Review

Medicinal orchids and their uses: Tissue culture a potential alternative for conservation

Bijaya Pant

Plant Biotechnology and Biochemistry Laboratory, Central Department of Botany, Tribhuvan University, Kathmandu, Nepal.

Accepted 19 August, 2013

Orchids are nature's most extravagant group of flowering plants distributed throughout the world from tropics to high alpine. They exhibit incredible range of diversity in size, shape and color of their flowers. Though orchids are grown primarily as ornamentals, many are used as herbal medicines, food, and other have cultural value by different cultures and tribes in different parts of the world. Orchids have been used in many parts of the world in traditional healing system as well as in the treatment of a number of diseases since the ancient time. Though Orchidaceae is regarded as a largest family of plant kingdom, few studies have been done regarding their medicinal properties. Linking of the indigenous knowledge of medicinal orchids to modern research activities provides a new reliable approach, for the discovery of novel drugs much more effectively than with random collection. Many of these orchids face the extreme danger of extinction due to over-exploitation and habitat loss. Plant tissue culture could be one of the most suitable alternative tools to minimize the pressure on natural population of medicinal orchids and their sustainable utilization.

Key words: Medicinal, orchids, propagation, conservation, culture.

INTRODUCTION

Orchids are nature's most extravagant group of flowering plants distributed throughout the world from tropics to high alpine (White and Sharma, 2000). They exhibit incredible range of diversity in shape, size and color of their flowers. They are important aesthetically, medicinally and also regarded as ecological indicators (Joshi et al., 2009). Several orchid species are cultivated for their various economic uses especially in floriculture. Orchids are grown primarily as ornamentals and are valued as cut flowers because of their exotic beauty and their long lasting blooming period (Hew et al., 1997). Though orchids are grown primarily as ornamentals, many are used as herbal medicines, food, and other cultural value by many different cultures and tribes in the different parts of world (Khasim and Rao, 1999; Kasulo et al., 2009). Though large population of orchid is still confined in their natural habitat, in many parts of the

world their number is decreasing due to their high demand and population pressure. Many orchid species are threatened due to their habitat destruction and indiscriminate collection.

At present, the orchids also figure prominently in the Red Data Book prepared by International Union for Conservation of Nature (IUCN). In fact, the entire family is now included in Appendix-II of Convention on International Trade in Endangered Species of Wild Fauna and Flora (CITES), where the international trade is strictly controlled and monitored.

Use of orchids in traditional medicine

Orchidaceae is regarded as the largest family of plant kingdom comprising 25,000-35,000 species (Dressler, 1993; Hossain, 2011). Very less study has been done

E-mail: pant_bijaya@yahoo.com or bijayapant@gmail.com.

regarding their medicinal properties. Limited information on medicinal values of orchids regarding their therapeutic properties in different parts of worlds is available and specifically corresponded to particular regions and community. Compilation of such information is very important to provide the reference for the drug development of many problematic diseases at present.

Scientists have traced orchids as far back as 120 million years ago. The history of orchids might started with their uses in the medicinal purpose. Chinese were the first to cultivate and describe orchids (Jalal et al., 2008). These plants first received recognition in the herbal writings of China and Japan 3,000 to 4,000 years ago, and they were the first to describe orchids for medicinal use (Reinikka, 1995; Bulpitt, 2005). Medicinal orchids belong mainly to genera: Anoctochilus, Bletilla, Cymbidium. Calanthe. Coelogyne, Cypipedium, Dendrobium, Ephemerantha, Eria, Galeola, Gastrodia, Gymnadenia, Habenaria, Ludisia, Luisia, Nevilia and Thunia (Szlachetko, 2001). Recently, more species belonging to different genera have been reported to have medicinal properties and in future more will be added in the list (Gutiérrez, 2010; Pant et al., 2011). We have listed ninety species of orchids from Nepal with their medicinal uses (Pant and Raskoti, 2013) (Table 1)

Dendrobium nobile, Bletilla striata and Gastrodia elata are routinely used in Traditional Chinese Medicine (TCM). Several species of Anoectochilus are used in Chinese folk medicines, such as Anoectochilus formosanus Hayata, Anoectochilus koshunensis Hayata, and Anoectochilus roxburghii (Wall.) Lindl. Anoectochilus roxburghii, which is distributed in southern China, Japan, Sri Lanka, India, and Nepal (Li and Zou, 1995), is also called "King Medicine" in China (Tseng et al., 2006). Different species of Dendrobium Sw are important in Chinese medicine used as Shi-hu (Shi hu: plant living on rocks) since the Han dynasty, 200 BC to 200 AD and are still used as a strengthening medicine and to cure varieties of diseases (Chen et al., 1994). Shi-hu is a term used to describe all Dendrobium and some Flickingeria species in China.

Different species of Dendrobium (Shi-hu) are used in Taiwan, Korea and Japan for various proposes such as a stomache, to treat night sweats, to fortify a person's body, to strengthen the kidneys and to cure impotence and as tonic. The tuber of Bletilla striata, which is called Baiji in China, has been used in traditional medicine to treat pneumonorrhagia and pneumono-phthisis. The medicine prepared from these tubers is used to treat tuberculosis, hemoptysis, gastrisis and duodenal ulcers, as well as bleeding, and cracked skin on the feet and hands. Other uses in China, Mongolia, Korea and Japan include the introduction of euphoria, purification of blood, strengthening and consolidation of lungs, as well as the treatment of pus, boils, abscesses, malignant swellings, ulcers and breast cancer (Zhang et al., 2006). Bulbophyllum kwangtungense Schlecht (Chinese name

"Shi dou-lan") has long been used in traditional Chinese medicine as a Yin tonic (Yi et al., 2005).

The earliest Middle East report of plant remedies is in a 4000-year-old Sumerian clay tablet which included some orchids (Kong et al., 2003). For flavoring, both Vanilla and Salep are well known and widely used long ago, the former is used as a delicious flavoring and wonderful perfume (Bechtel et al., 1992). Both are used in making ice-cream and beverages (Bulpitt, 2005).

Orchids are also one of the ingredients in ancient systems of medicine called Indian "Ayurveda". Asthavarga an important ingredient in many classical formulations viz., Chavyanprasa is reported to contain 4 species of orchids namely, Malaxis muscifrea, Malaxis acuminata, Habenaria intermedia, Habenaria edgeworthi (Singh and Duggal, 2009). Dendrobium macraei is another important orchid from Ayurvedic point of view as it is reported to be source of 'Jivanti'. Cypripedium parviflora is widely used as aphrodisiac and nervine tonic (Khasim and Rao, 1999). The tubers and pseudobulbs of several orchids like Orchis latifolia, Orchis mascula, Cymbidium aloifolium, Zeuxine strateumatica, and some species of Dendrobium, Eulophia and Habenaria are used as a restorative and in the treatment of various diseases (Puri. 1970).

Dendrobium fimbriatum, Papilionanthe teres, Eria musicucola, Eulophia compestris, Satyrium nepalense, Laparis odorata, Orchis latifolia, Vanda cristata, V. tessalata, V. coerula, V. spathulata, Cymbidium gigantieum, C. aloifolium, C. williomsoni, Dendrobium nobile, D. moschatum, Phaius tancarvilleae are some of the important medicinal plants used by traditional healer in Indian subcontinent (Suresh et al., 2000; Kong et al., 2003; Hossain et al., 2009; Medhi and Chakrabarti, 2009). Other Asian countries such as Indonesia, Malaysia, Taiwan, Singapore, Vietnam, Shri Lanka, Thailand, Myanmar, etc. have been using orchids in traditional medicine since the ancient time till date (Basu et al., 1971; Kumar et al., 2000; Hernández-Romero et al., 2005; Luo et al., 2007).

Similarly, use of orchids in America also has a long history. In Mexico, Vanilla has been used since ancient time to add aroma and flavor cocoa. In America, Vanilla planifolia was used as useful herb for the treatment of hysteria, fevers, impotence, rheumatism and to increase the energy of muscular systems since 15th century. Encyclia citrina, used by natives on infected wounds was described in the earliest literature. Laelia autumnal is for Stanhopea hernandezii for coughs; sunstroke; Arpophyllum spicatum, Bletia catenulate and Epidendrum pastoris for dysentery. Different species of Cypripedium were used in North America by different ethnic groups for its sedative and antispasmodic properties and to counter insomnia and nervous tension (Wilson, 2007). In North America, species collected for medicinal purposes include Cypripedium acaule, C. reginae, C. candidum and C. parvifolium (Cribb 1997; Richards, 1998; Duke,

Table 1. Uses of medicinal orchids in Nepal.

S/N	Botanical name	Habitat	Part used	Uses
1	Acampe papilliosa (Lindl.) Lindl.	Epiphytic	Root	Used to treat rheumatism (Ref. 1, 3, 5, 6)
2	Aerides multiflora Roxb.	Epiphytic	Leaves, Bulbs, Roots	Leaf paste applied to treat cuts and wounds. Plant parts possess antibacterial properties (Ref. 1, 5, 6)
3	Aerides odoratum Lour.	Epiphytic	Leaves	Leaf paste is used to treat cuts and wounds. Antibacterial properties (Ref. 1, 5, 6)
4	Anoectochilus roxburghii (Wall.) Lindl.	Terrestrial	Whole plant	Consumed to treat tuberculosis (Ref. 5)
5	<i>Arundina graminifolia</i> (D.Don) Hochr.	Terrestrial	Root	Root is used to relieve body ache (Ref. 4)
6	Brachycortis obcordata (Lindl.) Summerh.	Terrestrial	Root	Used in dysentery. Taken with milk as a tonic, nutritious (Ref. 1, 2, 5, 6)
7	Bulbophyllum careyanum (Hook.) Sprengel	Epiphytic	Leaves and pseudobulb	Fresh pulp of pseudo bulb is used in burns, powder of leaves is used to cause abortion and recovery during childbirth (Ref. 6)
8	Bulbophyllum leopardinum (Wall.) Lindl.	Epiphytic	Whole plant	Fresh pulp or juice is used in burns (Ref. 6)
9	Bulbophyllum odoratissimum (Sm.) Lindl.	Epiphytic	Whole plant	Used to treat tuberculosis and fracture (Ref. 6)
10	<i>Bulbophyllum umbellatum</i> Lindl.	Epiphytic	Whole plant	Used to enhance congenity (Ref. 2, 5)
11	Calanthe plantaginea Lindl.	Terrestrial	Rhizome	Dry powder with milk is taken as tonic and also as an aphrodisiac (Ref. 6)
12	Calanthe puberula Lindl.	Terrestrial	Rhizome	Dry powder with milk is taken as tonic (Ref. 6)
13	Calanthe sylvatica (Thou) Lindl.	Terrestrial	Flower	Juice is applied to stop nose bleeding (Ref.1, 6)
14	Calanthe tricarinata Lindl.	Terrestrial	Leaf, Pseudobulbs	Leaf paste applied on sores and eczema. Leaves and pseudobulbs are aphrodisiac (Ref. 1, 4)
15	Cephalanthera longifolia K. Fritsch		Rhizome	Appetizer, tonic, it heals wound (Ref. 1, 2)
16	Coelogyne corymbosa Lindl.	Epiphytic	Pseudobulbs	Juice of pseudobulbs applied in wound, paste applied in forehead to cure headache (Ref. 1, 2, 4, 5, 6)
17	Coelogyne cristata Lindl.	Epiphytic	Pseudobulbs	Pseudobulbs are given in constipation as also as an aphrodisiac. Juice of pseudobulbs is applied in wound and boils. Gum from psudobulb are used for sores (Ref. 3, 4, 5, 6)
18	Coelogyne flaccida Lindl.	Epiphytic	Pseudobulbs	Paste of pseudobulb is applied to forehead to cure headache and fever, juice is taken for indigestion (Ref. 4, 5, 6)
19	Coelogyne fuscescens Lindl.	Epiphytic	Pseudobulbs	Paste and juice for abdominal pain and in burns (Ref. 1, 5, 6)
20	Coelogyne nitida (Wall. ex Lindl) D. Don.	Epiphytic	Pseudobulbs	Paste and juice are applied in headache and fever and in burns (Ref. 6)
21	Coelogyne ovalis Lindl.	Epiphytic	Pseudobulbs	Aphrodisiac (Ref. 4, 5)
22	Coelogyne prolifera Lindl.	Epiphytic	Pseudobulbs	Paste is used to relieve from fever and headache and also applied in burns (Ref. 6), Paste is used for boils and backache (Ref. 4, 5)
23	Coelogyne stricta (D. Don) Schltr	Epiphytic	Pseudobulbs	Paste to relieve headache and fever (Ref. 1, 2, 5, 6)
24	Conchidium muscicola (Lindl.) Lindl.	Epiphyte	Whole plant	Used in cardiac, respiratory and nervous disorder (Ref. 2, 5)
25	Crepidium acuminatum (D. Don) Szlach	Epiphytic	Rhizome, root, psudobulb	Root powder is used for burns (Ref. 6), One of the ingredients of "Astavarga" of Ayurveda. Bulbs are used to treat bronchitis, fever, tuberculosis and weakness. Also given as a tonic (Ref. 1, 2, 5)

Table 1. Cont.

26	Cymbidium aloifolium (L.) Sw.	Epiphytic	Rhizome, root, bulbs	Paste is used for bone fracture and dislocated bones. Powder is used as a tonic (Ref. 1, 2, 4, 5, 6), Bulbs is used as demulcent agent (Ref.1,5)
27	Cymbidium devonianium Lindl. ex Paxton	Epiphytic	Whole plant	Root paste is applied to treat boils; concentrated decoction is taken in cough and cold (Ref. 4, 5)
28	Cymbidium elegans Lindl.	Epiphytic	Leaves, Pseudobulbs, roots	Fresh juice is coagulating, applied in deep wound to stop bleeding (Ref.1, 5, 6)
29	Cymbidium iridioides D. Don	Epiphytic	Leaves, Pseudobulbs, roots	Fresh juice is used to stop bleeding. Powder is used as tonic (Ref.1, 5, 6)
30	Cypripedium cordigerum D. Don	Terrestrial	Roots	Tonic, edible as a vegetable (Ref.1, 5)
31	Cypripedium elegans. Reichenb .f. Nep	Terrestrial	Roots	Nervine tonic in hysteria, spasm, madness, epilepsy and rheumatism (Ref.1, 5)
32	Cypripedium himalaicum (Rolfe) Kranzl	Terrestrial	Whole plant	Urine blocks treatment, Stone disease, heart disease, Chest disorder and cough (Ref. 5, 6)
33	Dactylorhiza hatagirea (D. Don) Soo	Terrestrial	Tubers	Tonic, wound healing and control bleeding, burns. Also used as a farinaceous food. Used to treat fever and various other body disorders (Ref.1, 2, 3, 4, 5, 6)
34	<i>Dendrobium amoenum</i> Wall. Ex Lindl.	Epiphytic	Pseudobulbs	Fresh paste is applied to cure burnt skin and dislocated bones (Ref. 6)
35	Dendrobium crepidatum Griff.	Epiphytic	Psudobulbs	Paste is used in fracture and dislocated bone. Ref. 8
36	Dendrobium densiflorum Lindl.	Epiphytic	Pseudobulbs	Pulps of the pseudobulbs are used in boils and pimples and other skin eruption (Ref. 4, 5, 6)
37	Dendrobium eriaeflorum Griff.	Epiphytic	Pseudobulbs	Paste is used to treat fractured and dislocated bones. Dried powder is used as tonic (Ref. 6)
38	Dendrobium fimbriatum Hook.	Epiphytic	Whole plant	Used in Liver upset and nervous debility (Ref.1, 5)
39	Dendrobium heterocarpum Wall.ex Lindl.	Epiphytic	Pseudobulb	Paste is used to treat fractured and dislocated bones (Ref. 6)
40	Dendrobium longicornu Lindl.	Epiphytic	Whole plant	Plant juice is used to relieve fever; boiled roots are used to feed livestock suffering from cough (Ref. 4, 6)
41	Dendrobium macaraei (Lindl.) Seidenf.		Whole plant	Paste is used against snake bite, general stimulant and demulcent (Ref. 6), Used in Asthma, Bronchitis, throat trouble, and fever, aphrodiasic (Ref. 5)
42	<i>Dendrobium monticola</i> P.F. Hunt & Summerh.	Epiphytic	Whole plant	Pulps of the pseudobulbs are used in boils and pimples and other skin eruptions (Ref.1, 2, 5)
43	Dendrobium moschtum Lindl.	Epiphytic	Pseudobulb	Paste is used to treat fractured and dislocated bones (Ref. 8)
44	Dendrobium nobile Lindl.	Epiphytic	Stem	Tonic useful in thirst and dryness of tongue. Given in weakening and fever (Ref. 1, 2, 5)
45	<i>Dendrobium transparens</i> Wall. ex Lindl.	Epiphytic	Pseudobulb	Paste is used to treat fractured and dislocated bones (Ref. 6)
46	Dienia cylindrostycha Lindl.	Terrestrial	Pseudobulb	Power is used as a tonic (Ref. 6)
47	Epipactis helleborine (L.) Crantz.	Terrestrial	Tubers	Used to treat insanity, gouts, headache & stomachache (Ref. 1, 2, 5, 6)
48	<i>Eria spicata</i> (D. Don) Hand. Mazz.	Epiphytic	Stem	Paste is taken internally to reduce stomachache and applied externally to reduce, headache (Ref. 1, 5, 6)
49	Eulophia dabia (D. Don) Hochr.	Terrestrial	Rhizome	Appetizer, tonic and aphrodisiac. Used in purulent cough and heart trouble. Tubers are given to infants in cough and cold (Ref. 1, 5, 6)
50	Eulophia nuda Landl.	Terrestrial	Tubers	Appetizer, useful for tuberculosis glands in neck, tumors and bronchitis (Ref. 5)

Table 1. Cont.

51	Flickingeria fugax (Rchb. f.) Seidenf.	Terrestrial	Whole plant	Powder is used as a tonic general debility stimulant (Ref. 6)
52	<i>Galeri</i> s <i>strachaeyi</i> (Hook. f.) P. F. Hunt	Epiphytic	Tubers	Used as tonic and to cure headache (Ref. 2, 5, 6)
53	Goodyera repens (L.) R. Br.	Terrestrial	Tuber	Plant paste externally applied in syphilis, extract is taken as a blood purifier (Ref. 7)
54	Gymnadenia orchidis Lindl.	Terrestrial	Roots, Pseudobulbs	Powered pseudobulbs are used to treat cuts and wounds. Also used for liver and urinary disorders and gastric (Ref. 1, 4, 5, 6)
55	Habenaria commelinifolia (Roxb.) Wall. ex Lindl.	Terrestrial	Whole plant	Used as Salep in combination of dried tubers of various orchids, and also used as spices (Ref. 5) The One of the ingredient of Astavarga of
56	Habenaria intermedia D. Don.	Terrestrial	Tubers	Ayurveda, used as tonic. Tuber paste is used to cure various diseases such as hyperdipsia, fever cough, asthma leprosy skin diseases (Ref. 2, 5, 6)
57	Habenaria marginata Colebr.	Terrestrial	Tubers	Thoroughly boiled plant extract taken in flatulence. in wound, tonic (Ref. 7).
58	<i>Habenaria pectinata</i> (Sm.) D. Don	Terrestrial	Tubers	Leaf juice applied in snake bites. Tuber used against arthritis (Ref. 6)
59	Herminium lanceum (Thunb. ex Sw.) Vuijk	Terrestrial	Whole plant	Extract of plant given in suppressed urination (Ref. 7).
60	Herminium monorchis (Linn.) R .Br.	Terrestrial	Roots	Tonic (Ref. 7).
61	Liparis nervosa (Thunb) Lindl.	Terrestrial	Tubers	Used to treat stomachache, malignant ulcers (Ref. 2, 5)
62	Luisia trichorhiza (Hook.) Bl.	Epiphytic	Tubers	Paste is applied externally to cure muscular pain (Ref. 2, 5, 6)
63	Luisia zeylanica Lindl.	Epiphytic	Leaves	Juice is used to treat chronic wounds, boils and burns (Ref. 2, 4, 5, 6)
64	<i>Malaxi</i> s <i>muscifera</i> (Lindl.) Kuntze	Terrestrial	Swollen stem base	Useful in sterility, seminal weakness, dysentery, fever and general debility as a tonic (Ref. 5, 6)
65	Neottianthe calcicola (W.W. Sm.) Soo.	Terrestrial	Rhizome	Tonic (Personal communication)
66	Nervilia aragoana Gaudich.	Terrestrial	Whole plant	Used in uropathy, haemoptysis cough asthma, vomiting, diarrhoea & mental instability (Ref. 1, 5)
67	Oberonia caulescens Lindl.	Epiphytic	Tubers	Used in liver ailments (Ref. 1, 5)
68	Otochilus albus Lindl.	Epiphytic	Whole plant	Powder is used as a tonic (Ref. 6)
69	Otochilus lancifolius Griff.	Epiphytic	Pseudobulb	Used to treat fractured and dislocated bones (Ref. 6)
70	Otochilus porrectus Lindl.	Epiphytic	Whole plant	Used as tonic & also in the treatment of sinusitis rheumatism (Ref. 3, 5)
71	Papilionanthe teres (Roxb.) Schltr.	Epiphytic	Whole plant	Paste is applied to treat dislocated bones (Ref. 4, 5, 6)
72	Phaius tankervilliae (Banks) Blume.	Terrestrial	Tubers	Tonic (Personal communication)
73	Pholidota articulata Lindl.	Epiphytic	Roots, fruits	Whole plant used as tonic. Root powder is used to treat cancer, juice berries is used to treat skin ulcers and skin eruptions (Ref. 1, 5, 6)
74	Pholidota articulata Lindl. var. grifithii Hook. f.	Epiphytic	Pseudobulb	Paste is applied to treat dislocated bones (Ref. 4, 5)
75	Pholidota imbricata (Roxb.) Lindl.	Epiphytic	Bulbs, Pseudobulb	Juice is applied to relieve naval pain, abdominal pain, and rheumatic pain. Also as a tonic (Ref. 1, 2, 5)
76	Pholidota pallida Lindl.	Epiphytic	Roots, Pseudobulb	Juice is applied to relieve naval pain, abdominal pain, and rheumatic pain. Powder is used to induce sleep (Ref. 1, 2, 5, 6)

Table 1. Cont.

	Platanthera edgeworthii			
77	(Hook. f. ex Collett) R. K. Gupta.	Terrestrial	Root, Leaves	Powder is used as a blood purifier (Ref. 6)
78	Platanthera sikkimensis (Hook. f.) Kraenzlin.	Terrestrial	Bulbs, Pseudobulb	Juice is applied to relieve naval pain, abdominal pain, and rheumatic pain (Ref. 1, 2, 5)
79	Pleione humilis (Sm.) D. Don	Epiphytic	Pseudobulb	Dried powder is tonic; Paste is used in cut and wounds (Ref. 4, 5, 6)
80	Pleione maculata (Lindl.) Lindl.	Epiphytic	Rhizome	Used for liver and stomach ailments (Ref. 1, 2, 5)
81	Pleione praecox (Sm.) D. Don	Epiphytic	Pseudobulb	Dried powder is tonic; paste is used in cut and wounds (Ref. 4, 5, 6)
82	Rhynchostylis retusa (L.). Bl.	Epiphytic	Whole plant	Leaves are used to treat rheumatism Root juice is applied to cuts and wounds (Ref. 2, 3, 4, 6)
83	Satyrium nepalense D. Don.	Terrestrial	Tubers	As a tonic and also used in diarrhea and malaria, tubers edible, juice is used externally in cut and wounds (Ref. 1, 2, 4, 5, 6)
84	<i>Smitinandia micrantha</i> (Lindl.) Holttum	Epiphytic	Whole plant	Root powder as a tonic and stem has antibacterial property (Ref. 1, 5)
85	Spiranthes sinensis (Pers.) Ames	Terrestrial	Tuber	Decoction of plant given in intermittent fever, tubers used as tonic. Paste of roots and stem is applied in sores (Ref. 1, 5, 6)
86	Thunia alba (Lindl.) Rchb. F.	Epiphytic	Whole plant	Plant paste is applied to treat dislocated bones (Ref. 4, 5, 6)
87	<i>Trudelia cristata</i> (Lindl.) Senghas	Epiphytic	Roots & Leaves	Root paste is applied in cuts, wounds, boils and dislocated bones (Ref. 3, 4, 5)
88	Vanda tessellata (Roxb.) Rchb. f.	Epiphytic	Roots, leaves	Used in rheumatism and allied disorders, paste of leaves is used for fever (Ref. Subramoniam and Pushpangadan, 2000). Used in rheumatism and allied disorders (Ref. 2, 5).
89	Vanda testacea (Lindl.) Rchb.f.	Epiphytic	Leaves	Used as antiviral and anticancer agent. Leaf drops are used for earache (Ref. 1, 2, 5, 6)
90	Zeuxine strateumatica (L.) Schltr.	Terrestrial	Roots and tubers	Dry powder is used as tonic (Ref. 6)

References: Vaidhya et al., 2000, (1); Shrestha, 2000, (2); Joshi and Joshi, 2000, (3); Manandhar, 2002, (4); Baral and Kurmi, 2006, (5); Subedi 2011, (6); Joshi et al., 2009 (7). Source: Medicinal orchids of Nepal, Pant and Raskoti, 2013.

2002; Moerman, 1998). Several species of *Goodyera* have been used as herbal remedy by the natives in North America. *Goodyera pubescens*, commonly known as 'Downy Rattlesnake Orchid', were used for infallible cure of the bite of a mad dog and to cure scrofula (Moerman, 1986).

The history of use of orchid in Europe is very long and is being used even today in various preparations. Langham, 1579, in his Garden of Health, he reported antipyretic, anti-consumption and anti-diarrhoeal effects of many European terrestrial orchids. The number of orchid species recorded as medicinal throughout Europe such as Ophrys apifera, O. muscifera, O. fuciflora, O. sphegodes, Orchis simia, O. mascula, Himantoglossum hircinum, Serapias vomeracea, S. lingua, Dactylorhiza majalis, D. majalis, foliosa etc. were used as aphrodisiac and have other healing properties (Turner, 1568). In Europe, some species of Epipactis have been used in traditional medicine preparation. The roots of Epipactis

gigantea, commonly known as 'Giant Orchid', have been used in a severe case of illness as a tonic. *Epipactis helleborine* was valued as a remedy for gout in European folklore. Its rhizome is also used as aphrodisiac infusion or decoction (Balzarini et al., 1992). The roots of *E. latifolia* were used in rheumatism. Several species of *Spiranthes* have also been used medicinally in various diseases, for instance *Spiranthes diuretica* is effective as a diuretic in children, *Spiranthes autumnalis*'s roots are used as a strong aphrodisiac as reported by Balzarini.

The early settlers and Australian aborigines in Australia used orchids in the earliest time (Lawler and Slaytor, 1970). Bulbsofmanyorchidssuchas *Gastrodia sesamoides*, *Dendrobium speciosum* and *Caladenia* species were used as emergency food (Bulpitt, 2005). The infusion or decoctions from the leaves of *Dendrobium aurantiacum* were used to cure diabetes (Yang et al., 2005). *Selenipedium chica*, considered as the tallest plants of the orchid family, was used occasionally as a substitute for vanilla.

Pseudobulbs of *Cymbidium madidam* were chewed for dysentery and its seeds were used as an oral contraceptive. In addition, *Cymbidium canaliculatum, Dendrobium teratifolium* and *Dendrobium discolor* were used for treating different ailments such as dysentery, to relieve pain and control ringworm (Lawler and Slaytor, 1970).

In Africa, the Zulus used several orchids for therapeutic purposes. Several species of Eulophia were used to prevent miscarriage and cure barrenness. Powdered form of Eulophia flaccida were applied to incisions made on the skin to relieve pain. Eulophia aha, commonly known as 'Wild cocow', was introduced in South Africa in the early days of the slave trade for its various medicinal uses. The Zulus also used the stems of Ansellia gigantea for their aphrodisiac intent. Morris (2003) has described twelve orchids currently used as medicine in Malawi. Nine of these are used for stomach complaints and two for fertility problems. Cyrtorchis arcuata and Eulophia cucullata are used to treat diabetes or skin infections and Eulophia cucullata to prevent epilepsy. An infusion of the leaves and pseudobulbs of Bulbophyllum maximum is used to protect against sorcery, and to treat madness. In Zambia and East sub-Saharan Africa, the boiled root tubers of some terrestrial orchid are used to make a food dish (Davenport, 2004). In Africa, an amulet of leaves of Ansellia africana infused with a paste made from the pseudobulbs of the same species is said to function as a short term contraceptive (Berliocchi, 2004). Stems infusion or decoction of Galeola foliate is used for the treatment of some infections in Morobe, Papua New Guinea (Khan and Omoloso, 2004).

Orchids as a rich source of natural compounds and their pharmacological uses

Researchers have found the various activities of metabolites and extracts of different orchid species in the treatment of various diseases. They have been used variously in different diseases as anti rheumatic, antiinflammatory, antiviral, anti carcinogenic, anticonvulsive, diuretic, neuroprotective, relaxation, anti-aging, wound healing, hypoglycemic, antitumor and anticancer, antimicrobial, antiviral and many other activities (Ghanaksh and Kaushik, 1999; Shyur et al., 2004; Li et al., 2001; 2006; Shimura et al., 2007; Wang et al., 2006; Prasad and Achari, 1966; Kumar et al., 2000; Zhao et al., 2003; Satish et al., 2003; Watanabe et al., 2007, Won et al., 2006; Lawler and Slaytor, 1970; Balzarini et al., 1992; Nayak et al., 2005; Miyazawa et al., 1999). For drugs derived from orchids, some novel discoveries, both in phytochemical and pharmacological properties, were reported by some researcher. Studies have reported the isolation of wide range of important phytochemicals from different genera of orchids such as alkaloids, flavonoids, stilebnoids, anthocyanins, triterpedoids, orchinol, hircinol,

cypripedin, bibenzyl derivatives, phenanthrenes, jibantine, nidemin and loroglossin which are present in leaves, pseudobulb, roots, flowers or in the entire plant (Okamoto et al., 1966; Williams, 1979; Majumder and Sen, 1991; Majumder et al., 1996; Zhao et al., 2003; Yang et al., 2006; Singh and Duggal, 2009).

Thus, from various studies, it is well known that orchids have been used all over the world in traditional healing and treatment system of a number of diseases. Knowledge of different ethnopharmacological studies, linking of the indigenous knowledge of medicinal orchids to modern research activities provides a new reliable approach, which makes the chances of discovery of drugs much more effective than with random collection. In this perspective, orchids which have been used for centuries are the potential resources for many novel drugs. It can be predicted that more genera and species of orchid possesses the possibility of having medicinal properties and in future they can be utilized for the ever demanding life saving drugs (Figures 1a to d).

Threats to orchids

Globally, orchids are the most threatened species among the flowering plants. Due the various reasons like overexploitation, illegal trade and encroachment of land, change in climate, orchids species are threatened rampantly (Shrestha, 2000; Pant et al., 2007). Medicinal orchids are under considerable threat due to habitat destruction, degradation - fragmentation and illegal collection for trade and consumption (Pant et al., 2002). Most of these species has been categorized as critically endangered, rare and listed under appendix II of CITES (IUCN status). There are some species such as Liparis olivacea, which have already extinct from the wild (Subedi, 2011). There is a wide gap between the supply and demands of medicinal orchids. Collection and sale of wild orchid from the orchid rich area specially by the rural community is the routine activity, uprooting the whole plant causes the extinction of many species and providing the huge amount of such orchids to the local and international traders (Kala, 2004). Due to such various levels of disturbances, destruction of number of economically important plants in alpine meadows has continued like reduction of Dactylorhiza hatagirea, a high valued medicinal orchids of the region from its natural population in the Himalayas which has been categorized as critically endangered listed under appendix I of CITES (Badola and Aitken, 2003; Giri et al., 2008). Due to the recent trend of using traditional medicine in western countries its demand is increasing. Such rapid depletion from the wild requires urgent conservation measures.

CONSERVATION MEASURES

As many valuable orchids are now at the verge of



Figure 1a. Some medicinal orchids of Nepal; Cymbidium devonianium.

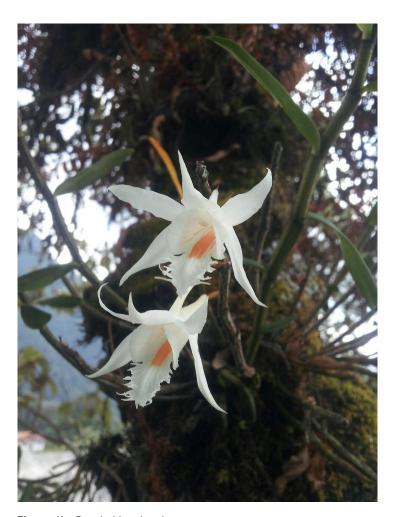


Figure 1b. Dendrobium longicornu.



Figure 1c. Dendrobium fimbriatum.



Figure 1d. Pholidota articulate.

extinction, so it is high time to conduct effective strategies to conserve them throughout all geographical regions. Conservation of orchids is an important issue that should be seriously considered by both government and private sector of the concern nation in participation with research institutions, non-government organization, community growers as well as through international collaboration. Conservation of medicinal orchids can be addressed by both *in situ* and *ex situ* measures in association with participation of local people.

In situ conservation

In situ conservation, the conservation of species in their natural habitats, is considered the most appropriate way of conserving biodiversity. Habitat protection could be the most important in situ conservation strategies for orchids. Because of their small population size and restricted distribution, intensive care and habitat management is highly recommended for their in situ conservation. Thus the Protected Areas (PAs) form a central element of any national strategy to conserve biodiversity. However, illegal collections of species from their natural habitat continue even from the PAs in many part of the world due to poor enforcement and regulation of law (Chaudhary et al., 2002; Dixon et al., 2003). Moreover, in situ conservation is not always a viable option because of the modification of habitat and migration or absence of the pollinators due to unfavorable modifications environment (Swarts, 2007; Swarts and Dixon, 2009). There is no substitution for conservation of threatened medicinal orchid species in their natural habitat by natural propagation method as their propagation rate is very slow.

Ex-situ conservation

Ex situ conservation is the preservation of components of biological diversity outside their natural habitats. Ex situ conservation measures can be complementary to in situ methods as they provide an "insurance policy" against extinction. These measures also have a valuable role to play in recovery programmes for endangered species. In this context, ex situ conservation is very important aspect of orchid conservation which can include both seed banks and in vitro culture plant tissue collections. Therefore there is an urgent need to develop such conservatory for long term conservation and recovery programme specifically for medicinal orchids in the threatened area.

Domestication and participation of community based organization

Propagation techniques for medicinal orchids are yet to

be perfected and encouraged to sell the cultivated orchid for the income generations of the people which will promote both *ex situ* and *in situ* conservation. In this regards, participation of community based organization, community forest users' groups (CFUs), private nurseries and orchid enthusiastic is very important. Cultivation of medicinal orchids could be one of the effective ways of income generation of ethnic inhabitants worldwide who are using medicinal orchids for their primary health care and trade to support their livelihood. This will be one of the best alternatives for the more sustainable use of wild orchids (Pant et al., 2007).

Propagation of orchids

Orchids are propagated either sexually or asexually. Propagation of orchids through sexual means is a very slow process as their seeds lack endosperm and need fungal stimulant for germination in nature. Since most of the commercial orchids are highly heterozygous they are not raised through seeds and are propagated through vegetative means to get true-to-type plants. Conventional methods such as cuttings division of shoots are applied for the vegetative propagation of orchids. Orchids are highly heterozygous and their vegetative propagation through division of clumps of rhizomes, bulbs or by the rooting of off shoots also takes long time and difficult to obtain desired number of orchids. This difficulty in natural population drives the many orchids including medicinal orchids to be threatened and some are reached to extinction. It is therefore important to take initiative for their mass propagation and reestablish them in nature.

Propagation through seeds: Symbiotic seed germination

The physiology of its seed germination has made the family Orchidaceae most interesting as their seeds are unique and adaptive in several respects. The small, dust like seeds of the orchids produced in each capsule, are highly fragile, nearly microscopic in size and are produced in very large numbers (Mitra, 1971). As many as 1,300 to 4,00,000 seeds per capsules are produced(Figures 2 and 3). Their color may be white, cream, pale green, reddish orange or dark brown and have very diverse shapes. Orchid seeds are characterized by lack of storage tissues required for seed germination and seedling development.

Orchid requires a combination of multiple factor for their continued reproduction in nature. In nature, association with a specific fungal partner, the orchid mycorrhiza is a pre-requisite for orchid seed germination (Mitra, 1986). Most of the mycorrhizal fungi of orchids fall into a non-sporing group known as Rhizoctonia, the major species



Figure 2. Immature capsules of orchids in their natural habitat.

being Rhizoctonia repens, Rhizoctonia mucoroides and Rhizoctonia languinosa. Orchid seeds cannot utilize their own reserve or do so very slowly, they also cannot hydrolyse large molecules like starch or cellulose. As a result, asymbiotic germination in the absence of sugar proceeds only to the early protocorm stage, after which they wait for external supply of simple sugars through the help of mycorrhizal fungus. The fungus is believed to augment the carbohydrate, auxin and vitamin transport in the orchid which is called symbiotic germination (Arditti et al., 1992). Symbiotic associations between orchids and mycorrhizal fungi are a competitive struggle. The fungi always try to invade the cytoplasm of orchid cells to obtain nutritional compounds. On the other hand, the orchid cells restrict the growth of the infecting hyphae and obtain nutrition by digesting them. It is assumed that antifungal compounds are involved in the restriction of fungal growth inside the orchid (Shimura et al., 2007). The rate of seed germination in nature is very poor, that is, 2-5% (Rao, 1977; Vij, 2002). Even in the symbiotic germination, the seeds take a long time for their

germination and any disturbance in the habitat or physical environment destroys the whole population.

Plant tissue culture, a breakthrough for the orchid propagation

Plant tissue culture technique has been accepted as a potential alternative method for mass scale propagation and conservation of rare, threatened and endangered orchids. Invention of *in vitro* propagation technique has saved the many naturally growing orchids and their collection from the wild has reduced. Increasing popularity of orchids for cut flower and medicinal purpose has added new dimension to *in vitro* propagation technique through which a significant number of identical clones can be raised from a single protocorm or shoot tip explants (Arditti and Ernst, 1993; Deb and Pongener, 2012). Thus methods for rapid multiplication of orchids are essential to meet the commercial demand. Various works on *in vitro* culture include propagation from seed or

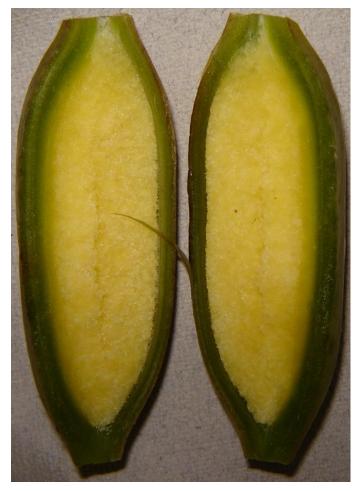


Figure 3. LS of orchid capsule showing immature seeds.

different explants such as shoot tips, stems, rhizomes protocorms, etc. More information on *in vitro* culture of orchids is described under separate heading below.

Asymbiotic seed germination in synthetic nutrient media

The propagation and cultivation of orchid was revolutionized after the discovery of Knudson (1922), media. The asymbiotic method for orchid seed germination developed by Knudson (1884-1958) was the first procedure for *in vitro* propagation of any plant in pure culture (Yam et al., 2009). Lewis Knudson in 1916, while working on the influence of the carbohydrates on green plants, also started experimenting with the germination of orchid seeds on the basis of the analysis of orchid salep which contained starch, protein, sugars and minerals. He formulated a medium and successfully germinated seeds of *Cattleya*, *Laelia*, *Epidendrum* and concluded that fungus was not necessary for orchid seed germination. Development of asymbiotic germination methods of orchid seeds took place following the formulation of

Knudson B and C medium (Knudson, 1922, 1946). After his demonstration of the possibility of by-passing the fungal requirement for germination of orchid seeds during *in vitro* culture, non-symbiotic seed germination has been accepted as an important tool for propagating orchids (Ernst, 1982). This method was a major methodological, biological and technological advance which contributed to modern biotechnology (Johnson et al., 2007).

Different media used for Orchid culture

Successes of plant tissue culture primarily depend on the formulation of nutrient media. With the advent of Knudson medium, a large number of modified media have been standardized and became available in market even with pre-adjusted pH. The most common media used for orchid culture are Knudson C medium (Knudson, 1922, 1925, 1927, 1946); MS medium (Murashige and Skoog, 1962); Vacin and Went medium (Vacin and Went, 1949); and many others (Arditti, 1968; Ernst, 1974; Mitra, 1986; Jonn, 1988). Different media range from simple three-salt solution to complex containing 20 or more salts of macro



Figure 4. Germinating seeds of Cymbidium aloifolium in MS medium after 21 weeks of culture.

and micro elements (Chang and Chang 2000a). Some media are designed for specific genera, while others for a broad spectrum. Plant growth regulators such as auxins and cytokinins are added in the media to enhance the seed germination in some orchids (Pant and Gurung, 2005; Stewart and Kane, 2006; Deb and Temjensangba, 2006; Johnson et al., 2007; Hossain, 2008; Pradhan and Pant, 2009; Pant et al., 2011). A large number of complex additives like coconut water, banana pulp, peptone, tomato juice, salep, honey and beef extract have been used in different media to show their effect on orchid seed germination (Mitra, 1971; Vij, 1993; Hua and Zhiguo, 1998; Mohammad et al., 2009). The germination rate of seeds, protocorm formation and complete development of seedlings depend on the genotype, maturity of seeds and culture condition.

Importance of seed culture in orchids

In vitro germination of seeds is an important aspect in the orchid multiplication and conservation program since the dust like tiny seeds have the capability of developing into complete seedlings without any fungal aid, Arditti (1967). After the successful pioneer work of Knudson, commercial propagation of orchids advanced remarkably. The hybridizing potential found in orchids, both at the

inter-specific and inter-generic level, has been utilized by amateur and commercial orchid growers to produce thousands of artificial hybrids. The desire to observe the resulting hybrids quickly leads to research and advances in *in vitro* culture techniques which have led to many innovations. Production of artificial seeds and propagation through them is another potential area of orchid conservation and propagation (Rederbangh et al. 1993; Fujii et al., 1989; Datta et al., 1999) (Figures 4, 5 and 6 a,b).

Propagation of orchids using different explants

Micropropagation, monoculture of desirable clones under uniform conditions and stopping the use of plants collected from the wild may solve problem of loss of orchid gene pool and help to conserve the gene bank of medicinal orchids. *In vitro* propagation using seed culture is less desirable in many cases especially for horticultural uses due to the long juvenile period before flowering (Decruse et al., 2003). As orchids are outbreeders, their propagation using seeds leads to the production of heterozygous plants. To obtain similar clones from the superior mother plants, regeneration from various vegetative parts of mature plants are essential. After the development of protocol for *in vitro* micropropagation by

461



Figure 5. Development of protocorms from germinating seeds of *Cymbidium iridioides*.



Figure 6a. In vitro propagation of Cymbidium aloifolium through artificial seeds.



Figure 6b. In vitro propagation of Cymbidium aloifolium through artificial seeds.

Morel (1960) who cultured shoot tips for the production of large number of virus free *Cymbidiums*, commercial orchids were predominantly produced by tissue culture and this technique is routinely used worldwide for mass scale production of orchids by the orchid growers (Wimber, 1963). During the last 50 years, tissue culture techniques using different explants have been exten-

sively exploited, not only for the rapid and large-scale propagation of orchids but also for their ex situ conservation. Different protocols have been developed for the large-scale propagation of a number of orchid species and hybrids through *in vitro* culture of various parts including shoot tips, flower stalk, nodes, buds, stems, root tips and rhizome segments (Vij, 1993; Nayak



Figure 7. In vitro propagation of Dendrobium densiflorum from root culture.

et al., 1998; Kanjilal et al., 1999; Chang and Chang, 2000b; Chen et al., 2003b; Chugh et al., 2009; Pant and Shrestha, 2011; Deb and Pongener, 2012; Paudel and Pant, 2012). Mass propagation of medicinal orchids using in vitro culture technique has been reported by some workers (Sharma and Chandel, 1996; Liu and Zhang, 1998); Nalawade et al., 2003; Shiau et al., 2005; Basker and Bai 2006; Sharma et al., 2007; Pant et al., 2008; Hossain et al., 2009, 2012; Kaur and Bhutani, 2009, 2010; Nongdam and Chongtham, 2011; Pant and Thapa, 2012; Pradhan et al., 2013). There are very few reports on reintroduction of in vitro propagated species of medicinal orchids to natural habitat (Stewart and Kane, 2006; Aggarwal and Zettler, 2010; Lesar, 2012.) or their cultivation, which are always collected from the wild for trade. To substitute for the habitat protection and species recovery, it is very important to reintroduce over exploited species (Figures 7, 8 and 9). At the mean time, there are ample opportunity and possibility of production of desired phytochemicals in culture (Pant, 2008; Mazumder et al. 2010).

CONCLUSION

Extensive research is still necessary to be able to fullyrecommend the orchid species for their medicinal uses. Due to their small population size and restricted distribution, intensive care and habitat management is highly recommended. Very little effort has been made to cultivate the medicinal orchids for commercial scale. The species which has reached the threatened category because of the human activities can survive only with human support. Plant tissue culture could be one of the most suitable alternative tools to minimize the pressure on natural population of medicinal orchids and their sustainable utilization.

ACKNOWLEDGEMENTS

The author gratefully acknowledges Ms Shreeti Pradhan, Ms Tripti Regmi, Mr. Babulal TIruwa and Mr. Bishnu Joshi for their cooperation in the research and Dr. Basant



Figure 8. *In vitro* grown plantlets of *Phaius tancarvilleae* form shoot tip culture.



Figure 9. Acclimatization of in vitro grown plantlets of Cymbidium aloifolium.

Pant for the photography of some medicinal orchids. All the tissue culture photographs are used from our research work. The author gratefully acknowledges all the team members involved in orchids research project.

REFERENCES

- Aggarwal S, LW Zettler (2010). Reintroduction of an endangered terrestrial orchid, *Dactylorhiza hatagirea* (D. Don) Soo, assisted by symbiotic seed germination: First report from the Indian subcontinent. Nat. Sci. 8(10):139-145.
- Arditti J (1967). Factors affecting the germination of orchid seeds. Bot. Rev. 33:1-97.
- Arditti J (1968). Germination and growth of orchids on banana fruit tissue and some of its extracts. Am. Orchid Soc. Bull. 37:112-116.
- Arditti J (1992). Fundamentals of Orchid Biology. John Wiley & Sons, New York.
- Arditti J, Clementsm MA, Fast G, Hadley G and Nishimura, G (1982). Orchid seed germination and seedling culture-A manual. In: Orchid Biology-Reviews and perspectives, Vol II, Arditti J (Ed.), Cornell University Press, Ithaca, New York. pp. 243-370.
- Arditti J, Ernst R (1984). Physiology of orchid seed germination. In: Arditti J (ed) Orchid biology: reviews and perspectives, New York.
- Badola HK, Pal M (2002). Endangered Medicinal plant in Himachal Pradesh. Curr. Sci. 83:797-798.
- Balzarini J, Neyts J, Schols D, Hosoya M, Van Damme E, Peumans W, De Clercq E (1992). The mannose-specific plant lectins from *Cymbidium hybrid* and *Epipactis helleborine* and the (Nacetylglucosamine) n-specific plant lectin from *Urtica dioica* are potent and selective inhibitors of human immunodeficiency virus and cytomegalovirus replication *in vitro*. Antiviral Res. 18:191-207.
- Baral SR, Kurmi PP (2006). A Compendium of Medicinal Plants of Nepal. Publisher Rachana Baral, Printed in Nepal by Mass Printing Press, Kathmandu.
- Basker S, Bai VN (2006). Micropropagation of coelogyne stricta (D. Don) Schltr. Via pseudo bulb segment cultures. Trop. Subtrop. Agroecosyst. 6:31-35.
- Basu K, Dasgupta B, Bhattacharya S, Lal R, Das P (1971). Anti-inflammatory principles of *Vanda roxburghii*. Curr. Sci. 40:80-86.
- Bechtel H, Cribb P, Launert E (1992). The manual of cultivated orchid species. 3rd ed. Blandford Press, London.
- Berliocchi L (2004). In: Griffiths M, editor. The orchid in lore and legend. Portland, Oregon: Timber Press.
- Bulpitt C (2005). The uses and misuses of orchids in medicine. QJM 98:625-631.
- Chang C, Chang WC (2000). Micropropagation of *Cymbidium ensifolium var.* misericors through callus-derived rhizomes. *In vitro* Cell. Dev. Bio. Plant. 36:517-520.
- Chang, C, Chang, WC (2000). Effect of thidiazuron on bud development of *Cymbidium sinense* Willd *in vitro*. Plant Growth Regul. 30:171-175.
- Chaudhary R, Subedi A, Shakya L, Karki D, Vetass O, Gupta V (2002) .Orchid diversity in Arun river and Marsyagdi river basins of Nepal: distribution and conservation priorities. Vegetation and society: their interaction in the Himalayas pp.108-117.
- Chen CC, Wu LG, Ko FN, Teng CM (1994). Antiplatelet aggregation principles of *Dendrobium loddigesii*. J. Nat. Prod. 57:1271-1274.
- Chen JT, Chang WC (2003). 1-Aminocyclopropane-1-carboxylic acid enhanced direct somatic embryogenesis from *Oncidium* leaf cultures. Biol. Plant 46:455-458.
- Chugh S, Guha S, Rao IU (2009). Micropropagation of orchids: a review on the potential of different explants. Scientia Horticulture 122:507-520.
- Cribb P (1997. The genus *Cypripedium*. Portland, Oregon: Timber Press; p.301.
- Datta K, Kanjilal B, De Sarker D (1999). Artificial seed technology: Development of a protocol in *Geodorum densiflorum* (Lam) Schltr.-An endangered orchid. Curr. Sci. 76:1142-1144.
- Deb CR, Temjensangba S (2006). Effect of different factors on nonsymbiotic seed germination, formation of protocorm-like bodies and plantlet morphology of *Cleisostoma racemiferum* (Lindl.) Garay. Indian J. Biotechnol. 5:223.

- Deb CR, Pongener A (2012). Studies on the *in vitro* regenerative competence of aerial roots of two horticultural important *Cymbidium* species. J. Plant Biochem. Biotechnol 6:1-7.
- Dixon KW (2003). Orchid conservation. Natural History Publications (Borneo).
- Dressler RL (1993). Phylogeny and classification of the orchid family: Cambridge University Press.
- Duke JA, Bogenschutz-Godwin M, Cellier J (2002). Handbook of medicinal herbs: Boca Raton, Florida CRC.
- Ernst R (1982). Orchid seed germination and seedling culture-a manual: *Paphiopedilum*. Orchid Biol. Rev. Perspect. 2:350-353.
- Fujii J, Slade A, Redenbaugh K (1989). Maturation and greenhouse planting of alfalfa artificial seeds. In vitro Cell Dev. Biol. 25:1179.
- Ghanaksh A, Kaushik P (1999). Antibacterial effect of *Aerides multiflora* Roxb.: a study *in vitro*. J. Orchid Soc. India 13:65-68.
- Giri D, Arya D, Tamta S, Tewari LM (2008). Dwindling of an endangered orchid *Dactylorhiza hatagirea* (D. Don) Soo: A case study from Tungnath Alpine meadows of Garhwal Himalaya. India. Natl. Sci. 6:6-9.
- Gutiérrez RMP (2010). Orchids: A review of uses in traditional medicine, its phytochemistry and pharmacology. J. Med. Plants. Res. 4:592-638.
- Hernández-Romero Y, Acevedo L, Sánchez MLÁ, Shier WT, Abbas HK, Mata R (2005). Phytotoxic activity of bibenzyl derivatives from the orchid *Epidendrum rigidum*. J. Agric. Food. Chem. 53:6276-6280.
- Hew CS, Arditti J, Lin WS (1997). Orchid cut-flower production in ASEAN countries. In: Arditti, J(Ed.), Orchid Biol. Rev. Perspect. 6:363-401.
- Hossain MM (2011). Therapeutic orchids: traditional uses and recent advances-An overview. Fitoterapia 82:102-140.
- Hossain MM, Sharma M, Pathak P (2009). Cost effective protocol for *in vitro* mass propagation of *Cymbidium aloifolium* (L.) Sw.-a medicinally important orchid. Eng. Life. Sci. 9:444-453.
- Hossain MM, Sharma M, Pathak P (2012). *In vitro* propagation of *Dendrobium aphyllum* (Orchidaceae)-seed germination to flowering. J. Plant Biochem. Biotechnol. pp.1-6.
- Hua L, Zhiguo Z (1998). Studies on plantlet strengthening medium for Dendrobium candidum Wall. et Linde. of clonal propagation in vitro. China J. Chin. Mater. Med. 11:654-655
- Jalal JS, Kumar P, Pangtey Y (2008). Ethnomedicinal Orchids of Uttarakhand, Western Himalaya. Ethnobotanical Leaflets 2008:164.
- Johnson TR, Stewart SL, Dutra D, Kane ME, Richardson L (2007). Asymbiotic and symbiotic seed germination of *Eulophia alta* (Orchidaceae)-preliminary evidence for the symbiotic culture advantage. Plant Cell Tiss. Org. Cult. 90:313-323.
- Jonn SD (1988). Simple medium for regeneration of Pclb's of Debdrobium walter Oumae. Bull. Penelitian Hort. 16:73-78.
- Joshi KK, Joshi SD (2000). Genetic Heritage of Medicinal and Aromatic Plants of Nepal Himalayas. Buddha Academy Publisher and Distributors, Pvt. Kathmandu, Nepal.
- Joshi G, Tewari LM, Lohani N, Upreti K, Jalal JS, Tewari G (2009). Diversity of orchids on Uttarakhand and their conservation strategy with special reference to their medicinal importance. Rep. Opin. 1:47-52.
- Kala C (2004). Assessment of species rarity. Curr. Sci. 86:1058-1058.
- Kanjilal B, Sarker DDE, Mitra J, Datta KB (1999). Stem disc culture: Development of a rapid mass propagation method for *Dendrobium moschatum* (Buch.-Ham.) Swartz- An endangered orchid. Curr. Sci. 77:497-500.
- Kasulo V, Mwabumba L, Munthali C (2009). A review of edible orchids in Malawi. J. Hortic. For. 1:133-139.
- Kaur S, Bhutani K (2011). Micropropagation of *Malaxis acuminata* D. Don: A Rare Orchid of High Therapeutic Value. Open Access J. Med. Aromat. Plants 1:29-33.
- Khan M, Omoloso A (2004). Antibacterial activity of *Galeola foliata*. Fitoterapia 75:494-496.
- Khasim S, Rao PRM (1999). Medicinal importance of orchids. The Botanica 49:86-91.
- Knudson L (1922). Nonsymbiotic germination of orchid seeds. Botanical Gazette: pp.1-25.
- Knudson L (1925). Physiological study of the symbiotic germination of orchid seeds. Bot Gaz 79:345-379.
- Knudson L (1927). Symbiosis and asymbiosis relative to orchids. New Phytol. 26:328-336.

- Knudson L (1946). A nutrient for germination of orchid seeds. Am. Orchid Soc. Bull.15:214-217.
- Kong JM, Goh NK, Chia LS, Chia TF (2003). Recent advances in traditional plant drugs and orchids. Acta Pharmacologica Sinica 24:7-21
- Kumar PKS, Subramoniam A, Pushpangadan P (2000). Aphrodisiac activity of *Vanda tessellata* (Roxb.) Hook. ex Don extract in male mice. Ind. J. Pharma. 32:300-304.
- Langham W (1579). The garden of health; London.
- Lawler L, Slaytor M (1970). Uses of Australian orchids by Aborigines and early settlers. The Medical journal of Australia 2:1259.
- Lesar H, Hlebec B, Čeranic N, Kasteles D, Luthar Z (2012). Acclimatization of terrestrial orchid *Bletilla striata* Rchb.f.(Orchidaceae) propagated under *in vitro* conditions. Acta agriculturae Slovenica, 99(1):69-75.
- Li H, Yan Z, Zhou Z, Xu L, Daikonya A, Wang J (2006). Anti-allergic agents from natural sources. Heterocycles 68:1259-1265.
- Li M, Zou D (1995). Study on pharmacology activity of *Anoectochilus* roxburghii from three sources. Information Chin. Pharmacol. Soc. 3:26-28
- Li YM, Wang HY, Liu GQ (2001). Erianin induces apoptosis in human leukemia HL-60 cells. Acta Pharmacologica Sinica 22:1018.
- Liu H, Zhang Z (1998). Studies on plantlet strengthening medium for Dendrobium candidum Wall ex. Lindl. clonal propagation in vitro. Zhongyuo Zhongyuo Zachi 23(11):654-656.
- Luo H, Lin S, Ren F, Wu L, Chen L, Sun Y (2007). Antioxidant and antimicrobial capacity of Chinese medicinal herb extracts in raw sheep meat. J. Food Protect. 70:1440-1445.
- Majumder P, Banerjee S, Sen S (1996). Three stilbenoids from the orchid *Agrostophyllum callosum*. Phytochem. 42:847-852.
- Majumder P, Sen R (1991). Pendulin, a polyoxygenated phenanthrene derivative from the orchid *Cymbidium pendulum*. Phytochem. 30:2432-2434.
- Manandhar NP (2002). Plants and People of Nepal. Timber Press, Portland, Oregon, USA.
- Mazumder PB, Sharma GD, Dutta M, Choudhury DN, Das T, Mazumder B (2010). *In vitro* propagation and phytochemical screening of *Papilionanthe teres* (Roxb.) Schltr. Assam University, J. Sci. Technol. Biol. Environ. Sci. 5(1):37-42.
- Medhi R, Chakrabarti S (2009). Traditional knowledge of NE people on conservation of wild orchids. Indian J. Trad. Knowl. 8:11-16.
- Mitra G (1971). Studies on seeds, shoot tips and stem discs of an orchid grown in aseptic culture. Indian J. Exp. Biol 9:79-85.
- Mitra G (1986). *In vitro* culture of orchid seeds for obtaining seedlings. Biology, conservation, and culture of orchids East-West Press.
- Miyazawa M, Shimamura H, Nakamura S, Sugiura W, Kosaka H, Kameoka H (1999). Moscatilin from *Dendrobium nobile*, a naturally occurring bibenzyl compound with potential antimutagenic activity. J. Agric. Food Chem. 47:2163-2167.
- Moerman D (1986). Medicinal plants of the Native Americans. University of Michigan Museum of Anthropology technical report, number 19.University of Michigan. p. 534.
- Moerman DE (1998). Native american ethnobotany. Timber Press Portland,
- Morel G (1960). Producing virus-free *Cymbidiums*; Am. Orchid Soc. Bull 29:495-497.
- Morris B (2003). Children of the Wind-Orchids as Medicines in Malawi. The Orchid Review, 111:271-277.
- Murashige T, Skoog F (1962). A revised medium for rapid growth and bio assays with tobacco tissue cultures. Physiol. Plant. 15:473-497.
- Nalawade SM, Sagare AP, Lee CY, Kao CL, Tsay HS (2003). Studies on tissue culture of Chinese medicinal plant resources in Taiwan and their sustainable utilization Bot. Bull. Acad. Sin. 44:79-98.
- Nayak NR, Chand PK, Rath SP, Patnaik SN (1998). Influence of some plant growth regulators on the growth and organogenesis of *Cymbidium aloifolium* (L.) Sw. seed derived rhizomes *in vitro*. In vitro Cell. Dev. Biol. Plant. 34:185-188.
- Nongdam P, Chongtham N (2011). In vitro rapid propagation of Cymbidium aloifolium (L.) Sw.: A medicinally important orchid via seed culture. J. Biol. Sci. 11:254-260.
- Okamoto T, Natsume M, Onaka T, Uchmaru F, Shimizu M (1966). The structure of dendramine (6-oxydendrobine) and 6-oxydendroxine. The

- fourth and fifth alkaloid from *Dendrobium nobile*. Chem. Pharm. Bull. 14:676-680.
- Pant B, Chaudhary RP, Subedi A, Shakya LR (2002). Nepalese Himalayan Orchids and the conservation priorities. In :Proceeding of International Seminar on Mountains. Royal Nepal Acad. Sci. Technol. pp. 485-495.
- Pant B, Gurung R (2005). *In vitro* seed germination and seedling development in *Aerides odorata* Lour. J. Orchid Soc. India 19:51-55.
- Pant B, Swar S, Gurung, R (2007). Current Status and ex situ conservation of threatened orchids of Nepal. In:Proceedings 9th Asia Pacific Orchid Conference (APOC 9), Seol, Korea. pp.307-318.
- Pant B (2008). Application of tissue culture for conservation of medicinal plants. In:PK. Jha et al (eds). Medicinal plants in Nepal:An anthology of contemporary research, Publisher, Ecological Society (ECOS) Kathmandu Nepal. pp. 240-245.
- Pant B, Swar S, Karanjeet A (2008). Micropropagation of *Coelogyne cristata* Lindl. The Journal of Orchid Society of India. 22(1,2):45-48.
- Pant B, Shrestha S, Pradhan S (2011). *In vitro* seed germination and seedling development of *Phaius tancarvilleae* (L'Her.) Blume. Scientific World 9:50-52.
- Pant B, Thapa D (2012). *In vitro* mass propagation of an epiphytic orchid, *Dendrobium primulinum* Lindl. through shoot tip culture. Afr. J. Biotechnol. 11:9970-9974.
- Pant B, Raskoti BB (2013). Medicinal Orchids of Nepal. Himalayan Map House, Pvt. Ltd. (Publisher).
- Paudel MR, Pant B (2012). In vitro plant regeneration of Esmeralda clarkei Rchb. f. via protocorm explant. Afr. J. Biotechnol. 11:11704-11708.
- Pongener A, Deb CR (2011). *In vitro* regeneration of plantlets of *Cymbidium iridioides D. Don* using nodal segments as explants. Int. J. Appl. Biotechnol. Biochem. 1(4):389-400.
- Pradhan S, Pant B (2010). *In vitro* seed germination in *Cymbidium elegans* Lindl. and *Dendrobium densiflorum* Lindl. ex Wall.(Orchidaceae). Botanica Orientalis: J.Plant Sci. 6:100-102.
- Pradhan S, Paudel YP, Pant B (2013). Efficient regeneration of plants from shoot tip explants of *Dendrobium densiflorum* Lindl., a medicinal orchid. Afr. J. Biotechnol. 12(12):1378-1383.
- Prasad DN, Achari G (1966). A study of anti-arthritic action of *Vanda roxburghii* in albino rats. J. Indian Med. Assoc. 46:234-237.
- Puri H (1970) Salep-the drug from orchids. Bull. Am. Orchid Soc. p.39. Rao A (1977). Tissue culture in the orchid industry. Applied and
- Fundamental Aspects of Plant Cell Tissue and Organ Culture. J. Reinert and YPS Bajaj (eds.). McGraw-Hill, New York:pp.44-69.
- Reinikka MA (1995). A history of the Orchid. Portland Timber Press.
- Richards J , Cribb P (1998). The genus *Cypripedium*. Royal Botanic Gardens, Kew Botanical Magazine Monograph. Timber Press, Portland, Oregon.
- Rederbangh K, Fujii JA, Slade D (1993). Hydrated coatings of synthetic seeds. In: Synseeds (Ed.): K. Redenbaugh. CRC Press, Boca Raton. pp.35-46.
- Sharma U, Rama R, Mohan J, Reddy A (2007). *In vitro* propagation of *Dendrobium microbulbon* A. Rich a rare ethnomedicinal herb. Indian J. Biotechnol. 6:381-384.
- Sharma N, Chandel KPS (1996). *In vitro* conservation of *Orchis latifolia*: A threatened, medicinal terrestrial orchid Indian J. Plant Genet. Res. 9:109-113.
- Shimura H, Matsuura M, Takada N, Koda Y (2007). An antifungal compound involved in symbiotic germination of *Cypripedium macranthos* var. *rebunense* (Orchidaceae). Phytochem. 68:1442-1447.
- Shiau YJ, Nalawade SM, Hsia CN, Mulabagal V, Tsay HS (2005). *In vitro* propagation of Chinese medicinal plant, *Dendrobium candidum* Wall. Ex. Lindl. from axennic nodal segments. *In Vitro* Cell. Dev. Biol. Plant. 41:666-670.
- Shrestha R (2000). Some medicinal orchids of Nepal. In:The Himalayan plants, can they save us? Proceeding of Nepal-Japan joint symposium on conservation and utilization of Himalayan medicinal resources (Eds. T. Watanabe, A. Takano, M.S. Bista and H.K. Saiju), Society for the Conservation and Development of Himalayan Medicinal Resources (SCDHMR). pp. 153-156.
- Singh A, Duggal S (2009). Medicinal Orchids: An Overview. Ethnobotanical Leaflets, 13:351-363.

- Shyur LF, Chen CH, Lo CP, Wang SY, Kang PL, Sun SJ, Chang CA, Tzeng CM, Yang NS (2004). Induction of apoptosis in MCF-7 human breast cancer cells by phytochemicals from *Anoectochilus formosanus*. J. Biomed. Sci. 11:928-939.
- Stewart SL, Kane ME (2006). Asymbiotic seed germination and *in vitro* seedling development of *Habenaria macroceratitis* (Orchidaceae), a rare Florida terrestrial orchid. Plant Cell Tiss. Org. Cult. 86:147-158.
- Subedi A (2011). New species, pollinator interactions and pharmaceutical potential of Himalayan orchids. Ph.D. Thesis, Leiden University, The Netherlands.
- Suresh PK, Subramoniam A, Pushpangadan P (2000). Aphrodisiac activity of *Vanda tessellata*. Indian J. Pharmacol. 32:300-304.
- Swarts N (2007). Integrated conservation of the rare and endangered terrestrial orchid Caladenia huegelii HG Reichb. Ph.D. Thesis, Australia.
- Swarts ND, Dixon KW (2009). Terrestrial orchid conservation in the age of extinction. Ann. Bot. 104:543-556.
- Szlachetko DL (2001). Genera et species Orchidalium. 1. Polish Bot. J. 46:11-26.
- Tseng CC, Shang HF, Wang LF, Su B, Hsu CC, Kao HY, Cheng KT (2006). Antitumor and immunostimulating effects of *Anoectochilus formosanus Hayata*. Phytomed. 13:366-370.
- Turner W (1568). The first and seconde partes of the Herbal of William Turner, doctor in Phisick, lately oversene, corrected and enlarged with the Third Parte. Cologne,. (Original publication 1551).
- Vacin EF, Went F (1949). Some pH changes in nutrient solutions. Botanical Gazette: pp.605-613.
- Vaidya B, Shrestha M, Joshee N (2002). Report on Nepalese orchid species with medicinal properties. Proceeding of Nepal-Japan joint Symposium on conservation and utilization of Himalayan medicinal resources. Society for the Conservation and Development of Himalayan Medicinal Resources (SCDHMR), Japan, pp.146-152.
- Vij S (1993) Regeneration response of orchid roots: A study in vitro. J. Orchid Soc. India 7:61-72.
- Vij S (2002). Orchids and tissue culture: current status. Role of plant tissue culture. In biodiversity conservation and economic development. Gyanodaya Prakashan, Nainital India. p.491.
- Wang J, Matsuzaki K, Kitanaka S (2006). Stilbene derivatives from *Pholidota chinensis* and their anti-inflammatory activity. Chem. Pharma. Bull. 54:1216-1218.

- Watanabe K, Tanaka R, Sakurai H, Iguchi K, Yamada Y, Hsu CS, Sakuma C, Kikuchi H, Shibayama H, Kawai T (2007). Structure of cymbidine A, a monomeric peptidoglycan-related compound with hypotensive and diuretic activities, isolated from a higher plant, Cymbidium goeringii (Orchidaceae). Chem. Pharma. Bull. 55:780-783.
- White KJ, Sharma B (2000). Wild orchids in Nepal: the guide to the Himalayan orchids of the Tribhuvan Rajpath and Chitwan Jungle: Bangkok, Thailand: White Lotus Press.
- Williams CA (1979) The leaf flavonoids of the Orchidaceae. Phytochem. 18:803-813.
- Wilson MF (2007). Medicinal plant fact sheet: *Cypripedium*:lady's slipper orchids. Virginia: Arlington.
- Wimber D (1963). Clonal multiplication of *Cymbidiums* through tissue culture of the shoot meristem. Am. Orchid Soc. Bull. 32:105-107.
- Won JH, Kim JY, Yun KJ, Lee JH, Back NI, Chung HG, Chung SA, Jeong TS, Choi MS, Lee KT (2006). Gigantol isolated from the whole plants of *Cymbidium goeringii* inhibits the LPS-induced iNOS and COX- 2 expression via NF-kappaB inactivation in RAW 264.7macrophages cells. Planta Med.72:1181-1187.
- Yam TW, Arditti J, Cameron KM (2009). "The orchids have been a splendid sport"-an alternative look at Charles Darwin's contribution to orchid biology. Am. J. Bot. 96:2128-2154.
- Yang L, Wang Z, Xu L (2006). Simultaneous determination of phenols (Bibenzyl, phenanthrene, and fluorene) in *Dendrobium* species by high-performance liquid chromatography with diode array detection. J. Chromatogr. A 1104:230-237.
- Yi Y, Xing F, Huang X, Chen H, WANG F (2005). Medicinal plants of *Bulbophyllum* species in China. J. Trop. Subtrop. Bot. 13:65-69.
- Zhang D, Zhang Y, Liu G, Zhang J (2006). Dactylorhin B reduces toxic effects of β-amyloid fragment (25-35) on neuron cells and isolated rat brain mitochondria. Naunyn-Schmiedeberg's Arch. Pharmacol. 374:117-125.
- Zhao C, Liu Q, Halaweish F, Shao B, Ye Y, Zhao W (2003).
 Copacamphane, Picrotoxane, and Alloaromadendrane
 Sesquiterpene Glycosides and Phenolic Glycosides from
 Dendrobium moniliforme. J. Nat. Prod. 66:1140-1143.