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Trichoderma-bean interaction: Defense enzymes activity and endophytism

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Biotic inducing agents such as *Trichoderma* spp. are an alternative for the induction of resistance by activating defense mechanisms of plants. This work aimed to study the interaction between bean and *Trichoderma* spp. evaluating the effects of the induction of resistance in bean through the defense enzymes activity and ability to endofitism. The experimental was design in randomized blocks, with four replications, being 21 isolates of *Trichoderma* spp. and control. The isolates were inoculated at planting, with adding 0.8 ml of the suspension 1×10^8 conidia/ml in groove, in sterilized soil. After 30 days, leaf and roots samples were taken to determine the enzyme activity. To verify the endophytic colonization capacity of isolated, bean root fragments were sanitized and placed in Petri dishes containing PDA medium. Data were submitted to analyzes of variance and compared by Scott-Knott test ($p < 0.05$). The enzymatic activity of peroxidase, polyphenoloxidase and phenylalanine ammonia-lyase was not significant, but the β -1-3-glucanase activity in leaf tissue was suppressed by isolates T11, TM1, TLB15 (*Trichoderma virens*), T12, TLB3, TLB4, TLB12, TOD1, TOD3 (*Trichoderma harzianum*), TLB6 (*Trichoderma asperellum*), TLB14, TLB17 (*Trichoderma koningiopsis*) and TOD2B (*Trichoderma longibrachiatum*). The isolates TM4, TLB9, TLB15 (*T. virens*), T12, T14, TLB2, TOD1 (*T. harzianum*) and TLB17 (*T. koningiopsis*), were able to colonize endophytically the bean roots. *Trichoderma* has endophytic capacity and interfere in β -1-3-glucanase activity.

Key words: Induced resistance, *Phaseolus vulgaris* L., *Trichoderma* spp.

INTRODUCTION

The common bean (*Phaseolus vulgaris* L.) is exposed to factors that causes reduction on productivity and final quality of grain. Brazil is one of the major producer and

consumer of this legume, with national production of 3,185,400,000 tons in the 2014/1015 harvest (Conab, 2015). However, there are several phytosanitary

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Table 1. Name, specie of the isolated used on trials.

Isolated	Specie
TI1	<i>Trichoderma virens</i>
TI2	<i>Trichoderma harzianum</i>
TI3	<i>Trichoderma virens</i>
TI4	<i>Trichoderma harzianum</i>
TM1	<i>Trichoderma virens</i>
TM2	<i>Trichoderma virens</i>
TM3	<i>Trichoderma virens</i>
TM4	<i>Trichoderma virens</i>
TLB2	<i>Trichoderma harzianum</i>
TLB3	<i>Trichoderma harzianum</i>
TLB4	<i>Trichoderma harzianum</i>
TLB6	<i>Trichoderma asperellum</i>
TLB9	<i>Trichoderma virens</i>
TLB12	<i>Trichoderma harzianum</i>
TLB14	<i>Trichoderma koningiopsis</i>
TLB15	<i>Trichoderma virens</i>
TLB17	<i>Trichoderma koningiopsis</i>
TOD1	<i>Trichoderma harzianum</i>
TOD2A	<i>Trichoderma longibrachiatum</i>
TOD2B	<i>Trichoderma longibrachiatum</i>
TOD3	<i>Trichoderma harzianum</i>

problems, that have great destructive potential, causing enormous losses in production of the country.

The *Trichoderma* is a fungus naturally found in different types of soils, living saprophyte and parasitizing other fungi, behaving as an antagonist to various phytopathogens, being the most searched fungus nowadays, once they are able to act as biological control agents, growth promoters and plant disease resistance inducers (Louzada et al., 2009).

Biological control is considered to be one of the method that may occur simultaneously throughout the antagonist life cycle, whose capacity can induce the synthesis of substances with antagonistic action by producing secondary metabolites which inhibit or stimulate the development of a microorganism or plant. In addition to biological control, there may be resistance induction, in which the plant activates defense mechanisms, signaled by elicitors of biotic and abiotic origin (Papavizas, 1985; Ridout et al., 1988).

The interaction between plant and fungus may have endophytic relations, establishing symbiotic relations still not well understood, but of great importance for plants (Azevedo, 1998). In interactions, the microorganisms produce or induce the production of secondary metabolites, which can provide many benefits, such as inducing resistance by the activation of defense mechanisms present at low levels in plant (Hammerschmidt and Dann, 1997; Schwan-Estrada et al., 2000).

Thus, the research aimed to study the interaction of different isolates of *Trichoderma* spp. in bean plants, by the effect on enzymes activity related to plant defense and endophytic capacity.

MATERIALS AND METHODS

The experiment was conducted in laboratory and greenhouse. Isolates of *Trichoderma* spp., from native forest soils and of agricultural property in system of organic production, were isolated by the bait method (Ethur et al., 2005) and identified in species level by the sequencing region method internal transcribed spacer (ITS) and part of gene elongation factor (EF) gene (Table 1).

To prepare the *Trichoderma* spp. inoculum, potato dextrose agar (PDA) discs having mycelium of each isolates, were transferred to erlenmeyers containing rice (20 g of rice to 25 ml of water and autoclaved for 20 min at 120°C), then, were incubated at 25 ± 2°C at photoperiod of 12 h, according to Silva et al. (2011) with adaptations.

After seven days, conidia suspension was prepared with 50 mL of sterile and distilled water, added to the Erlenmeyer to make rice washing. The suspension was filtered in gauze and the concentration was adjusted to 1×10⁶ conidia mL⁻¹, with the aid of a Neubauer chamber.

The experiment was conducted in plastic pots for up to three liters, which contained soil, sand and organic compound in a proportion of 3: 2: 1, autoclaved for 25 min at 120°C, repeating the operation in 24 h.

Seed were treated by the suspension of *Trichoderma* spp. isolates at planting, by application of 0.8 ml of the adjusted suspension (Broetto, 2013). In each pot were seeded two bean seeds of the cultivar IAPAR 81 (Iapar, 2015).

The experimental design was in randomized blocks, being 22 treatments with four replications (21 *Trichoderma* spp isolates and a control (without *Trichoderma* spp. inoculation)), deployed in 88 experimental plots.

For carrying out the biochemical analysis, 30 days after the emergence of bean the samples were collected samples from leaf and root tissue, which were weighed and packed in aluminum envelopes, frozen and kept in a freezer at -20°C until the moment of analysis.

Samples from leaves and roots (0.5 g) were macerated mechanically in 4 mL of sodium acetate buffer 100 mM (pH 5.0), being added 0.04 g of polyvinylpyrrolidone. The mixture was centrifuged at 20.000 g for 25 minutes at 4°C. The supernate was taken as the enzyme extract and frozen at -20°C for determination of peroxidase (POD), polyphenoloxidase (PPO), phenylalanine ammonia-lyase (PAL) and β-1,3-glucanase (β-GAZE) activities (Lusso and Pascholati, 1999).

The POD activity was determined by direct spectrophotometric method, by the measurement of the conversion of guaiacol in tetraguaiacol to 30°C, with spectrophotometric reading at 470 nm, for a period of 2 minutes, with optical density recorded every 15 s (Hammerschmidt et al., 1982). The reaction mixture contained 100 μL of the enzyme preparation and 900 μL of solution (12.5 mL of guaiacol and 310 μL of hydrogen peroxide in 87.5 ml of sodium phosphate buffer 0.01 M (pH 6.8)). The activity was expressed by the variation between the values in the linear increase range (E.U. (enzyme unit) min⁻¹ g⁻¹ fresh weight).

The PPO activity was determined using the methodology of Duangmal and Apenten (1999), adapted by Kuhn et al. (2010). The assay measured the catechol oxidation converted into quinone, reaction measured by the enzyme PPO. The substrate was composed by catechol in a concentration of 0.002 M, dissolved in sodium phosphate buffer 0.01 M (pH 6.8). The reaction was developed by mixing 900 μL of substrate and 100 μL of enzymatic

preparation, with readings in spectrophotometer at 420 nm, for a period of one minute, with optical density recorded every 15 s. The activity was determined by the variation between the values in the linear increase range E.U. (enzyme unit) $\text{min}^{-1} \text{g}^{-1}$ fresh weight.

The PAL activity was determined by quantitation of trans-cinnamic acid released from the substrate phenylalanine; the methodology is described by Umesha (2006), in which 50 μL of the enzyme preparation were added to 450 μL of Tris-HCl 0.025 M buffer (pH 8,8) and 500 μL of a solution L-phenylalanine 0.05 M, incubated at 40°C for 2 h. The reaction was stopped by adding 60 μL of HCl 5 M, with read in a spectrophotometer at 290 nm. The PAL activity constitutes of the difference between the absorbance of the mixture and of the control (100 μL of protein extract and 900 μL of Tris-HCl 0.025 M buffer (pH 8.8)), which was guinea fowl in the standard curve to trans-cinnamic acid ($y = 0.0095x + 0.0255$, where y is the absorbance at 290 nm and x is the concentration of trans-cinnamic acid (μg)), expressed in μg trans-cinnamic acid $\text{h}^{-1} \text{g}^{-1}$ fresh weight.

The β -GAZE activity was determined by colorimetric measurement of reducing sugars released from the laminarin (Vogelsang and Barz, 1993). The reaction involves 100 μL of the enzyme preparation, 50 μL of sodium phosphate buffer 0.01 M (pH 6.0) and 150 μL of laminarin (2 mg mL^{-1}), which was developed at 40 ° C for one hour. In control, laminarin was added after the incubation. The sugars formed were quantified by Lever Method (1972), which consists of extracting an aliquot part of 30 μL of the previous solution and adding to this 1.5 mL of hydrazine solution of the p-hydroxybenzoic acid 0.5% (PAHBAH). The mixture was maintained at 100°C for 10 min, cooled on ice for 10 min, and the absorbance determined at 410 nm. The β -1,3-glucanase activity was the difference between the absorbance of the mixture containing the sample and control, which was painted in standard for glucose curve ($y = 0.002x + 0.0046$, where y is the absorbance and x the concentration of reducing sugars (μg)) and expressed as glucose equivalent $\mu\text{g}^{-1} \text{mL}^{-1} \text{h}^{-1} \text{g}^{-1}$ fresh weight.

After collecting the roots samples for biochemical analysis, a portion of 10 cm of root from each plant was washed to evaluate the ability of *Trichoderma* spp. isolates, about its endophytic colonization capacity. This was performed by the indirect method of isolation, using root fragments of 1.5 cm, superficially disinfected with 70% alcohol for one minute, immersed in 3% sodium hypochlorite for one minute and washed three times in distilled autoclaved water. Then, three fragments were transferred into Petri plate, containing PDA médium, in three repetitions. The petri dishes were incubated in BOD at 25 ± 2 ° C with a photoperiod of 12 hours. The evaluation was made by viewing the *Trichoderma* spp. growth, from the root.

Once tabulated, the data were submitted to analysis of variance, and the averages compared by Scott-Knott, at 5% probability, with the assistance of statistical software version 5.3 Sisvar (Ferreira, 2010).

RESULTS AND DISCUSSION

Despite the importance of *Trichoderma* sp. and research for resistance induction, just a little is understood about the relationship of microorganism-plant interaction of this fungus in the bean, by the fact of the paucity of studies involving the mechanisms by which *Trichoderma* spp. interacts, activating the biochemical defense systems of bean plants. Several hypotheses are presented, but few of them have been cogent, which are evaluated by chemical and cytological searches from plant tissues presenting these fungal agents (Hanson and Howell,

2004).

According to Table 2, the results of the enzymatic activity of POD, PPO and PAL in the leaves and roots showed no statistical difference between isolated of *Trichoderma* spp., but, the β -GAZE was reduced depending on the isolated, presenting statistical difference in bean leaves.

The POD activity in bean, even with no statistical difference between the isolated, can observe a tendency of suppression by the isolated TM1, TM4, TLB15 (*T. virens*), TOD2A (*T. longibrachiatum*) and TOD3 (*T. harzianum*) on the expression of this enzyme activity in leaf tissue of bean, as compared to control.

Similarly, the isolated TI1, TLB15 (*T. virens*), TOD2B (*T. longibrachiatum*) and TLB6 (*T. asperellum*), suppressed the enzymatic activity in root tissue. However, the isolated TM3 (*T. virens*) and TI2, TLB4 (*T. harzianum*), showed a tendency to increase the POD enzyme in bean plants, compared to the control.

Similar results for POD were obtained by Silva et al. (2011), that observed the effect of 60 isolated of *Trichoderma* spp. on growth promoting and systemic resistance induction to anthracnose in cucumber plants. They found no significant difference regarding the activation of the PPO enzyme in the plant between the 7th and 14th day after treated with *Trichoderma* spp., as compared to control.

However, Yedidia et al. (1999), working with hydroponic cucumber plants inoculated with *Trichoderma*, observed an increase in the POD enzyme after the time interval until seven days. However there was no increase in activity at 7 and 14 days after treatment, while the untreated plants showed a gradual increase over time, it was observed increase in plant growth, being confirmed, for the first time, the direct impact of this fungus in the interaction plant-fungus.

According to Yedidia et al. (1999), the increase in POD enzyme in leaves observed in their study, may be due to either a systemic defense response, as the presence of *Trichoderma* in the rhizosphere, probably decrease of the enzymatic activity could be temporarily related to the root colonization by different isolated of *Trichoderma* spp.

A similar result was obtained by Dildey et al. (2013), which have observed no POD and PPO activity induction in bean plants against the pathogen *Macrophomina phaseolina*, in inoculation of seeds with different isolates of *T. harzianum*. However, even it does not demonstrate significant results for enzymes here, there was a tendency of such, on one of the different isolated of *Trichoderma* compared with the treatment control. From the results, the POD activity is not necessarily involved in this process, but further defense mechanisms such as chitinases, β -GAZE among others, may be acting in the induction of resistance of bean plants.

For the PPO activity, the results of this research showed no statistical difference, however, the isolated TI1, TI3, TLB15, TM4 (*T. virens*), TOD2A, TOD2B (*T.*

Table 2. Peroxidase activity (POD - unit of enzyme⁻¹ min⁻¹ g⁻¹ fresh tissue), polyphenoloxidase (PPO - unit of enzyme⁻¹ min⁻¹ g⁻¹ fresh tissue), phenylalanine ammonia-lyase (PAL - µg⁻¹ trans-cinnamic acid h⁻¹ g⁻¹ fresh tissue) and β-1,3 glucanase (β-GAZE - µg⁻¹ h⁻¹ g⁻¹ fresh tissue) in roots and leaves of bean in the treating seed with *Trichoderma* spp. isolated, 30 days after germination.

Isolated	Peroxidase activity							
	Leaf				Root			
	POD	PPO	PAL	β-GAZE	POD	PPO	PAL	β-GAZE
TI1	1066.22 ^a	438.72 ^a	31.48 ^a	869.70 ^a	776.85 ^a	655.15 ^a	51.54 ^a	1876.52 ^a
TI2	973.85 ^a	593.90 ^a	81.43 ^a	1318.42 ^a	1120.42 ^a	566.82 ^a	44.53 ^a	1678.07 ^a
TI3	779.27 ^a	506.65 ^a	21.25 ^a	1546.45b	924.02 ^a	848.20 ^a	58.63 ^a	1592.40 ^a
TI4	850.67 ^a	516.97 ^a	30.03 ^a	1633.8b	864.85 ^a	569.87 ^a	65.83 ^a	1630.20 ^a
TM1	747.05 ^a	629.37 ^a	43.11 ^a	1234.85 ^a	879.32 ^a	643.27 ^a	60.32 ^a	1640.30 ^a
TM2	827.52 ^a	601.92 ^a	97.78 ^a	1527.75b	1018.20 ^a	852.25 ^a	42.87 ^a	1883.07 ^a
TM3	1065.47 ^a	581.02 ^a	162.43 ^a	1873.90b	1226.87 ^a	741.62 ^a	50.75 ^a	1864.10 ^a
TM4	689.47 ^a	537.27 ^a	36.40 ^a	1698.47b	819.25 ^a	500.25 ^a	39.15 ^a	1356.00 ^a
TLB2	911.25 ^a	512.05 ^a	94.99 ^a	2064.77b	1080.15 ^a	655.22 ^a	49.71 ^a	1602.47 ^a
TLB3	1069.52 ^a	601.85 ^a	71.69 ^a	1302.17 ^a	1040.37 ^a	566.27 ^a	110.46 ^a	1833.70 ^a
TLB4	1045.30 ^a	590.30 ^a	38.95 ^a	1415.10 ^a	1121.90 ^a	775.07 ^a	48.95 ^a	1704.45 ^a
TLB6	1025.45 ^a	598.65 ^a	57.22 ^a	1395.10 ^a	746.45 ^a	715.12 ^a	67.54 ^a	1907.62 ^a
TLB9	973.52 ^a	662.97 ^a	64.39 ^a	2045.00b	922.75 ^a	673.80 ^a	92.21 ^a	2050.42 ^a
TLB12	944.85 ^a	549.72 ^a	32.47 ^a	1033.17 ^a	878.35 ^a	671.85 ^a	71.14 ^a	2184.27 ^a
TLB14	911.07 ^a	583.15 ^a	48.25 ^a	1322.77 ^a	988.47 ^a	605.30 ^a	38.46 ^a	1791.10 ^a
TLB15	800.97 ^a	418.45 ^a	71.75 ^a	1308.60 ^a	641.70 ^a	546.20 ^a	58.21 ^a	1803.82 ^a
TLB17	1049.35 ^a	613.80 ^a	70.71 ^a	1263.40 ^a	854.45 ^a	700.82 ^a	54.02 ^a	2118.07 ^a
TOD1	1043.97 ^a	581.95 ^a	100.60 ^a	1366.50 ^a	982.92 ^a	522.27 ^a	56.49 ^a	1771.67 ^a
TOD2A	922.87 ^a	439.45 ^a	41.02 ^a	1831.80b	933.87 ^a	595.92 ^a	70.58 ^a	1960.02 ^a
TOD2B	733.80 ^a	427.17 ^a	17.91 ^a	1187.20 ^a	714.12 ^a	632.15 ^a	71.45 ^a	2093.87 ^a
TOD3	827.20 ^a	519.42 ^a	77.40 ^a	1126.05 ^a	828.82 ^a	553.22 ^a	66.46 ^a	2252.90 ^a
Controle	1014.50 ^a	638.12 ^a	111.30 ^a	1747.75b	890.17 ^a	644.25 ^a	64.07 ^a	2195.00 ^a
(CV%)	30.50	32.80	125.40	32.90	37.70	30.50	47.18	26.80
DMS	747.52	481.48	212.34	1274.71	921.97	523.91	75.90	1319.19

TI1, TI3, TM1, TM2, TM3, TM4, TLB9 and TLB15 (*T. virens*); TI2, TI4, TLB2, TLB3, TLB4, TLB12, TOD1, TOD3 (*T. harzianum*); TLB6 (*T. asperellum*); TLB14, TLB17 (*T. koningiopsis*); TOD2A, TOD2B (*T. longibrachiatum*). * Means followed by the same letter in the column do not differ by the Scott-Knott test at 5% significance. Control: soil without *Trichoderma* spp. Transformed data by $\sqrt{x+1.0}$.

longibrachiatum) and TI4, TLB2 (*T. harzianum*), had a suppression tendency of PPO enzymatic activity, compared with the control, for the leaf tissue.

For the PPO activity in the root tissues of bean plants, they have shown an increasing trend for the isolated TI3, TM2, TM3 (*T. virens*) and TLB4 (*T. harzianum*), however it can be seen that the isolated TLB15, TM4 (*T. virens*) and TI2, TI4, TLB3, TOD1, TOD3 (*T. harzianum*), suppressed this enzyme activity.

Pereira et al. (2008), working with chitosan extracted from *Trichoderma* spp. and *Rhizopus* sp., obtained a protective effect on cocoa plants, by noting that the application of biotic inducers are able to promote increased in POD and PPO enzymes activity, in addition to reducing Verticillium-wilt.

According to Pinol and Palazón (1996), in some cases, the polyphenols metabolism may act as antioxidant, leading to the inactivation of fenolases enzyme production. In another aspect, the biotic biosynthesis of

secondary metabolites is attached the plant genetic constitution determining the activation of genes that synthesize the enzymes.

The PAL activity, showed no significant difference in bean plants. But, for leaf tissue, it can be noticed a sharp increase in the enzymatic activity of isolated TM3 (*T. virens*) compared to the control. On the other hand, all isolated present a suppression tendency for this enzyme, being this trend more evident in isolated TI1, TI3, TM4 (*T. virens*), TI4, TLB4, TLB12 (*T. harzianum*) and TOD2B (*T. longibrachiatum*), with no statistical difference.

In expression of this enzyme in the root system, it is observed a increase tendency for isolated TLB3 (*T. harzianum*) and TLB9 (*T. virens*), however, the isolated TI1, TM2, TM3, TM4 (*T. virens*), TI2, TLB2, TLB4 (*T. harzianum*), TLB6 (*T. asperellum*) and TLB14 (*T. koningiopsis*), showed suppression in PAL enzymatic activity and no statistical difference.

Karthikeyan et al. (2006), evaluated the induction of

Table 3. Presence (+) and absence (-) of endophytic colonization in bean roots by *Trichoderma* spp. isolated, 30 days after germination.

Colonization in bean roots		
Isolated	Specie	Colonization
TI1	<i>T. virens</i>	-
TI2	<i>T. harzianum</i>	+
TI3	<i>T. virens</i>	-
TI4	<i>T. harzianum</i>	+
TM1	<i>T. virens</i>	-
TM2	<i>T. virens</i>	-
TM3	<i>T. virens</i>	-
TM4	<i>T. virens</i>	+
TLB2	<i>T. harzianum</i>	+
TLB3	<i>T. harzianum</i>	-
TLB4	<i>T. harzianum</i>	-
TLB6	<i>T. asperellum</i>	-
TLB9	<i>T. virens</i>	+
TLB12	<i>T. harzianum</i>	-
TLB14	<i>T. koningiopsis</i>	-
TLB15	<i>T. virens</i>	+
TLB17	<i>T. koningiopsis</i>	+
TOD1	<i>T. harzianum</i>	+
TOD2A	<i>T. longibrachiatum</i>	-
TOD2B	<i>T. longibrachiatum</i>	-
TOD3	<i>T. harzianum</i>	-
Control*		-

*Soil without *Trichoderma* spp.

phenol compounds, the defense enzymes involved in phenylpropanoid pathway and the protein-PR accumulation against *Ganoderma* in reply to the application of *Pseudomonas fluorescens*, *T. viride* and *T. harzianum* combined with chitin. The authors observed a high increase in PAL activity in all treated palms following application of *P. fluorescens*, *T. viride* and chitin. The highest level was reached 3 days after treatment and, then, declined slowly.

According to De Meyer et al. (1999) the rhizosphere colonization by *P. aeruginosa*, produced PAL activity in bean roots and, consequently, increased the salicylic acid level in the leaves.

As stated by Campos et al. (2003), a complex response which involves production of phytoalexins and isoflavonoids may make a different role in the related response for the enzyme PAL induction. In addition of there being a complex response, involving phytoalexins and isoflavonoids production, it may play a different role in the response related to induction of that enzyme.

On the other hand, Nicholson and Hammerschmidt (1992), reported that the increase in PAL activity might be directly involved with a biochemical marker for induced resistance, once this enzyme is considered essential for

the phenols synthesis, being important for resistance, enabling occurrence of a specific interaction between plant and inducer microorganism.

For β -GAZE activity, there was significant difference for the results of leaf tissue treated with the isolated TI1, TM1, TLB15 (*T. virens*), TI2, TLB3, TLB4, TLB12, TOD1 and TOD3 (*T. harzianum*). The isolated TLB6 (*T. asperellum*), TLB14, TLB17 (*T. koningiopsis*), TOD2B (*T. longibrachiatum*) suppressed the this enzyme activity. In tissue root system there was no difference, but was observed a more evident suppression trend in the isolated TM4 (*T. virens*).

According to Saksirirat et al. (2009), by working with fifteen isolated of *Trichoderma* spp. to control *Xanthomonas campestris* pv. *vesicatoria* there was a decrease of 69,32% of the symptoms, showed that four isolated of species *T. harzianum* e *T. asperellum*, induced activity of β -GAZE and chitinase in the leaves, showing that these species are promising for resistance induction in tomato plants. The same authors, They report that these enzymes are known to play an important role in the degradation of fungus cell wall. Besides that, are related to pathogenesis-related proteins (PR proteins) in several plants, including beans (Van Loon and Van Stien, 1999).

As stated by Salzman et al. (1998), these induced proteins are not always present during the infection process, they can be found in healthy tissue of certain plant part, as leaves or roots, or at specific developmental stages.

According to Akrami et al. (2011), several factors may interfere in the efficiency of biotic inducing agents among them the abiotics, mainly temperature, humidity and soil pH, which are considered crucial on inducer efficiency, ensuring a successful performance of *Trichoderma* spp. isolated.

Among the 21 isolates of *Trichoderma* spp. evaluated for the colonization of bean plants roots, it was verified that 8 isolated feature the endophytism capacity in roots (Table 3), The results show that these isolates can be considered endophytic microorganisms

These data confirm those of Benítez et al. (2004), which found that isolated of *Trichoderma* spp. are able of colonizing roots, producing secondary compounds that confer stimulate to plant growth by, as well as, induce resistance, by activation of plant defense mechanism. The same has been verified by Pedro (2012), that working with *Trichoderma* spp. isolated, in growth stimulating and anthracnose resistance induction in bean plants, showed that seven isolated of *Trichoderma* spp. evaluated were able to colonize the seedlings roots, being possible to check their growth on petri dishes, from the roots.

The gender *Trichoderma* may be associated with the roots of several plants of several species. Its penetration and colonization, associated with its antibiosis capacity, enhances an hydrolytic enzymatic arsenal, degrader of

the plant cell wall. This input on epidermal layers of the roots confers a higher growth, improving nutrient absorption, and, thereby, increasing plant productivity (Harman et al., 2004).

According to the results of the present work, the reduction of enzymatic activity verified in bean plants inoculated with *Trichoderma* spp., is not very well clarified, it is believed that the *Trichoderma* spp., stimulates the plant to the equilibrium of its enzymatic arsenal, for the sake of its own benefit, so that, the plant will not endanger the symbiotic relationship between its interaction with the *Trichoderma* spp., because in case of the plant be stimulated to produce defense enzymes, *Trichoderma*, probably, would be hampered by the degradation of its structures and its cell death would be occurred.

Lucon (2009), verify that, in the absence of phytopathogens, the action of the mechanisms may be related to the production of hormones or growing factors, increase in the availability and absorption of nutrients or efficiency in the use of some nutrients by the plant.

These same *Trichoderma* spp. isolated, tested in this study, were used for Broetto (2013), which verified the effectiveness of the isolated TLB2, TLB3, TLB12 and TLB17 in bean against *M. phaseolina*. Also, stood out isolated TI1, TM3, TLB3, TLB12 and TLB17 as good growth promoters of the bean plants, whereas, the isolated TLB12 presented highlighted grain yield.

Several studies have been conducted to extend the knowledge of the action mechanisms by the biocontrol in diverse cultures. It is believed that, to the extent that the potential of these mechanisms will getting to know, in interactions with plants and microorganisms, lead to promising expectations for use in agriculture, as well as contribute towards the environment. preservation.

Conclusion

Isolated of *Trichoderma* TI1, TM1, TLB15 (*T. virens*), TI2, TLB3, TLB4, TLB12, TOD1, TOD3 (*T. harzianum*), TLB6 (*T. asperellum*), TLB14, TLB17 (*T. koningiopsis*) and TOD2B (*T. longibrachiatum*), suppressed the β -1,3-glucanase activity in leaf tissue. Neither of the isolated interfered in the activity of peroxidase, polyphenoloxidase and phenylalanine-ammonia-lyase. Isolated of *Trichoderma* spp., TM4, TLB9, TLB15 (*T. virens*), TI2, TI4, TLB2, TOD1 (*T. harzianum*) and TLB17 (*T. koningiopsis*), colonized the bean root in endophytic form.

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