

Full length Research paper

Mineral nutrition of mycorrhized tropical gum tree *A. senegal* (L.) under water deficiency conditions

Ndiaye Malick^{1*}, Cavalli Eric², Leye El Hadji Malick³, and Diop Tahir Abdoulaye¹

¹Laboratoire de Biotechnologies des Champignons, Département de Biologie Végétale, Faculté des Sciences et Techniques, Université Cheikh Anta Diop, BP. 5005 Dakar-Fann, Sénégal.

²Laboratoire de Nanomédecine, Imagerie et Thérapeutique, EA 4662, UFR Sciences Médicales et Pharmaceutiques, Université de Franche-Comté, 19 rue Ambroise Paré, 25030 Besançon cedex, France.

³Laboratoire National de Recherche sur les Productions Végétales/ Institut Sénégalais de Recherche Agricole, 'LNRPV/ISRA', Dakar, Sénégal.

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A pot experiment was set to examine the effects of arbuscular mycorrhizal fungi (AMF) on mineral nutrition of a tropical legume tree (*Acacia senegal*) under three different water status. *Acacia senegal* seedlings were inoculated with three species of AMF, *Glomus intraradices*, *Glomus fasciculatum* or *Glomus mosseae*. Three water levels (field capacity, moderate water deficiency and severe water deficiency) were applied to the pots after transplantation. *A. senegal* seedlings were colonized by the three AM fungi. Twelve weeks after water stress imposition, uptake nutrient of *A. Senegal* was enhanced by mycorrhizal inoculation under moderate and severe water status. Root colonization varied from 30.4 to 62.5%. The lowest intensity (30.4%) was observed on field capacity associated with *G. intraradices* and the highest root colonization (62.5%) was observed on severe water deficiency associated with *G. fasciculatum*. Relative improvement was noted in the foliar nutrient content nitrogen (N), phosphorus (P), potassium (K) and shoot water content of the inoculated plants, whatever the water regime. Mycorrhizal inoculation has no significant effect on shoot calcium (Ca), magnesium (Mg) and sodium (Na) compared to uninoculated plants. *G. fasciculatum* was the most efficiency fungus in nutrient foliar of *A. senegal* plants under water deficiency conditions. Inoculating *A. senegal* plant with the arbuscular mycorrhizal fungus *G. fasciculatum* increased ability to acquire N, P, and K under water deficiency conditions.

Key words: Arbuscular mycorrhizal fungi, *Acacia senegal*, mineral nutrient, water deficiency.

INTRODUCTION

The major factors affecting plant growth in Sahelian zone agrosystem are water and nutrient (Floret et al., 1993). Water availability has been recognized as the most critical determinant of plant survival rate after transplantation in Sahelian zones. Thus, it is necessary to improve the level of efficiency in the plant capture and

use of nutrients which is important in plant growth. In recent years, there has been increasing evidence that the microbial communities of soil and plants have an important role in the development of sustainable agriculture. Among the microorganism living in plant rhizosphere, arbuscular mycorrhizal fungi (AMF) have

*Corresponding author. E-mail: papmalic@yahoo.fr. Tel: + 221 77 534 84 79. Fax: + (221) 33 86466 58.

Table 1. Characteristics of the soil of study.

Component	Contents
Clay	3.6%
Silt	1.6%
Fine silt	2.9%
Fine sand	51%
Coarse sand	40.9%
Organic matter	1.06%
Total carbon	2.5
Total nitrogen	0.33
Conductivity ($\mu\text{S}/\text{cm}$)	658
Total phosphorus (ppm)	47
Available phosphorus (ppm)	3.1
pH (sol/water ratio 1:2)	6.7
pH (sol/KCl ratio 1:2)	4.5

been found to be essential components of sustainable soil-plant systems (Bucher, 2007). Therefore, mycorrhizal inoculation with suitable fungi has been proposed as a promising tool for improving restoration success in semi-arid degraded areas (Garbaye, 2000).

The symbiotic relationship between AMF and the roots of higher plants contributes significantly to plant nutrition and growth (Augé, 2001). These positive responses in productivity to AMF colonization have mainly been attributed to the enhanced uptake by AMF of relatively immobile soil ions such as phosphorus (P), potassium (K), calcium (Ca), magnesium (Mg), and manganese (Mn) (Smith and Read, 2008), but it involves the enhanced uptake and transport of more mobile nitrogen (N) ions, particularly under drought conditions (Liu et al., 2007). Since nutrient mobility is limited under drought conditions, AMF may have a larger impact on overall plant growth and development in water deficiency conditions (Sánchez-Díaz and Honrubia, 1994).

The objective of the present study was to examine the effects of AMF on foliar nutrient of *A. senegal* seedlings under water deficiency conditions.

MATERIALS AND METHODS

Planting of materials

The experiment was conducted at the Department of Plant Biology (University Cheikh Anta Diop/Senegal). The soil used in this study was collected from Sangalkam (50 km from Dakar, Senegal). Soil was sterilized by autoclaving at 12°C for 1 h. Characteristics of the soil were given in Table 1. Seeds were scarified and surface-sterilized with concentrated sulfuric acid for 10 min then washed in sterile distilled water before germinated on sterile water agar at 0.8 at 30°C in the dark for 48 h. The germinated seeds were transferred to plastic bags used in nursery. Four germinated seeds were sown into each bag containing approximately 500 g sandy soil of Sangalkam and thinned to one seedling per pot, 1 week after emergence. All seedlings were watered daily for 1 week to allow proper establishment.

AMF Inoculation

The three species of AMF were used: *Glomus mosseae*, *Glomus fasciculatum* and *Glomus intraradices* obtained from the Laboratory of Fungal Biotechnology (LBC) of the Plant Biology Department (University Cheikh Anta Diop, Sénégal). Mycorrhizal inoculum for each endophyte consisted of mixed soil, spores, mycelium and infected root fragments obtained using maize as host plant. Inoculum from each AM fungus possessed similar infected characteristics (an average of 40 spores per gram and 85% of infected roots).

Experimental design and growth conditions

Plants were inoculated with one of three AMF species by placing 20 g of inoculum directly in the substrate at the position of the roots. The seedlings were subdivided into three blocks. Each block had a plant control and AM inoculation treatment with *G. mosseae*, *G. fasciculatum* or *G. intraradices* each in five replicates. The first block of control plants was kept to 100% of field capacity (FC). The second block of moderate water-deficiency (MWD) plants was maintained at 50% FC and the last block of severe water-deficiency (SWD) plants was maintained at 25% FC. Plants were raised from January to March in a greenhouse with the following conditions: average day/night temperature 30/25 \pm 2°C, and relative humidity maintained between 55 and 65%. Plants were harvested after 3 months.

Measurements and chemical analysis

The percentage of mycorrhizal root infection was estimated by microscope observation of fungal colonization after clearing washed roots in 10% KOH and staining with 0.05% trypan blue in lactophenol (v/v), according to Phillips and Hayman (1970). The relative arbuscular richness (that is, percentage of arbuscules observations among the number of AMF in roots) were calculated using the grid-line intersect method (Giovannetti and Mosse, 1980). Chemical analysis of plants and soil were conducted at the Analysis and Characterization Service (SERAC) of the University of Franche-Comte in France. Plant mineral element compositions [Ca, Mg, sodium (Na), and K] were assessed after digestion with HNO₃ + H₂O₂. Elements were determined using flame atomic emission spectrometry (VARIAN spectra A 220FS) following the French AFNOR standards (FD T90-019, 1984). Kjeldahl Nitrogen was determined by volumetric determination using the French AFNOR standards (NF EN 25663, 1994). The content of the four mineral elements was expressed in mg. kg⁻¹ of dry weight. The phosphorus was determined by atomic absorption (GANIMEDE P) using a molecular adaptation following the French AFNOR standards (NF EN 6878, 2005).

Statistical analysis

Statistical procedures were carried out with the software package R version 2.5. Two factors analysis of variance (ANOVA) was performed to partition the variance into the main effects and the interaction between inoculation and water status.

RESULTS

Symbiotic development

As shown in Table 2, roots of control seedlings were

Table 2. Effects of water status on mycorrhizal colonization (%) of *A. Senegal* seedlings.

Treatment	AM colonization (%)		
	FC	MWD	SWD
Control	0.00 ^d	0.00 ^d	0.00 ^d
<i>G. intraradices</i>	30.40 ^c	37.20 ^{bc}	43.60 ^c
<i>G. fasciculatum</i>	50.10 ^a	55.60 ^a	62.50 ^a
<i>G. mosseae</i>	40.95 ^b	50.35 ^a	55.40 ^b

Values in columns followed by different letters are significantly different ($p \leq 0.05$). FC: Field capacity; MWD: Moderate water deficiency; SWD: Severe water deficiency.

Table 3. Foliar nutrient (mg/kg dry matter) of *A. Senegal* shoot under water status subjected to four different treatments.

Treatment	N	P	K	Na	Ca	Mg
FC						
Control	34.30 ^a	1.58 ^a	11.73 ^b	2.75 ^a	34.18 ^a	6.15 ^a
<i>G. intraradices</i>	32.60 ^a	1.40 ^a	13.22 ^b	1.88 ^b	32.22 ^a	5.16 ^a
<i>G. fasciculatum</i>	39.80 ^a	1.64 ^a	15.71 ^a	2.44 ^a	30.01 ^a	6.10 ^a
<i>G. mosseae</i>	37.40 ^a	1.51 ^a	12.97 ^b	2.23 ^a	29.99 ^a	5.00 ^a
MWD						
Control	36.30 ^c	1.48 ^b	9.63 ^c	2.10 ^a	31.74 ^a	4.41 ^a
<i>G. intraradices</i>	35.80 ^c	1.65 ^b	14.96 ^b	1.70 ^b	25.60 ^b	4.88 ^a
<i>G. fasciculatum</i>	57.00 ^a	2.27 ^a	20.54 ^a	1.93 ^a	30.02 ^b	4.46 ^a
<i>G. mosseae</i>	43.00 ^b	1.70 ^a	17.18 ^a	1.87 ^b	25.46 ^b	4.47 ^a
SWD						
Control	32.80 ^d	1.18 ^b	7.00 ^c	0.89 ^a	25.30 ^a	3.99 ^b
<i>G. intraradices</i>	60.00 ^c	1.55 ^b	10.33 ^b	0.79 ^b	23.50 ^a	3.36 ^b
<i>G. fasciculatum</i>	74.10 ^a	2.02 ^a	17.10 ^a	0.90 ^a	25.28 ^a	5.07 ^a
<i>G. mosseae</i>	67.00 ^b	1.46 ^b	10.38 ^b	0.92 ^a	24.19 ^a	4.05 ^a

Values in columns followed by different letters are significantly different ($p \leq 0.05$). FC: Field capacity; MWD: Moderate water deficiency; SWD: Severe water deficiency.

observed after water treatments, indicating the absence of mycorrhiza. *A. senegal* seedlings were infected by any of AM fungi and the average root colonization varies from 30.4 to 62.5% (Table 2). Results showed that *G. fasciculatum* and *G. intraradices* had the highest and the lowest AM fungus colonization among the three AMF, regardless of water status. Water deficiency increased root colonization.

Foliar nutrient

Nutrient concentrations in the shoots of the seedlings after harvesting are shown in Table 3. *A. senegal* shoot N, P, and K contents showed that the nutrient uptake was a function of the applied treatment. Colonization by *G.*

fasciculatum improved the content of these nutrients more than the other two fungi. The greatest differences among inoculated plants were found for foliar K. This fungal effect was more obvious under water deficiency conditions where nutrient uptake by *A. senegal* plants is more limited. Moreover, the damaging effects of water deficiency conditions on nutrient acquisition of control plant were reduced in inoculated plant (Table 3). As observed for mycorrhizal colonization, the highest foliar concentrations of N, P, and K were seen in the plants inoculated with *G. fasciculatum* under water deficiency condition. Likewise, the field capacity did not have an effect on foliar nutrient concentrations in shoots.

The presence of water deficiency conditions reduced the contents of N, P and K by 13, 25.3 and 36.4%, respectively, in the control plants. However, the most

Table 4. Significance level (F-values) of effects of different factors and factors interaction on foliar nutrient based on analysis of variance (ANOVA).

Variable	Inoculation	Water status	Inoculation x Water status
Df	3	2	6
N	1779.07***	9671.15***	181.10***
P	1636.975***	18.437***	32.294***
K	365.666***	23.892***	47.761***
Na	4.4865**	92.4347***	1.1571 ^{ns}
Ca	5.4989**	30.8064***	1.8789 ^{ns}
Mg	8.6854***	25.7115***	2.0110 ^{ns}

*, **, *** and ns indicate the level of significance at $P \leq 0.05$, 0.0, 0.001 and the absence of significance respectively.

active fungus in increasing water deficiency tolerance was *G. fasciculatum*. It increased N, P, and K concentrations by 125, 86.4 and 144%, respectively over those values recorded in control plants under severe water stress (Table 3). Shoot N, P, and K concentrations for water deficiency conditions were significantly higher than field capacity. Analysis of variance was seen to have significantly increased all foliar nutrient concentration by inoculation and water status. Statistical results also show that combined factors were significant for the N, P, and K contents (Table 4).

Na, Ca, Mg concentration declined with increasing water deficiency conditions (Table 3). Na concentrations of inoculated plant with *G. intraradices* decreased significantly with the water regime. Ca concentrations of inoculated plant decreased significantly under moderate deficiency condition, but not under FC and SWD conditions. Magnesium concentrations in different treatments did not change significantly in comparison with control except for Mg shoot content in severe water deficiency condition.

DISCUSSION

Human activity is one of the main factors reducing soil fertility in the vast Sahelian area. As it was observed in tropical gum trees, water deficiency conditions reduce plant mineral nutrition in areas distress from water limitation. In this study, *G. fasciculatum* was the most efficient fungus in terms of *A. Senegal* performance, and mostly in improving foliar nutrient concentration. Differences among AMF presenting foliar nutrient improvement under water deficiency conditions have already been reported (Qu et al., 2004; Miransari et al., 2007). When soil water is limited, K plays an important role in the control of water relations, helping to maintain a high tissue water level, even under osmotically impaired conditions. It is well documented that K, as the most prominent inorganic solute, plays a key role in

osmoregulation processes in photosynthesis (Smith et al., 2004; Li et al., 2006). It should also be noted that AM plants are able to absorb higher rates of P, which, among other important roles in the plant, can markedly enhance root growth (Miransari et al., 2008; Smith and Read, 2008). The increase of the uptake may help explain the important growth effects obtained in inoculated plants under water deficient conditions (Audet and Charest, 2006).

In our study N, P, K concentrations in inoculated plants were particularly increased under drought conditions. This is in agreement with the studies of other researchers comparing AM species under different stresses (Ruiz-Lozano et al., 2003). While water status significantly affected the concentration of N, P, and K, AMF species exerted a significant effect on all nutrient concentrations. Table 3 shows that the high efficiency of all AMF on the use of these macronutrients by tropical gum tree. Accordingly, previous researchers have found that the efficiency of AM increases with increased level of stress (Audet and Charest, 2006; Subramanian et al., 2006).

Significant interaction of water status and AMF on the concentrations of N, P, and K indicate differential nutrient uptake efficiencies for different AM-host plant combinations (Table 4). In agreement with these ideas, Subramanian and Charest (1997) contend that AM symbiosis improved drought tolerance of a tropical tree, primarily through the enhanced uptake of slowly diffusing nutrients. In fact, foliar nutrient have been found to increase in inoculated plants (Mortimer et al., 2008), but not under water deficiency condition. The opposite effect was observed by Huett et al. (1997). Na, Ca, Mg concentrations in inoculated plants were lower than those of the controls, however, improved Mg concentration under severe water condition was observed. The same results were found by Monzon and Azcon (1996) for inoculated plants.

The support of symbiotic associations between these fungi and tropical gum tree may be suggested in application of inoculation for successfully improve mineral

nutrient uptake of seedlings in Sahelian zone agrosystem.

Conclusion

We may conclude that especially under water deficiency conditions, AMF enhances foliar essential nutrient uptake. This role is very much dependent on different species of arbuscular mycorrhiza. Accordingly, the symbiosis of *A. Senegal* with more efficient AM specie *G. fasciculatum* under such conditions can be very beneficial in water deficiency condition. The next step should be transposing this greenhouse experiment to the field.

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Abbreviations: **AMF**, Arbuscular mycorrhizal fungi; **FC**, field capacity; **MWD**, moderate water deficiency; **SWD**, severe water deficiency.

REFERENCES

- Audet P, Charest C (2006). Effects of AM colonization on « wild tobacco » plants grown in zinc-contaminated soil. *Mycorrhiza* 16:277-83.
- Augé RM (2001). Water relations drought and vesicular-arbuscular mycorrhizal symbiosis. *Mycorrhiza* 11:3-42.
- Bucher M (2007). Functional biology of plant phosphate uptake at root and mycorrhiza interfaces. *New Phytol.* 173:11-26.
- FD T90-019 (1984). Essais des eaux - Dosage du sodium et du potassium - Méthode par spectrométrie d'émission de flamme, Afnor.
- Floret C, Pontanier R, Serpantié G (1993). La jachère en Afrique Tropicale. *Man and Biosphere*, Unesco. Paris. 16:86.
- Garbaye J (2000). The role of ectomycorrhizal symbiosis in the resistance of forests to water stress. *Agriculture* 29:63-69.
- Giovannetti M, Mosse B (1980). An evaluation of techniques for measuring vesicular-arbuscular mycorrhizal infection in roots. *New Phytol.* 84:489-500.
- Huett DO, George AP, Slack JM, Morris SC (1997). Diagnostic leaf nutrient standards for low-chill peaches in subtropical Australia. *Aust. J. Exp. Agric.* 37:119-126.
- Li H, Smith SE, Holloway RE, Zhu Y, Smith FA (2006). Arbuscular mycorrhizal fungi contribute to phosphorus-fixing soil even in the absence of positive growth responses. *New Phytol.* 172:536-543.
- Liu A, Plenchette C, Hamel C (2007). Soil nutrient and water providers: how arbuscular mycorrhizal mycelia support plant performance in a resource limited world. In: Hamel C, Plenchette C (eds) *Mycorrhizae in Crop Production*. Haworth Food and Agricultural Products Press, Binghamton. NY. pp. 37-66.
- Miransari M, Bahrami HA, Rejali F, Malakouti MJ (2008). Using arbuscular mycorrhiza to reduce the stressful effects of soil compaction on wheat (*Triticumaestivum* L.) growth. *Soil Biol. Biochem.* 40:1197-1206.
- Miransari M, Bahrami HA, Rejali F, Malakouti MJ, Torabi H (2007). Using arbuscular mycorrhiza to reduce the stressful effects of soil compaction on corn (*Zea mays* L.) growth. *Soil. Biol. Biochem.* 39:2014-2026.
- Monzon A, Azcon R (1996). Relevance of mycorrhizal fungal origin and host plant genotype to inducing growth and nutrient uptake in *Medicago* species. *Agric. Ecosyst. Environ.* 60:9-15.
- Mortimer PE, Perez-Fernandez MA, Valentine AJ (2008). The role of arbuscular mycorrhizal colonization in the carbon and nitrogen economy of the tripartite symbiosis with nodulated *Phaseolus vulgaris*. *Soil. Biol. Biochem.* 40:1019-27.
- NF EN 25663 (1994). Qualité de l'eau - Dosage de l'azote Kjeldahl - Méthode après minéralisation au sélénium, Afnor.
- NF EN 6878 (2005). Qualité de l'eau - Dosage du phosphore - Méthode spectrométrique au molybdate d'ammonium, Afnor.
- Phillips JM, Hayman DS (1970). Improved procedures for cleaning roots and staining parasitic and vesicular arbuscular mycorrhizal fungi for rapid assessment of infection. *Trans. Br. Mycol. Soc.* 55:93-130.
- Qu L, Shinano T, Quoreshi AM, Tamai Y, Osaki M, Koike T (2004). Allocation of C14 carbon in two species of larch seedlings infected with ectomycorrhizal fungi. *Tree Physiol.* 24:69-76.
- Ruiz-Lozano JM (2003). Arbuscular mycorrhizal symbiosis and alleviation of osmotic stress. *New perspectives for molecular studies. Mycorrhiza* 13:309-317.
- Sánchez-Díaz M, Honrubia M (1994). Water relations and alleviation of drought stress in mycorrhizal plants In: S. Gianinazzi and H. Schüepp, Editors, *Impact of Arbuscular Mycorrhizas on sustainable agriculture and natural ecosystems*, Birkhäuser Verlag, Basel. Switzerland. pp. 167-178.
- Smith SE, Smith FA, Jakobsen I (2004). Functional diversity in arbuscular mycorrhizal (AM) symbioses: the contribution of the mycorrhizal P uptake pathway is not correlated with mycorrhizal responses in growth and total P uptake. *New Phytologist.* 162:511-524.
- Smith SS, Read DJ (2008). *Mycorrhizal Symbiosis*. Academic Press.
- Subramanian KS, Charest C (1997). Nutritional, growth, and reproductive responses of maize (*Zea mays* L.) to arbuscular mycorrhizal inoculation during and after drought stress at tasselling. *Mycorrhiza*, 7:25-32.