academicJournals

Vol. 11(34), pp. 3203 -3209, 25 August, 2016 DOI: 10.5897/AJAR2016.11331 Article Number: 766A0C460186 ISSN 1991-637X Copyright ©2016 Author(s) retain the copyright of this article http://www.academicjournals.org/AJAR

African Journal of Agricultural Research

Full Length Research Paper

Estimation of the flower buttons per inflorescences of grapevine (*Vitis vinifera* L.) by image auto-assessment processing

Radhouane Benmehaia^{1,2*}, Derdour Khedidja³ and Mohamed El Moncef Bentchikou¹

Received 18 June, 2016; Accepted 9 August, 2016

The aim of this research is to develop a reliable tool by a special method, combining image processing based on watershed algorithm, and a predictive model to estimate automatically the flowers number per inflorescence. Eighty images of *Vitis vinifera* L. inflorescence (the Cardinal cultivar) were processed. Watershed algorithm was used for the image processing and this was followed by statistical analysis that provides robust predictive estimation of the flower button number. The results show a robust estimation, compared to manual flowers counting, with strong correlation. The developed algorithm shows that the watershed algorithm was able to provide an automatic assessment of the flower button number in the inflorescence. The method used is more robust and provides a more significant level compared with recent studies. In the applied research in viticulture, it is crucial to improve knowledge of yield forecasting and to study the fruit set rates estimation. The technique is used to determine, with a higher significance level, the fruitiness rate of grapevine at the early stage of flowering.

Key words: Flower button, fruit set rate, grapevine, image processing, watershed.

INTRODUCTION

Determining fruit set rates requires a flower button counting at stage H (separate flower buttons) of Baggiolini code (1952) in the development cycle of the vine, and at another stage (J). The first attempt of counting procedure

is done smoothly and cautiously by manual manipulation of a lot of inflorescences of vines. This monotonous procedure has some technical and spatiotemporal constraints. The manual flower counting is very difficult,

*Corresponding author. E-mail: benmehaiarad@univ-msila.dz.

Author(s) agree that this article remain permanently open access under the terms of the <u>Creative Commons Attribution</u> License 4.0 International License

¹Laboratoire de Développement et Valorisation des Ressources Phytogénétiques, Université de Constantine 1, B. P. 325 Route Ain El Bey, Constantine 25017, Algérie.

²Département des Sciences de la Nature et de la Vie, Faculté des Sciences, Université de Msila, B. P. 166 Ichbelia, 28000 M'sila, Algérie.

³Laboratoire d'Automatique et Productique, Département de Génie Industrielle, Université de Batna, 05 Avenue Chahid Boukhlouf, 05000 Batna, Algérie.

and deals with many problems having influence on the requested research results. The main problem is the large number of flower buttons (several hundreds or thousands) that can be grouped into clusters (cluster compactness). This may cause a loss in flower buttons during manipulation. Other disadvantages are the possible severe natural conditions of experimentation and time constraints.

The research of Bessis (1960) suggests a method for determination of the flower number. It is based on a linear regression between the cluster length and its richness in flower. He affirmed a positive relationship. This is actually the common method for flower assessment.

More recently, counting objects by using computer simulation has been of great research interest involving important methodological problems of image processing. Researchers have developed various techniques. Formerly, Girard et al. (2009) have developed a method of counting flexible oblong objects that may overlap. This method uses a combination of image processing morphological and statistical filtering.

Moreover, Vallotton and Thomas (2008) have introduced a system based on algorithms of image processing for counting the number of hairs and its length. Whereas, Sossa et al. (2003) have proposed a technique for counting objects in an image without separating the conglomerates of objects, this technique is based on the skeletonization. Similarly, in applied biological processes, Guérin et al. (2004) studied the feasibility of counting of wheat grain by colored images based on texture analysis.

In viticulture, Poni et al. (2006) found a relatively strong correlation between actual and manual count flower numbered on prints. In order to operate the automatic assessment, Cubero et al. (2014) developed a fast and accurate method for detecting and removing the pedicel in images of berries. This method is based on a novel signature of the contour. Diago et al. (2014) developed a simple and robust machine vision methodology to be applied on image taken under field conditions in order to estimate automatically the number of flowers per inflorescence. The later work is taken as reference for our comparative statistical analysis by comparing the robustness and the significance level of the methods.

About the applications on the grapevine, Dunn and Martin (2004) and Tardaguila et al. (2010) developed a program, which recognizes automatically the grapes from a digital image of the canopy of Cabernet Sauvignon grapevine. This method is used to predict vineyard yield. In order to count berries in grapes, Font et al. (2014) presented an automatic method for counting red grapes from high-resolution images of vineyards taken under artificial lighting. The proposed method is based on detecting the specular reflection peaks from the spherical

surface of the grapes. Ivorra et al. (2015) propose a three dimensional computer vision approach to assessing grape yield components based on new 3D descriptors. More advanced methodologies applied in viticulture is found in the studies of Nuske et al. (2011), Liu et al. (2013), Nuske et al. (2014), Aquino et al. (2015), Diago et al. (2015), Font et al. (2015) and Schöler et al. (2015).

The main objective of this work is to develop a new method automatic assessment system using image processing and based on watershed in order to detect and separate contiguous flower buttons. It will be pursued by a comparative statistical analysis to obtain a predictive estimation of the flower assessment in the vineyard, which the utility lies in determining fruit set rate.

MATERIALS AND METHODS

In this section, the image acquisition and the process analysis carried on these images and flower counting are described. In order to get a better counting system, a large number of images, which are considered as learning data are worked on. The study was carried out with 80 grapevine inflorescences from different Cardinal (*Vitis vinifera* L.) cultivar.

The images were taken in the field conditions using a Bridge Digital Camera (Sony HX100v) at stage H (separate flower buttons) of Baggiolini (1952) code against a dark background to assure a high contrast. The camera was set to automatic mode and the distance between the inflorescence and the camera lens was not fixed because of the inflorescence size. In the same time, a cautious and precise manual counting was carried in the field for each photographed inflorescence.

Images were processed using Matlab (v7.14) in particular the Image Processing Toolbox. The proposed algorithm is divided in two parts. In a first step, a pre-treatment is necessary to prepare the image. Then, the watershed is applied to the processed images to obtain the final segmentation of flower buttons.

The pre-treatment is based on image processing. It is a very interesting scientific topic that provides more applications (Serra, 1982; Soille, 2002). The objective is to prepare the image by cleaning (noise elimination) or by reducing the information quantity to be processed in order to keep only the most significant information. Consequently, this step is based on the mathematical morphology. The basic idea is to compare objects to be analyzed with another object of a known shape called "structuring element".

Therefore, to prepare our image (Figure 1a), the following steps were realized: (i) The images are converted to grayscale (Figure 1b). (ii) The images background is removed using the operation "Top Hat", which represents a residue for amplifying the contrast (Figure 1c). (iii) The resulting image is then converted into a binary image by using thresholding (Figure 1d). The function graythresh automatically computes an appropriate threshold to use to convert the grayscale image to binary (iv) The flower buttons are separated by morphological operations (Erode the binary image with a disk structuring element using "imerode" function, then remove small objects by the function bwareaopen and imopen) (Figure 1e).

The watershed introduced for image analysis is one of the most powerful methods to accomplish the delineation steps in image segmentation chains. The watershed technique allows partitioning the image pixels into a set of connected regions, separated by a closed contour. It is an adapted tool for segmentation (Cousty,

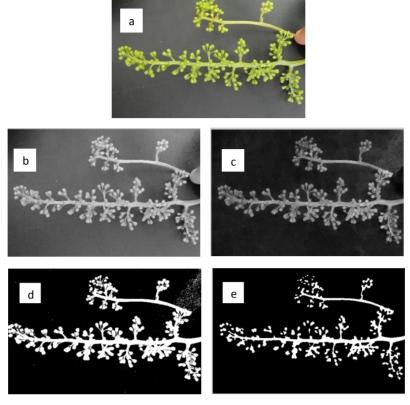


Figure 1. (a) The original image; (b) The image converted to grayscale; (c) Separating the background from the flowers clusters; (d) The image after binarisation; (e) Separation of flower buttons by morphological operations.

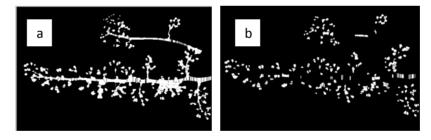


Figure 2. (a) Image after watershed function application; (b) Estimation of flower buttons number after elimination of stalk objects.

2007; Najman and Cousty, 2014; Dias and Nonato, 2015). It is the principal tool of morphological segmentation. Its major advantages, according to Meyer (2012), are the following: (i) It produces closed contours: To each minimum or to each marker corresponds one region. (ii) Flooding a topographic surface fills some minima and the watershed of the flooded surface has less catchment basin. The catchment basins of successive floodings form a hierarchical segmentation. (iii) It is possible to flood a surface so as to impose minima at some predetermined places: This leads to marker based

segmentation.

This technique was used to separate contiguous flower buttons (Figure 2a) where it is possible to label the obtained objects. For the proposed automatic counting method, it should take into account the following issues: (1) The flower buttons are grouped in clusters (superimposed flower buttons), which are not being handled by the simple counting. (2) The flower button size differs from one image to another, depending on the distance between the camera and the inflorescence. (3) There are inflorescences containing flower

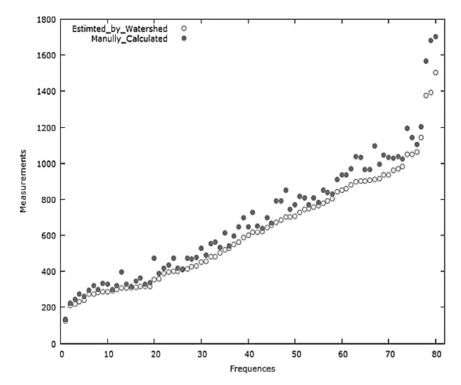


Figure 3. Graphic of the manually counted and those estimated by watershed.

buttons with different sizes in the same image. (4) The image contrast has an influence on the separation of the inflorescence from the background due to light during the image acquisition. (5) The bad separation of raffles that appears before taking images results a superimposed flower button. (6) The emergence of additional objects. These issues require changes in the developed algorithm (threshold adjustment and structural element size) from one image to another in order to eliminate the additional objects, and to separate the inflorescence from background for better detection of flower buttons.

The main objective was the determination of the correlation between manual counting (MC) procedure and automatic counting estimated by Watershed (EW) of flower buttons in our proposed method. Evaluation of the performance of the developed method was carried by the correlation coefficient (r) and the coefficient of determination (R²). Statistical analysis was conducted using SAS (v9.1).

RESULTS AND DISCUSSION

To determine the number of flower buttons per inflorescence on the image, the objects were divided into three categories: Flower buttons, objects removed because of the smaller size, the unidentified objects (stalks).

By eliminating the stalk and the small objects, a better estimation of the number of flower buttons was obtain (Figure 2b). The number found by the program (Figure 2b) is 283 flowers on the total of 301 manually counted.

This reflects an approximation of manually calculated number.

After the algorithm implementation on the total images, results named "Estimated by Watershed" (EW) were gotten. They are graphically presented with the manually counted measurements (MC) in Figure 3.

The uniform lightness from an image to another is corrected by the elimination of background (distinguishing inflorescence from background) that makes the method independent of lightness. This involves changes in the level from a grayscale image to another. The difference in brightness between different images of the database shows that the method is robust for this factor.

A sensitivity to the shape and size of the morphological filter shows that a change in size of the structuring element "disk" is required for each picture due to volume of flower buttons.

The noise has the effect to perturb the labelling of objects (flower buttons) by the existence of more objects. However, the threshold easily removes these very small objects.

Both measures showed a strong correlation (with R²: 0.99), which leads us to draw that regression line as shown in Figure 4.

Poni et al. (2006) found, by a regression between actual (real) flower number and the number of flowers

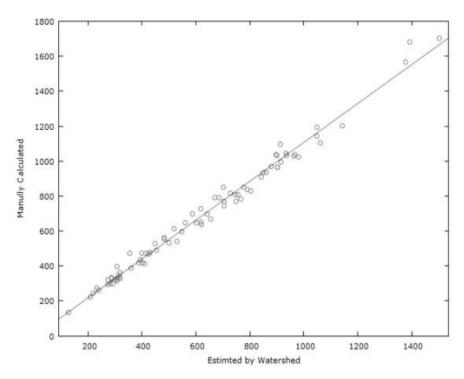


Figure 4. Relationship between the number of flower buttons manually counted and those estimated automatically by watershed ($R^2 = 0.988$ at p<0.00001].

counted on photo prints, that the coefficient of determination was 0.88 and 0.87 for the two worked varieties. Compared with our results based on automatic counting, it seems to be more significant and presents a higher correlation (R^2 =0.98).

Diago et al. (2014), by using the same software but with another function in toolbox labeled *bwconncomp*, have similar findings. Comparing our results with Diago et al. (2014, 2015) results, regardless of the used technique, it seems that our results have a more significant level. They found the coefficient of determination between 0.76 and 0.89 among the four studied cultivars.

The flower buttons assessment is done for a single upper surface (a perpendicular view of the image, where flower buttons on the other side of the inflorescence were invisible in images and consequently, undetectable by the algorithm). This makes the counting of the total flower buttons per inflorescence impossible. For this reason, we explain the underestimation of the occurred flower number.

To solve this problem, the implementation of a regression method giving low counting error (with no influence on the results) is required. Through the linear regression method, we could obtain predicted values (Figure 5). By comparing the predicted values with those counted manually, the subject of our estimation, we have

found that there is a strong correlation with quite a strong significance with a coefficient of determination of 0.99 (with p<0.00001).

To evaluate the error in the resulting model, the analysis of the linear regression estimator has shown that the values of residuals are reduced. In fact, they have low values by which its standard error of 34.4.

Returning to the precedent example, the number found by the prediction is 312 flowers, which is closer to the manual count (301 flowers) than that obtained from the watershed program (283 flowers). In order to get closer to the manually counted values, our research has shown that we should add 5% of the estimated value to the original one.

Most studies of flower number per cluster were used to estimate fruit set. Therefore, account of the flower button number per inflorescence is essential for accurate assessment of fruit set. In this research paper, we have developed a method of grapevine flower buttons assessment per inflorescence by image processing. This method is based on image processing techniques (The mathematical morphology and the watershed), with a comparative statistical analysis. The comparative analysis has shown that the applied technique presents a more significant level and high robustness compared with some previous studies.

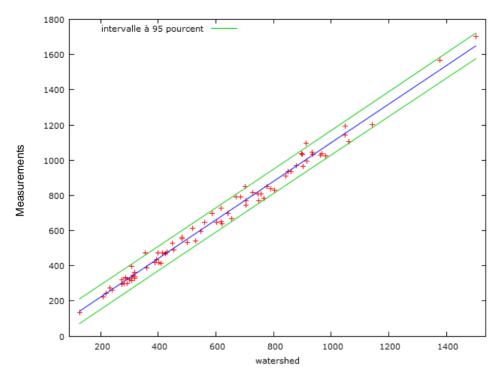


Figure 5. Graphical presentation of predictions with confidence interval of 95%.

Our study is in progress to overcome the difficulties of automatic counting and to implement a reliable tool to the flower buttons estimate per inflorescence automatically, which will be very useful and may be of great help for researchers seeking for the determination of estimated fruit set of grapevine. In the future research, efforts will focus on the correlation estimation between the automatic grape counting results and the ripeness degree of the clusters of grapes and on applying the proposed counting method to other grape varieties and to other families of fruit to estimate yield production.

Conflict of Interests

The authors have not declared any conflict of interests.

REFERENCES

Aquino A, Millan B, Gutiérrez S, Tardáguila J (2015). Grapevine flower estimation by applying artificial vision techniques on images with uncontrolled scene and multi-model analysis. Comput. Electron. Agric. 119:92-104.

Baggiolini M (1952). Les stades repères dans le déroulement annuel de la vigne et leur utilisation pratique. Rev. Romande Agric. Vitic. 8:4-6.

Bessis R (1960). Deux méthodes rapides d'appréciation du nombre de fleurs dans les grappes de la vigne. Compte Rendu Hebdomadaire des Séances de l'Acad. d'Agric. France 46:823.

Cousty J (2007). Lignes de partage des eaux discrètes: Théorie et

application à la segmentation d'images cardiaques. Thèse Doctorat, Université de Marne la Vallée.

Cubero S, Diago MP, Blasco J, Tardáguila J, Millán B, Aleixos N (2014). A new method for pedicel/peduncle detection and size assessment of grapevine berries and other fruits by image analysis. Biosyst. Eng. 117:62-72.

Diago MP, Sanz-Garcia A, Millan B, Blasco J, Tardaguila J (2014). Assessment of flower number per inflorescencein grapevine by image analysis under field conditions. J. Sci. Food Agric. 94(10):1981-1987.

Diago MP, Tardaguila J, Aleixos N, Millan B, Prats-Montalban JM, Cubero S, Blasco J (2015). Assessment of cluster yield components by image analysis. J. Sci. Food Agric. 95(6):1274-82.

Dias F, Nonato LG (2015). Some Operators from Mathematical Morphology for the Visual Analysis of Georeferenced Data. ICMC, University of Sao Paulo, Brazil.

Dunn GM, Martin SR (2004). Yield prediction from digital image analysis: A technique with potential for vineyard assessments prior to harvest. Austr. J. Grape Wine Res. 10(3):196-8.

Font D, Pallejà T, Tresanchez M, Teixidó M, Martinez D, Moreno J, Palacín J (2014). Counting red grapes in vineyards by detecting specular spherical reflection peaks in RGB images obtained at night with artificial illumination. Comput. Electron. Agric. 108:105-111.

Font D, Tresanchez M, Martínez D, Moreno J, Clotet E, Palacín J (2015). Vineyard yield estimation based on the analysis of high resolution images obtained with artificial illumination at night. Sensors 15(4):8284-301.

Girard A, Charbonnier B, Durso G (2009). Une approche par traitement statistique d'images du comptage de poils de brosses à mascara. Colloques sur le Traitement du Signal et des Images, GRETSI, Dijon-France.

Guérin D, Cointault F, Gee C, Guillemin JP (2004). Étude de faisabilité d'un système de comptage d'épis de blé par vision. Traitement de Signal 21:549-560.

- Ivorra E, Sánchez AJ, Camarasa JG, MP Diago, Tardaguila J (2015). Assessment of grape cluster yield components based on 3D descriptors using stereo vision. Food Control 50:273-282.
- Liu S, Marden S, Whitty M (2013). Towards automated yield estimation in viticulture. Proceedings of the Australasian Conference on Robotics and Automation, 24, Sydney, Australia.
- Meyer F (2012). The watershed concept and its use in segmentation: A brief history. Centre de Morphologie Mathématique, Département Maths et Systèmes, Mines-ParisTech.
- Najman L, Cousty J (2014). A graph-based mathematical morphology reader. Pattern Recogn. Lett. 1(47):3-17.
- Nuske S, Achar S, Bates T, Narasimhan S, Singh S (2011). Yield estimation in vineyards by visual grape detection. Intelligent Robots and Systems, IEEE/RSJ Int. Conf. pp. 2352-2358.
- Nuske S, Wilshusen K, Achar S, Yoder L, Narasimhan S, Singh S (2014). Automated visual yield estimation in vineyards. J. Field Robot. 31(5):837-860.
- Poni S, Casalini L, Bernizzoni F, Civardi S, Intrieri C (2006). Effects of Early Defoliation on Shoot Photosynthesis, Yield Components, and Grape Composition. Am. J. Enol. Vitic. 57:397-407.

- Schöler F, Steinhage V (2015). Automated 3D reconstruction of grape cluster architecture from sensor data for efficient phenotyping. Comput. Electron. Agric. 114:163-177.
- Serra J (1982). Image analysis and mathematical morphology. London Academic Press.
- Soille P (2002). Morphological image analysis: Principles and applications. 2nd Edition, Springer.
- Sossa H, Guzm G, Pogrebnyak O, Cuevas F (2003). Object counting without conglomerate separation. Fourth Mexican Int. Conf. on Computer Science, 8-12 Sept, Tlaxcala, Mexico pp. 216-220.
- Tardaguila J, de Toda FM, Poni S, Diago MP (2010). Impact of early leaf removal on yield and fruit and wine composition of *Vitis vinifera* L. Graciano and Carignan. Am. J. Enol. Vitic. 61(3):372-81.
- Vallotton P, Thomas N (2008). Automated body hair counting and length measurement. Skin Res. Technol. 14(4):493-497.