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Evaluation of soybean [*Glycine max* (L.)Merr.] genotypes for agronomic and quality traits in Kenya

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The interaction between genotypes and environments results in significant differences in the performance of genotypes when tested in various environments. Fifteen soybean [*Glycine max* (L.) Merr.] genotypes were evaluated for yield and protein content in Kenya aiming to quantifying and identifying high yielding genotypes for human food and livestock feed. The study was conducted in Eldoret (0° 35'N, 35°18'E), Lanet (0°18'S, 36° 09'E), Nakuru West (0° 33'S, 36° 0'E) and for two seasons at Njoro (0° 20'S, 35° 56'E). The results indicated significant ($p \le 0.01$) effects due to genotype, environment and genotype × environment interaction for days to flowering harvest maturity, plant height, number of pods per plant, number of nodes per plant, seed yield, oil and protein content. The mean seed yield was 1267.8 kg ha⁻¹. Genotype Nyala produced the highest yield across environments. The mean protein content ranged from 40.3% for genotype TGX 1740-2F (DPSB 19) to 35.2% for genotype 931/5/34 across the five test environments with the highest mean protein being recorded at Njoro and the lowest at Nakuru West. Genotype Nyala and SBH 7/1/1 may be recommended for production for their relatively high grain yield across the sites.

Key words: Soybean, yield, genotype, environment, protein.

INTRODUCTION

Soybean [*Glycine max* (L.) Merr.] is an important legume grown as source of vegetable oil and proteins as well as for feed and industrial uses (Wan et al., 2005). Soybean cultivation in Kenya is expected to gain popularity in the near future because of the increasing need for food and fodder (Mugendi et al., 2010). Livestock feed consumption accounts for over 90% of soybean utilization in Kenya and 60% of all livestock feed is compounded with soybean (Chianu et al., 2008.). Soybean improves soil fertility by fixing atmospheric nitrogen and some varieties fix 44 to 103 kg N ha⁻¹ annually (Sanginga et al., 2003). This can reduce the need for application of nitrogenous fertilizers that are detrimental to the environment (Zhang et al., 2003). Biological nitrogen fixation is environmental friendly and ideal for sustainable agriculture (Cheng, 2008). Soybean biomass is a source of feed, green manure and mulch (Chianu et al., 2008). Soybean farming is one of most cost effective ways smallholder farmers can maintain soil fertility and reap other benefits from subsequent crops (Osunde et al., 2003). Multi-environment trials are carried out to identify superior and stable soybean genotypes and to

*Corresponding author. E-mail: jamnjoros@gmail.com Author(s) agree that this article remain permanently open access under the terms of the <u>Creative Commons Attribution</u> <u>License 4.0 International License</u> understand the effects of genotype and environments on soybean performance. The interaction between genotypes and environments results in significant differences in the performance of genotypes when tested in various environments (Gauch and Zobel, 1997). The genotype x environment ($G \times E$) interaction plays a major role in the performance of any genotype and in the success of any breeding program for the development of genetic material, adapted to varying environments.

Quality aspects are affected by interactions between genotypes and the environment. Three aspects that affect soybean quality are seed appearance, protein and oil content and the chemical components of protein and oil (Liu et al., 1995). Primomo et al. (2002) investigated G \times E interaction for soybean fatty acids and found that genotype \times year interaction was significant for all fatty acids. Due to varying regional ecological conditions in Kenya, it is extremely important to select suitable cultivars for adaptability to specific as well as across environments.

Objectives of this study were to identify genotypes with high grain yield and related agronomic traits, protein and oil content for human food and feed consumption.

MATERIALS AND METHODS

Experimental sites

The study was conducted in Njoro (0° 20'S, 35° 56'E) for two seasons (Njoro I and Njoro II), Eldoret (0° 35'N, 35°18'E), Nakuru West (0° 33'S, 36° 0'E) and Lanet (0°18'S, 36° 09'E). Njoro (2185 m above sea level) is situated about 200 km West of Nairobi in Nakuru County and experiences an average daily minimum temperature of 9.5°C and maximum temperature of 24.2°C with an average precipitation of 1032 mm (average of 21 years 1992-2012 weather station number 9035021; Jaetzold et al., 2010). The soils are mollic andosols in eco-zone III (Jaetzold et al., 2010). Eldoret [2154 m above sea level (m.a.s.l.)] is located in Uasin Gishu County (FAO/UNESCO, 1994) and the soils are predominantly acidic (pH 4.7) rhodic ferrasols that are low in organic matter and deficient in nitrogen (N) and phosphorus (P). Lanet is 16 km South East of Nakuru town at an altitude of 1920 m.a.s.l. and experiences a bimodal rainfall pattern with an annual mean of 800 mm. The minimum and maximum temperatures that prevail in this site are 10 and 26°C, respectively (http://www.kalro.org). Nakuru West is elevated to an altitude of 2003 m.a.s.l. and is on the wind ward side overseeing Lake Nakuru.

Genotypes

Fifteen soybean genotypes obtained from Kenya Kenya Agricultural and Livestock Organization, Food Crops Research Centre (KARLO-Njoro) were used in this study. Eight of these were coded SBH (soybean hybridization) lines (SBH 10/5/6, SBH 1/12/9, SBH 7/1/1, SBH 4/4/4, SBH 10/2/3, SBH 4/6/6, SBH 6/6/6/2, SBH 3/8/4/1). Genotype EAI 3600, a stable genotype, was used as a check. Genotypes Nyala and Gazelle are medium maturing while genotype 931/5/34 gives high oil content. Genotype TGX1835-10E (DPSB 3) is resistant to Asian soybean rust disease [*Phakopsora pachyrhizi* (H. Sydow and Sydow)]. Genotypes TGX 1895-33F (DPSB-8) and TGX 1740-2F (DPSB-19) are promiscuous and have high biomass, trait that aids in improvement of poor soils and provision of feed to livestock.

Experimental procedure

The seedbed was prepared in order to become suitable for planting soybean. Planting was done on 13th June 2011 at Njoro in first season (Njoro I), 22nd June 2011 at Nakuru West, 8th July 2011 at Lanet, 14th June 2011 at Eldoret and on 15th July 2011 at Njoro in second season (Njoro II). The experiment was laid out in a randomised complete block design (RCBD) with three replicates. In all locations, the genotypes were planted in six row experimental units of 3 m x 2.7 m. Seeds were planted at a spacing of 45 cm x 10 cm and at an average depth of 5 cm. At planting time diammonium phosphate fertilizer (DAP) was applied to supply 22 kg of N ha-1 and 57.5 kg of P ha-1. A pre-emergent herbicide (Metribuzin) was applied at the rate of 360 g ha⁻¹ immediately after planting in order to control weeds. Weeding was done manually when weeds appeared after the waned effect of the herbicide. Foliar fungal diseases were controlled by applying Tebuconazole at the rate of 250 g ha⁻¹ weekly from flowering until the beginning of maturity (stage R7) (Fehr and Caviness, 1977). The crop was grown fully under rain fed conditions.

Data collection

Evaluation was done at vegetative and reproductive growth stages as described by Fehr and Caviness (1977). Days to 50% flowering was taken when there was at least one flower in 50% of all plants in the plot. Days to harvest maturity were observed when 95% of the pods had reached their mature pod colour (stage R8). At maturity, plant height, number of nodes plant⁻¹ and number of pods plant⁻¹ were determined from five plants of each genotype in the four middle rows. Mean seed count from twenty pods was observed as the number of seeds pod⁻¹. Seed yield was determined by weighing as the seed weight from four centre rows. The weight of 100 seeds was recorded by weighing a random sample of 100 seeds. Seed protein content was estimated using Near Infra red Refractometer (NIR) machine (Infratec TM 1241 Grain Analyzer ISW 3.20: Foss Analytical AB, Sweden).

Data analysis

Analysis of variance to estimate the effect of genotype, environment and $G \times E$ interaction on yield and protein content was done using the statistical analysis systems (SAS) general linear mode (GLM) procedure (SAS Institute, 2000 release 8.1). Means of genotypes were separated using least significant difference (LSD) test.

RESULTS

Agronomic performance of the soybean genotypes across the environments

The analysis of variance indicated that there was significant ($p \le 0.01$) effects due to genotype (G), environment (E) and genotype x environment interaction (Table 1) for all the traits studied. Variation among genotypes was observed in mean values of traits (Table 2). Days to 50% flowering ranged from 74 (DPSB 19) to 101 (DPSB 3). The contribution to the sum of squares

Source	df	50% flowering (days)	Harvest maturity (days)	Plant height (cm)	Pods plant ⁻¹ (No.)	Nodes plant ⁻¹ (No.)	Yield (Kg ha ⁻¹)	100-seed weight(g)	Oil content (%)	Protein content (%)
Environment (E)	4	1120.60 **	10411.87**	6458.81**	6203.01**	94.15**	4983331.16**	97.68**	134.68**	649.34**
Replicates R(E)	10	8.71	67.96	70.25	120.93	1.64	77842.17	1.62	0.81	6.88
Genotype(G)	14	714.72**	3774.52**	2996.69**	339.60**	59.85**	933486.56**	71.41**	26.98**	26.31**
G×E	56	30.24**	142.55**	439.69**	251.51**	8.64**	227898.57**	5.53**	0.97**	4.65**
Error	140	4.91	36.75	37.41	48.97	1.24	50436.56	1.81	0.39	3.02
C.V. %		2.64	3.72	8.72	17.83	8.29	17.71	9.85	3.23	4.61
R^2		0.95	0.95	0.94	0.86	0.90	0.86	0.87	0.94	0.88

Table 1. Mean squares for the examined traits of soybean genotypes evaluated at Njoro I, Njoro II, Eldoret, Nakuru West and Lanet, Kenya in 2011.

** Significant at $p \le 0.0I$, C.V.% - coefficient of variation, R²- coefficient of determination, df- degrees of freedom.

Table 2. Mean values of soybean agronomic traits of 15 genotypes evaluated at Njoro I, Njoro II, Eldoret, Nakuru West, and Lanet, Kenya in 2011.

Genotype	50% Flowering	Harvest maturity (days)	Plant height (cm)	Pods plant ⁻¹	Seeds Pod ⁻¹ (No.)	Nodes plant ⁻¹ (No.)	Grain yield (kg ha⁻¹)	100 Seed weight (g)	Oil content (%)	Protein content (%)
SBH 10/5/6	79.0 ^f	158.2 ^{fe}	87.4 ^b	35.3 ^{d-f}	2.4 ^{a-c}	15.7 ^{bc}	1303.5 ^{c-e}	13.3 ^{cd}	20.5 ^b	36.6 ^{gh}
SBH 1/12/9	79.8 ^{fe}	155.3 ^{f-h}	78.1 ^d	40.6 ^{bc}	2.4 ^{a-c}	14.4 ^{de}	1434.7 ^{bc}	12.6 ^{de}	19.6 ^{e-g}	37.0 ^{e-h}
SBH7/1/1	79.6 ^{fe}	162.5 ^e	87.9 ^b	39.4 ^{b-d}	2.4 ^{a-c}	16.8 ^a	1543.9 ^{ab}	12.9 ^{de}	20.1 ^{b-d}	36.8 ^{f-h}
Gazelle	89.3 ^c	168.5 ^d	81.6 ^{cd}	32.9 ^{ef}	1.9 ^f	14.8 ^d	1327.5 ^{с-е}	17.5 ^a	19.9 ^{c-f}	36.1 ^{hi}
SBH 4/4/4	79.2 ^{fe}	156.7 ^{fg}	85.7 ^{bc}	33.9 ^{ef}	2.4 ^{a-c}	15.1 ^{cd}	1367.4 ^{cd}	12.8 ^{de}	20.3 ^{bc}	36.9 ^{e-h}
SBH 10/2/3	83.1 ^d	154.2 ^{f-h}	60.5 ^{fg}	39.8 ^{b-d}	2.3 ^{b-d}	11.4 ^h	1124.9 ^f	14.5 ^b	19.7 ^{ef}	38.8 ^{bc}
SBH 4/6/6	82.6 ^d	153.8 ^{gh}	62.1 ^{ef}	37.3 ^{c-f}	2.3 ^{a-d}	11.1 ^h	1289.5 ^{c-e}	14.4 ^b	19.7 ^{d-f}	38.4 ^{bcd}
Nyala	84.3 ^d	174.5 [°]	60.7 ^{fg}	46.1 ^a	2.2 ^{de}	13.4 ^f	1600.9 ^a	17.6 ^a	19.5 ^{fg}	37.2 ^{d-h}
EAI 3600	83.2 ^d	155.8 ^{fg}	66.3 ^e	32.6 ^f	2.4 ^a	11.4 ^h	1375.2 ^{cd}	14.1 ^{bc}	20.0 ^{bc-e}	37.8 ^{c-g}
SBH 6/6/6/2	80.6 ^e	151.1 ^{ih}	57.8 ^{f-h}	43.2 ^{ab}	2.3 ^{a-d}	11.3 ^h	1197.7 ^{ef}	12.2 ^e	19.2 ^g	37.9 ^{c-f}
DPSB 8	91.7 ^b	185.1 ^b	95.8 ^a	37.5 ^{c-f}	2.4 ^{ab}	16.3 ^{ab}	1414.8 ^{bc}	12.3 ^e	17.9 ^h	38.0 ^{b-e}
DPSB 19	74.0 ^g	140.5j	59.6 ^{f-h}	37.9 ^{c-e}	2.3 ^{c-e}	12.5 ^g	807.9 ^g	10.7 ^f	17.0 ⁱ	40.3 ^a
931/5/34	93.1 ^b	188.9 ^b	55.3 ^h	47.1 ^a	2.4 ^{ab}	12.3 ^g	1323.3 ^{с-е}	16.7 ^a	21.9 ^a	35.2 ⁱ
DPSB 3	100.9 ^a	194.2 ^a	57.2 ^{gh}	39.1 ^{b-d}	2.1 ^e	13.8 ^{ef}	661.7 ^g	10.5 ^f	16.6 ⁱ	39.2 ^{ab}
SBH 3/8/4/1	80.6 ^e	147.3 ⁱ	56.8 ^{gh}	46.4 ^a	2.3 ^{a-d}	11.1 ^h	1243.5 ^{d-f}	12.9 ^{de}	19.2 ^g	38.9 ^{bc}
Mean	84.1	163.1	70.2	39.3	2.3	13.4	1267.8	13.7	19.4	37.7
C.V.%	2.6	3.7	8.7	17.8	8.8	8.3	17.7	9.8	3.2	1.3
LSD(0.05)	1.6	4.4	4.4	5.1	0.1	0.8	162.1	1.0	0.5	1.3

Means designated by the same letter within columns are not significantly different at p=0.05, CV%- coefficient of variation, df-degrees of freedom, LSD-least significance difference.

was 59% by genotypes, 26% by the environment while genotype by environment interaction (GEI) contributed 9% for the days to 50% flowering. The SBH lines were medium maturing with a range of 147-163 days while the TGX genotypes had the earliest maturing genotype ranging from 141 (DPSB19) to 194 days (DPSB 3). The check variety (EAI 3600) was medium in maturity. Genotype 931/5/34 had the highest number of pods plant and the shortest in height. Genotypes with pods plant¹ above the mean of 39.3 were SBH 1/12/19, SBH 7/1/1, SBH 10/2/3, Nyala, SBH 6/6/6/2, SBH 3/8/4/1 and 931/5/34. There was minimum variation on seeds pod⁻¹ since 30% of the genotypes (Gazelle, SBH 10/2/3, Nyala, DPSB 19, DPSB 3) had significantly different number of seeds pod⁻¹. Genotypes SBH 7/1/1 and DPSB 8 had the highest plant height and number of nodes plant¹ while genotypes SBH 3/8/4/1, SBH 4/6/6, SBH 6/6/6/2, EAI 3600 and SBH 10/2/3 had significantly lower number of nodes plant¹. Genotype Nyala had the highest mean grain yield across sites with the highest yield observed at Njoro in the second season (2397 kg ha⁻¹) while genotype DPSB 3 yielded the least (Table 2).

There were significant differences in seed weight among the genotypes. Genotypes Nyala had the highest seed, weight; however, this was not significantly different from Gazelle and 931/5/34. Only 40% of the genotypes (Gazelle, SBH 10/2/3, SBH 4/6/6, Nyala, EAI 3600, 931/5/34) observed seed weight above the mean (Table 2). Protein content was highest on genotype DPSB 19 while genotype 931/5/34 had the least. The contribution of environment sum of squares to yield, oil content, protein content, seed weight, maturity and pods plant⁻¹ was 37, 52, 70, 20, 38 and 48%, respectively while the genotype contributed 24, 37, 10, 51, 49, and 9%, respectively. The G × E interaction contributed 24, 5, 7, 16, 7 and 27% respectively. Based on their contribution, it is evident that protein and oil content are influenced by environment compared to other traits.

Performance of the genotypes in individual test environments

There was considerable variation in the performance of genotypes in individual test environments (Table 3). At Eldoret, genotype SBH 7/1/1 yielded highest while DPSB 19 yielded least (Table 3). Highest seed yield was observed at Lanet. Protein content was marginally higher at Njoro in the first season (Njoro I) compared to second season (Njoro II) (Table 3). The highest protein content (45%) was observed on genotype 931/5/ DPSB 19 at Njoro in the first season. Variability in the protein content among the genotypes was least in Nakuru west.

DISCUSSION

Significant genotype and $G \times E$ interactions for all the

traits studied suggested that genotypes responded differently to environments and there was G x E interaction, allowing for selection of genotypes for specific eco-zones. However, contribution to the total sum of squares differed depending on the trait. In a combined analysis of variance, Pfeiffer et al., (1995) observed that soybean genotypes differed significantly for seed yield and seed composition, however the genotype by environment interaction was not significant for seed yield but was significant for seed composition. The mean plant height range observed in the current study agrees satisfactorily with that observed by Aditya et al. (2011) while testing 31 soybean genotypes. A study by Karasu et al. (2009) indicated a similar range of mean plant height among the eight soybean genotypes tested in two locations for two years. The genotypes height differences in their study and the current study could be attributed to genetic and environmental influences.

Pods plant⁻¹, seeds pod⁻¹, number of nodes plant⁻¹ and seed weight are important components of seed yield. The results of the study on these traits are consistent with those of Aditya et al. (2011) and Karasu et al. (2009); however, Ojo (2003) recorded a higher range for pods plant⁻¹. The variations could be attributed to differences in genotypes and test environments. The oil content was significantly different among the genotypes. The percentage oil content was affected more by the environment and the genotype and less by G x E interaction effects. Protein in the current study varied among the genotypes and the variation was attributed to environment, genotypes and influence of environment on genotype. Protein and oil content are influenced by both genotype and environment cues (Clemente and Cahoon, 2009). The inverse relationship between oil and protein content exists, typically a 15% reduction in total oil content leads to a 2% increase in protein content (Clemente and Cahoon, 2009).

Environments and $G \times E$ interaction play a significant role in the performance of genotypes in a particular environment. There was significant $G \times E$ interaction for all traits studied which underscores the importance of selection for specific adoption to the given regions.

CONCLUSION AND RECOMMENDATIONS

The study has revealed that there are high yielding genotypes for different environments with high oil and protein content. Genotype SBH 7/1/1 and Gazelle, may be recommended for Eldoret and Lanet, respectively. DPSB 8 a late maturing genotype maybe recommended for Njoro in the first season. Genotype DPSB 19 and DPSB 3 had high mean protein content However, they had low grain yield and maybe useful in a breeding program to improve protein content. Among the sites tested seems to favor soybean production. Production targeting high protein content needs to be done in higher

Genotypes -	Seed yield (kg ha ⁻¹)							Protein content (%)					
	Eldoret	Lanet	Nakuru West	Njorol	Njoroll	Mean	Eldoret	Lanet	Nakuruwest	Njoro I	Njoroll	Mean	
SBH 10/5/6	1222.8 ^{a-c}	1748.8 ^{a-c}	912.3 ^{b-e}	1370.0 ^{b-d}	1263.0 ^{cd}	1303.5 ^{с-е}	35.5 ^{bc}	38.6 ^{a-d}	31.4 ^{a-c}	38.7 ^{d-f}	38.7 ^{e-f}	36.6 ^{gh}	
SBH 1/12/9	1204.3 ^{a-c}	1932.1 ^{a-c}	920.4 ^{b-e}	1495.1 ^{bc}	1621.6 ^{b-d}	1434.7 ^{bc}	37.5 ^{a-c}	36.9 ^{de}	31.3 ^{a-d}	38.6 ^{d-f}	40.8 ^{bc}	37.0 ^{e-h}	
SBH7/1/1	1492.0 ^a	1975.3 ^{a-c}	942.0 ^{b-d}	1542 ^b	1768.5 ^b	1543.9 ^{ab}	39.1 ^{ab}	37.7 ^{a-e}	31.2 ^{a-e}	37.8 ^{ef}	38.1 ^f	36.8 ^{f-h}	
Gazelle	1197.5 ^{a-c}	2179.0 ^a	959.2 ^{b-d}	827.8 ^{fg}	1474.1 ^{b-d}	1327.5 ^{с-е}	36.3 ^{bc}	36.1 ^e	30.8 ^{d-f}	37.9 ^{ef}	39.3 ^{c-f}	36.1 ^{hi}	
SBH 4/4/4	1217.3 ^{a-c}	1668.5 ^{bc}	918.5 ^{b-e}	1459.9 ^{b-d}	1572.8 ^{b-d}	1367.4 ^{cd}	39.1 ^{ab}	37.2 ^{b-e}	31.1 ^{a-e}	38.6 ^{d-f}	38.2 ^f	36.9 ^{e-h}	
SBH 10/2/3	812.3 ^{de}	1804.9 ^{a-c}	682.7 ^{ef}	997.5 ^{e-g}	1327.1 ^{b-d}	1124.9 ^f	38.0 ^{a-c}	40.0 ^{ab}	31.5 ^{ab}	43.3 ^{ab}	41.1 ^{cb}	38.8 ^{bc}	
SBH 4/6/6	943.8 ^{c-e}	1933.9 ^{a-c}	1070.4 ^{a-c}	1156.8 ^{d-f}	1342.6 ^{b-d}	1289.5 ^{с-е}	37.3 ^{a-c}	39.8 ^{a-c}	31.1 ^{a-e}	41.8 ^{bc}	41.9 ^{ab}	38.4 ^{bcd}	
Nyala	1165.4 ^{a-c}	1992.6 ^{ab}	1229.0 ^a	1237.7 ^{b-e}	2379.6 ^ª	1600.9 ^a	39.5 ^{ab}	37.1 ^{с-е}	31.0 ^{b-f}	39.9 ^{c-e}	38.4 ^{ef}	37.2 ^{d-h}	
EAI 3600	1125.9 ^{b-d}	2036.4 ^{ab}	1129.6 ^{ab}	1205.6 ^{с-е}	1378.4 ^{cd}	1375.2 ^{cd}	37.7 ^{a-c}	37.9 ^{a-e}	31.1 ^{a-e}	41.6 ^{b-d}	40.4 ^{b-f}	37.8 ^{c-g}	
SBH 6/6/6/2	1085.8 ^{b-d}	1538.9 ^{cd}	871.6 ^{c-e}	800 ^g	1692.0 ^{bc}	1197.7 ^{ef}	36.7 ^{bc}	38.6 ^{a-e}	31.4 ^{ef}	41.7 ^{b-d}	41.7 ^{ab}	37.9 ^{c-f}	
DPSB 8	927.2 ^{c-e}	1816.1 ^{a-c}	873.5 ^{с-е}	1919.1 ^a	1538.3 ^{c-d}	1414.8 ^{bc}	39.2 ^{ab}	39.1 ^{a-d}	31.4 ^{a-c}	40.6 ^{b-e}	39.9 ^{b-f}	38.0 ^{b-e}	
DPSB 19	675.3 ^e	1175.9 ^{ed}	768.5 ^{d-f}	246.9 ^h	1172.9 ^d	807.9 ^g	41.4 ^a	39.9 ^{a-c}	31.6 ^a	45.1 ^a	43.4 ^a	40.3 ^a	
931/5/34	1293.2 ^{ab}	1930.8 ^{a-c}	587.73 ^f	1569.1 ^b	1235.8 ^{cd}	1323.3 ^{с-е}	34.2 ^c	35.9 ^e	30.6 ^f	36.9 ^f	38.5 ^{ef}	35.2 ⁱ	
DPSB 3	1055.0 ^{b-d}	792.6 ^e	887.0 ^{bc-e}	827.8 ^{fg}	124.7 ^e	661.7 ⁹	41.6 ^a	40.5 ^a	30.9 ^{c-f}	42.1 ^{bc}	40.8 ^{b-d}	39.2 ^{ab}	
SBH 3/8/4/1	1184.0 ^{a-c}	1778.4 ^{a-c}	680.9 ^{ef}	937.7 ^{e-g}	1636.4 ^{b-d}	1243.5 ^{d-f}	39.2 ^{ab}	40.2 ^a	31.2 ^{a-e}	43.6 ^{ab}	40.7 ^{b-d}	38.9 ^{bc}	
Mean	1106.8	1153.62	895.6	1147.7	1435.2	1267.8	38.2	38.4	31.1	40.5	40.1	37.7	
C.V.%	18.0	15.30	16.7	17.3	19.5	17.7	7.1	4.3	1.0	4.5	3.1	1.3	
Lsd	333.9	448.8	248.9	331.8	469.2	162.1	4.5	2.8	0.5	3.0	2.1	1.3	

Table 3. Variation in yield and protein content of soybean genotypes grown at Njoro I, Njoro II, Eldoret, Nakuru West, and Lanet experimental sites, Kenya in 2011.

Means designated by the same letter within columns are not significantly different at p=0.05, CV %- coefficient of variation, df-degrees of freedom, LSD-least significance difference.

altitudes such as Njoro and Eldoret while production targeting high oil content should focus on lower altitude areas such as Nakuru west. The low GxE interaction of the genotypes for oil and protein content implies that there is still need to develop genotypes with high oil and protein content. Genotype Nyala and SBH 7/1/1 maybe recommended for production across studied environments due to their relatively high grain vield. Further evaluation in low and medium altitudes with more diverse soybean genotypes, especially promiscuous ones the is recommended.

Conflict of Interest

The authors have not declared any conflict of interest.

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