

Full Length Research Paper

Evaluation of antioxidant activity, quantitative estimation of phenols and flavonoids in different parts of *Aegle marmelos*

Nadeem Ahmad Siddique¹, Mohd Mujeeb^{1*}, Abdul Kalam Najmi² and Mohd Akram³

¹Department of Pharmacognosy and Phytochemistry, Faculty of Pharmacy, Jamia Hamdard (Hamdard University) New Delhi-110062, India.

²Department of Pharmacology, Faculty of Pharmacy, Jamia Hamdard (Hamdard University) New Delhi-110062, India.

³Department of preventive and social medicine, Faculty of medicine, Jamia Hamdard (Hamdard University) New Delhi-110062, India.

Accepted 5 of August, 2009

In existing study, we carried out an efficient record of the comparative antioxidant activity in methanolic extract of the selected parts (leaves, root and stem bark) of *Aegle marmelos*. Total content of phenol and flavonoid was quantitatively estimated in different parts of *A. marmelos*. The total phenolic contents varied from 9.8367 ± 0.0235 to 1.7281 ± 0.049 mg g⁻¹. Total flavonoid contents were between 8.248 ± 0.029 to 1.087 ± 0.002 mg g⁻¹. Free radical scavenging activity of different extracts was evaluated by using DPPH (1, 1 -Diphenyl- 2 -picryl hydrazyl) method. The highest free radical scavenging effect was observed in leaves with IC₅₀ = 2.096µg ml⁻¹. The effectiveness of radical scavenging activity of leaves extract was about 10 times greater than reference antioxidant butylated hydroxy toluene (BHT). The greater amount of phenolic compounds leads to more powerful radical scavenging effect as shown by methanolic extract of *A. marmelos* leaves.

Key words: *Aegle marmelos*, antioxidant, flavonoids, phenols, 1, 1 -diphenyl - 2 -picryl hydrazyl.

INTRODUCTION

In recent years, considerable interest has been evinced by the public and the medical professional regarding the use of indigenous drugs in the treatment of diseases. Several members of the family Rutaceae are being used traditionally for a wide variety of ethnomedical properties. *Aegle marmelose* (L) (Rutaceae) is 1 among them found in India. *A. marmelose* generally acknowledged as bael or koovalam (Malayalam, India) growing wild through out deciduous forest of India, climbing to a height of 1,200 m in Western Himalayas and also occurring in Andaman Island. Its fruits and leaves are valued in indigenous medicine (Charakbraty et al., 1960). The plant has been employed for long time in folk therapy. Poultice made of leaves are used for ophthalmia and ulcers. The leaves are use to lowering the blood glucose levels (Ayurvedic Pharmacopoeia of India, 1988). Other actions like anti-fungal (Renu, 1983), antibacterial (Banerji and Kumar,

1980), antiprotozoal (Banerjee, 1980), antispermatogenic (Sur et al, 1999) are also reported. The plant has been found to contain number of phytoconstituents like aegeline, agelinine, rutin, sterol (Chatterjee and Bose, 1952), β-sitosterol, β -D-glucoside, marmesinine (Sharma et al, 1980), lupeol (Patra et al., 1979), tannins, phlobatannins, flavonoids, umbelliferone, quercetin and volatile oils (Eugenol and methyl eugenol) (Banerjee, and Nigam, 1979). It has been reported that leaves possess cardiotoxic effect, antifungal, analgesic and antioxidant activities (Rai, 1996). No scientific evaluation of antioxidant activity of *A. marmelose* has been reported so far. Therefore, it was thought worthwhile to evaluate antioxidant activity of *A. marmelos* to confirm its folk medicine claim. Many naturally occurring products have been reported to contain large amount of antioxidant other than vitamin C, E and carotenoid (Javanmardi et al., 2003). These antioxidant play a vital role in delaying, intercepting or preventing oxidative reactions, catalyse by free radical (Vilioglu et al., 1998). This antioxidant activity might be due to the presence of phenolic compounds such as

*Corresponding author. E-mail: mujeeb_zaidi@yahoo.co.in.

flavonoids (Pieatta, 1998), phenolic acids and phenolic diterpene (Shahidi and Wanasundara, 1992). Antioxidants may guard against reactive oxygen species (ROS) toxicities by the prevention of ROS construction, by the disruption of ROS attack, by scavenging reactive metabolites and converting them to less reactive molecules or by enhancing the resistance of sensitive biological target to ROS attack (Sen, 1995). Free radicals, reactive oxygen species (ROS) and reactive nitrogen species (RNS) are associated with many pathological conditions such as atherosclerosis, arthritis, ischemia, reperfusion injury of many tissues, central nervous system injury, gastritis, cancer and AIDS (Kumpulainen and Salonen, 1999; Cook and Samman, 1996). Synthetic antioxidants like butylated hydroxy anisole (BHA, butylated hydroxy toluene (BHT), tertiary butylated hydroxy quinone and gallic acid esters have been suspected to be carcinogenic. Hence, strong limitations have been placed on their use and there is a trend to replace them with naturally occurring antioxidants. Moreover, these synthetic antioxidants also show low solubility and moderate antioxidant activity (Barlow, 1990; Branen, 1975). Hence, search for natural antioxidant has greatly been increased in the recent scenario (Jayaprakasha et al., 2003). In the literature many crude extracts and pure natural compounds have been reported which have potent antioxidant potential (Schuler, 1990; Chu, 2000; Koleva et al., 2002; Mantle et al., 2000; Oke and Hamburger, 2002). However there is still a need to find out more effective antioxidant having fewer side effects from natural source. It has been found out that plant having polyphenolic compounds such as flavonoids possess antioxidant activity (Cook and Samman, 1996). Some evidence suggests that the biological actions of these compounds are related to their antioxidant activity (Gryglewski et al., 1987). In the present investigation, phytochemical screening of methanolic extracts of different parts of *A. marmelos* revealed the presence of phenolic and flavonoids compounds. Hence the present study was design to evaluate antioxidant activity of *A. marmelos*.

MATERIALS AND METHODS

Chemicals

1, 1-Diphenyl-2-picryl hydrazyl (DPPH), rutin and ascorbic acid were purchased from Sigma Chemical Co. (St., Louis, USA). Tert-butyl-4-hydroxy toluene (BHT), gallic acid, Folin Ciocalteu reagent, and methanol were purchased from Merck Co. (Germany). All the chemicals and reagents used were of analytical grade.

Plant materials

Different parts of *A. marmelos* were collected from campus of Hamdard University, New Delhi, India, (July 2007), which was identified by Taxonomist, Department of Botany, Hamdard New Delhi. The voucher specimens were deposited in the Department of Pharmacognosy and Phytochemistry, Faculty of Pharmacy, Jamia

Hamdard (JHFP, 2023). Plants materials were shade dried at room temperature and ground in a mortar. 25g of each parts of *A. marmelos* powder were extracted in 250 ml of methanol by maceration for 48 h (yield-18.5%). The extracts were concentrated in vacuo at 50°C and the extracts were freeze dried.

Total phenols estimation

The total phenols of all extracts were measured at 765 nm by Folin Ciocalteu reagent (McDonald et al., 2001). The dilute methanolic extract (0.5 ml of 1:10 g ml⁻¹) or gallic acid (standard phenolic compound) was mixed with Folin Ciocalteu reagent (5 ml, 1:10 diluted with distilled water) and aqueous sodium carbonate (4 ml, 1 M). The mixture was allowed to stand for 15 min and the total phenols were determined by spectrophotometer at 765 nm. The standard curve was prepared using 0, 50, 100, 150, 200, 250 mg l⁻¹ solutions of gallic acid in methanol: water (50:50, v/v). Total phenol values are expressed in terms of gallic acid equivalent (mg g⁻¹ of dry mass), which is a common reference compound.

Total flavonoids estimation

Aluminium chloride colorimetric technique was used for flavonoids estimation (Chang et al., 2002). Each extract (0.5 ml of 1:10 g ml⁻¹) in methanol was separately mixed with 1.5 ml of methanol, 0.1 ml of 10% aluminium chloride, 0.1 ml of 1 M potassium acetate and 2.8 ml of distilled water. It was left at room temperature for 30 min after which the absorbance of the reaction mixture was measured at 415 nm with a double beam UV/Visible spectrophotometer, SHEMADZU (Japan). The calibration curve was plotted by preparing the quercetin solutions at concentrations 12.5 to 100 g ml⁻¹ in methanol.

Free radical scavenging activity

DPPH assay

The free radical scavenging activity of the different parts of *A. marmelos*, butylated hydroxy toluene (BHT), rutin and ascorbic acid was measured in terms of hydrogen donating or free radical scavenging ability by using the stable 1,1-diphenyl-2-picryl hydrazyl radical (DPPH) (Blois, 1957). Different concentration of methanolic plant extract (0.5 ml) was mixed in a test tube with a mixture of 2.5 ml of methanol and 75 µM DPPH, (stable free radical) and record the absorbance at 517 nm. The reaction mixture was set-aside in the dark at room temperature for 90 min and absorbance was recorded at 517 nm. The experiment was done in triplicate. BHT, rutin and ascorbic acid were used as standard controls. IC₅₀ value is the concentration of sample, required to scavenge 50% of DPPH free radicals.

Statistical analysis

The statistical significance between antioxidant activity values of the extracts were evaluated by analysis of variance (ANOVA) followed by Dunett's test. P values less than 0.05 were considered to be statistically significant.

RESULTS AND DISCUSSION

Flavonoid and total phenol contents of the methanolic extracts

Over the years, the study on medicinal plants to reveal

the mechanism of action and to justify their claims by traditional healers has been increase. An angle of this research has been the study of bioactive components and antioxidant properties of the *A. marmelos*. The present study has verified that remedial plants could be good source of antioxidant substances. It has been acknowledged that flavonoids show significant antioxidant action on human health and fitness. The flavonoids act through scavenging or chelating process (Kessler et al., 2003; Cook and Samman, 1996). The high potential of phenolics to scavenge free radicals may be due to many phenolic hydroxyl groups they possess (Sawa et al., 1999). The contents of total flavonoid that were measured by aluminium chloride colorimetric technique in term of quercetin equivalent (the standard curve equation: $y = 0.006x + 0.007$, $r^2 = 0.999$) were between $1.087 \pm$ contents in the extract of leaves ($8.248 \pm 0.029 \text{ mg kg}^{-1}$) and stem ($1.400 \pm 0.029 \text{ mg kg}^{-1}$) were higher than that in the extracts of root ($1.087 \pm 0.002 \text{ mg kg}^{-1}$) (Table 1). Table 2 also show the contents of total phenols that were measured by Folin Ciocalteu reagent in terms of gallic acid equivalent (standard curve equation: $y = 0.004x + 0.003$, $r^2 = 0.991$). The total phenol varied from 1.7281 ± 0.049 to $9.8367 \pm 0.02335 \text{ mg kg}^{-1}$. The total phenol in methanolic extract of the leaves ($9.8367 \pm 0.0235 \text{ mg kg}^{-1}$) and in stem extract ($7.4693 \pm 0.047 \text{ mg kg}^{-1}$) were higher than that in the extracts of root ($1.7281 \pm 0.049 \text{ mg kg}^{-1}$). Table 2 shows the comparison of DPPH free radical inhibitory concentration of the plant extracts and those of BHT, ascorbic acid and rutin. The % inhibition of leaf (64.12 ± 0.01), stem (76.883 ± 0.03) and root are (64.193 ± 0.05) in comparison to BHT (65.09 ± 0.22), ascorbic acid (52.163 ± 0.02) and rutin (72.686 ± 0.560) respectively. The compounds such as flavonoids, which hold hydroxyls groups, are responsible for the radical scavenging activity in the plants (Das and Pereira, 1990; Younes, 1981).

Antioxidant activity

Antioxidants are significant in the prevention of human illness and may function as free radical scavengers, complexes of pro-oxidant metals, reducing agents and quencher of singlet oxygen formation (Andlauer and Furst, 1998). Free radicals possess the ability to reduce the oxidative damage associated with many disease including neurodegenerative diseases, cancer, cardiovascular disease, cataracts and AIDS (Pietta et al., 1998; Lee et al., 2000; Middleton et al., 2000). Antioxidants through their scavenging power are useful for the management of these diseases. DPPH stable free radical method is an easy, rapid and sensitive way to survey the antioxidant activity of a specific compound or plant extracts (Blois, 1958). Figure 1 shows the IC₅₀ ($\mu\text{g ml}^{-1}$) values of plant extracts for free radical scavenging activity by DPPH radical. IC₅₀ of the standard compounds, BHT, ascorbic

Table 1. Total phenolic and flavonoid contents of different parts extract of *A. marmelos*.

Plant parts	Total phenolic contents (mg/kg)	Total flavonoid contents (mg/kg)
Leaf	9.8367 ± 0.0235	8.248 ± 0.029
Stem	7.4693 ± 0.047	1.400 ± 0.029
Root	1.7281 ± 0.049	1.087 ± 0.002

Each value in the table was obtained by calculating the average of three experiments \pm SEM.

Table 2. Comparison of DPPH free radical inhibitory concentration of the plant extracts and those of BHT, ascorbic acid and rutin.

Plant parts	Concentration ($\mu\text{g/ml}$)	% inhibition
Leaf	3.5	64.12 ± 0.01
Stem	3.5	76.883 ± 0.03
Root	3.5	64.193 ± 0.05
BHT	35	65.09 ± 0.22
Ascorbic acid	35	52.163 ± 0.20
Rutin	35	72.686 ± 0.56

Each value in the table was obtained by calculating the average of three experiments \pm SEM.

ic acid and rutin were 18.726 , 29.338 and $12.231 \mu\text{g ml}^{-1}$ respectively. The highest radical scavenging activity was shown by leaf extract with $\text{IC}_{50} = 2.096 \mu\text{g ml}^{-1}$ which is higher than that of BHT ($P < 0.05$). The radical scavenging activity in the plant extracts decreased in the following order: Leaves > stem > root. The radical scavenging effect of leaves at $3.5 \mu\text{g ml}^{-1}$ was similar to BHT at $35 \mu\text{g ml}^{-1}$. Therefore, the antioxidant effect of leaves extract was 10 times greater than that of the synthetic antioxidant, BHT.

Conclusion

The result of the present study revealed that the leaf extract of *A. marmelos*, which hold maximum amount of flavonoid and phenolic compounds, exhibited the best antioxidant activity. Despite widespread use of *A. marmelos* as folklore medicines in India, the literature contains few reports on its antioxidant activity. In present experiment, we conceded out a systematic record on the relative free radical scavenging activity in methanolic extract of selected parts of *A. marmelos*. We have also established the relationship of total flavonoid and phenolic contents by means of free radical scavenging activity.

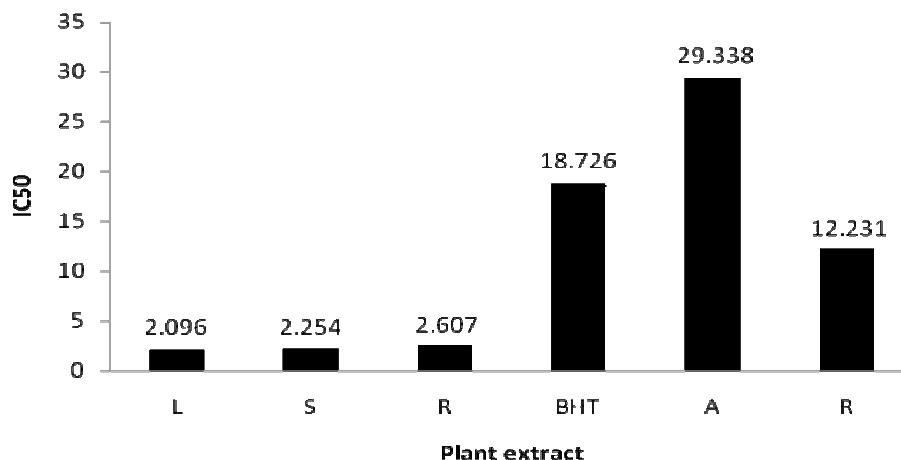


Figure 1. IC₅₀ (µgml⁻¹) values of plant extracts for free radical scavenging activity by DPPH radical. Lower IC₅₀ value indicates higher antioxidant activity. Extracts: L = Leaves, S = Stem, R = Root, BHT = Butylated hydroxyl toluene, A = Ascorbic acid and R = Rutin.

In this study, *A. marmelos* recognized as having high levels of antioxidant activity. Leaf extract of *A. marmelos* showed higher scavenging property it may be due to the present of hydroxyl groups existing in the phenolic and flavonoid compounds chemical configuration that can provide the essential constituents as a radical scavenger. Free radicals have been reported to be liable for the cataract formation, oxidative damage in the occurrence and development of vascular diseases (Langsethm, 1995; Alho and Leinonen, 1999). Free radical mediated processes have been implicated in the pathogenesis of most of the diseases. It is well documented that free radicals take part in the pathogenesis of a large number of diseases (Gyamfi et al., 1999). The present study showed that all extracts demonstrated different extent of antioxidant activity. It was also shown that leaf extract showed significantly higher antioxidant activity than BHT, rutin and ascorbic acid in scavenging of DPPH free radical. This may be support to the high amount of flavonoid and phenolic compounds in the mehanolic extract of *A. marmelos*.

ACKNOWLEDGMENT

The authors place, grateful and their heart felt thanks to Department of Pharmacognosy and Phytochemistry, Faculty of Pharmacy Jamia Hamdard New Delhi, for their valuable assistance for this study.

REFERENCES

Alho H, Leinonen J (1999). Total antioxidant activity measured by chemiluminescence method. *Meth. Enzymol.* 299: 3-15.
 Ayurvedic Pharmacopoeia of India (1988). The Ayurvedic Pharmacopoeia of India. Vol- IV, Part- I, Ist-Edition, Govt. of India, Ministry

of health and family welfare, Deptt. of AYUSH, New Delhi.
 Andlauer W, Furst P (1998). Antioxidative power of phytochemicals with special reference to cereales. *Cereal Foods World* 43:356-359.
 Banerjee A (1980). Antimicrobial and anti helminthic screening of the fixed oil and unsaponifiable matter of *Aegle marmelos*. *Conf. Proc. 67th session, Indian Science Congress. Calcutta Part iii. Chemistry Section, abstr.*, 247.
 Banerjee A, Nigam SS (1979). Studies on the fixed oil from the seeds of *Aegle marmelos* *Corr. J. Am. Oil Chem. Soc.*, 56: 647.
 Banerji N, Kumar R (1980) Studies on the seed oil of *A. Marmelos* and its effect on some bacterial species. *J. Inst. Chem., Calcutta*, 52.
 Barlow SM (1990). Toxicological aspects of antioxidants used as food additives. In *Food Antioxidants*, Hudson BJJ (ed.) Elsevier, London, pp 253-307.
 Blois MS (1958). Antioxidant determinations by the use of stable free radicals. *Nature* 26:1199- 1200.
 Branen AL (1975). Toxicology and biochemistry of butylated hydroxyanisole and butylated hydroxytoluene. *J. Am. Oil Chem. Soc.* 5: 59-63.
 Charakbraty B, Malik C, Bhatthacharya S (1960). Studies on the effect of green leaves of *Aegle marmelos* and *Piper nigrum* on the glucose and cholesterol levels of blood in diabetes mellitus. *Indaian. Med. Forum* 9: 285-28.
 Chang C, Yang M, Wen H, Chern J (2002) Estimation of total flavonoid content in propolis by two complementary colorimetric methods. *J. Food Drug Anal.* 10: 178-182.
 Chu Y (2000) Flavonoid content of several vegetables and their antioxidant activity, *J. Sci. Food Agric.* 80: 561-566.
 Cook NC, Samman S (1996) Flavonoids- chemistry, metabolism, cardioprotective effects, and dietary sources. *Nutr. Biochem.* 7: 66-76.
 Gyamfi MA, Yonamine M, Aniya Y (1999) Free- radical scavenging action of medicinal herbs from Ghana *Thonningia sanguinea* on experimentally- induced liver injuries. *General Pharmacol.* 32: 661- 667.
 Halliwell B (1994) Free radicals, antioxidants, and human disease: curiosity, cause, or consequence? *Lancet* 344: 721-724.
 Javanmardi J, Stushnoff C, Locke E, Vivaco JM (2003). Antioxident activity and total phenolic content of Iranian *Ocimum accessions*. *Food Chem.* 83:547-550.
 Kessler M, Ubeaud G, Jung L (2003) Anti- and pro-oxidant activity of rutin and quercetin derivatives. *J. Pharm. Pharmacol.* 55: 131-142.
 Kumpulainen JT, Salonen JT (1999). Natural Antioxidants and Anticarcinogens in Nutrition, Health and Disease, The Royal Society of Chemistry, UK pp. 178-187.
 Langsthem L (1995). Oxidants, antioxidants and disease prevention, ILSI Press: Washington, DC.

- Lee KG, Mitchell AE, Shibamoto T (2000). Determination of antioxidant properties of aroma extracts from various beans. *J. Agric. Food Chem.* 48: 4817-4820.
- Mantle D, Eddeb F, Pickering AT (2000) Comparison of relative antioxidant activities of British medicinal plant species *in vitro*. *J. Ethnopharmacol.* 72: 47-51.
- McDonald S, Prenzler PD, Autolovich M, Robards K (2001) Phenolic content and antioxidant activity of olive extracts. *Food Chem.* 73:73-84.
- Middleton E, Kandaswami C, Theoharides TC (2000). The effects of plants flavonoid on mammalian cell: implication for inflammation, heart disease, and cancer. *Pharmacol. Rev.* 52:673-751.
- Pietta P, Simonetti P, Mauri P (1998). Antioxidant activity of selected medicinal plants. *J. Agric. Food Chem.* 46: 4487-4490.
- Pereira TA (1990). Effects of flavonoids on thermal autooxidation of Palm oil: structure- activity relationship. *J. Am. Oil Chem. Soc.* 67: 255-258.
- Rai MK (1996). *In vitro* evaluation of medicinal plant extract against *Pestalotiopsis mangiferae*. *Hindustan Antibiot. Bull.* 38(1-4): 53-56
- Renu (1983). Fungitoxicity of leaf extracts of some higher plants against *Rhizoctonia solani* Kuehn. *Natl. Acad. Sci. Lett.* 6: 245-246.
- Sawa T, Nako M, Akaike T, Ono K, Maeda H (1999). Alkylperoxyl radical scavenging activity of various flavonoids and other phenolics compounds: Implications for the antitumor promoter effect of vegetables. *J. Agric. Food Chem.*, 47: 397- 492.
- Schuler P (1990) Natural antioxidants exploited commercially, In *Food Antioxidants*, Hudson BJJ (ed.). Elsevier, London, pp 99-170.
- Shahidi F, Wanasundara PKJPD (1992) Phenolic antioxidants. *Crit. Rev. Food Sci. Nutr.* 32: 67-103.
- Sen CK (1995). Oxygen toxicity and antioxidant: state of the art. *Indian J. Physiol. Pharmacol.* 39(3):177-96.
- Sur TK, Pandit S, Pramanik T (1999). Antispermato-genic activity of leaves of *Aegle marmelos*, corr. In albino rats: a preliminary report. *Biomed* 19:199-202.
- Sharma BR, Rattan RK, Sharma P (1980) Constituent of leaves and fruits of *Aegle marmelos*. *Indian J. Chem.* 19B, 162.
- Vilioglu YS, Mazza G, Gao L, Oomah BD (1998). Antioxidant activity and total phenolics in selected fruits, vegetables and grain products. *J. Agric. Food Chem.*, 46: 4113-4117.
- Younes M (1981) Inhibitory action of some flavonoids on enhanced spontaneous lipid peroxidation following glutathione depletion. *Planta Medica* 43: 240-245.