

Full Length Research Paper

Induction of antioxidant system in niger (*Guizotia abyssinica* Cass.) under drought stress

Kavya H. Naik and Varadahalli R. Devaraj*

Department of Biochemistry, Central College Campus, Bangalore University, Bengaluru, India.

Received 3 September, 2017; Accepted 27 September, 2017

Drought stress is a highly pervasive and economically damaging abiotic stress that affects plant yield and productivity worldwide. Physiologically, drought stress in plants is associated with oxidative stress leading to tissue damage. Drought stress imposed over 72 h in 10 days old seedlings of *Guizotia abyssinica* Cass, niger (cv; RCR-18) under greenhouse conditions resulted in elevated levels of oxidative stress markers such as H₂O₂, malondialdehyde, proline, reduced glutathione and ascorbate in a time-dependent manner. Levels of antioxidant enzymes: peroxidases and glutathione reductase, and metabolic enzyme: amylase and acid phosphatase were moderately enhanced. The levels of stress markers, antioxidants, and recovery upon re-watering suggested that the antioxidant system in niger could withstand the drought stress for up to 48 h under greenhouse conditions.

Key words: Drought stress, antioxidants, antioxidant enzymes, malondialdehyde, abiotic stress.

INTRODUCTION

As an important factor that determines geographical distribution of plant species, drought has major yield limiting ability (Nezhadahmadi et al., 2013). One of the earliest biochemical responses of eukaryotic cell to environmental stresses such as drought, salinity, high temperature, chilling etc., is generation of reactive oxygen species (ROS). Water deficit due to drought induces closing of stomata, limiting CO₂ assimilation, which in turn leads to NADPH accumulation and subsequent leakage of electrons. The leakage of electrons causes partial reduction of atmospheric O₂, generating enhanced levels of ROS (Fujita and Alam, 2015). There are four general forms of cellular ROS, singlet oxygen (¹O₂), superoxide radical (O₂⁻), hydrogen

peroxide (H₂O₂) and the hydroxyl radical (HO[·]), each having a characteristic half-life and oxidizing potential. Plants are endowed with an array of versatile and cooperative antioxidant systems comprising enzymatic and non-enzymatic components. Enzymatic components include antioxidant enzymes; guaiacol peroxidase (GPOX), catalase (CAT), ascorbate peroxidase (APX), glutathione reductase (GR) and metabolic enzymes; amylase (AMY) and acid phosphatase (AP). While, non-enzymatic components include antioxidants like ascorbic acid (ASC) and reduced glutathione (GSH), and osmoregulants like proline to cope with ROS generated in response to stress. ROS generation due to prolonged exposure to drought stress overwhelms the antioxidant

*Corresponding author. E-mail: devaraj@bub.ernet.in.

defense system, thereby causing extensive damage to cellular components like proteins, lipids, DNA and RNA, leading to cell death (Silva and Santos, 2015). Increasing population and growing soil aridity in future are expected to make water a scarce commodity. The profound impact of drought on agriculture and ecosystem thus makes the ability of plant to withstand stress of great economic importance (Rybka and Nita, 2015). Hence, it is important to look for alternative strategies to improve the abiotic stress tolerance of various crop plants.

Guizotia abyssinica (Niger) belonging to family *Asteraceae* is cultivated throughout India, West Indies and East Africa. Niger accounts for 3% of Indian oil seed production and contains 40% oil and 20% protein. Niger seed earns precious foreign exchange as it is exported as bird feed. That apart, it is mainly used for culinary purposes, manufacture of soaps, cosmetics, lighting and lubrication. Various attempts have been made to explore the potential of biodiesel production from Niger, which is cost effective, less corrosive, user and environmentally friendly green catalyst (Yerranguntla et al., 2012). Effects of polyethylene glycol 8000 and drought on *G. abyssinica* Cass cultivars (IGP 76, GA 10, No. 71 and IGP 2004) at seedling and maturity stages of the plant had been reported (Nikam and Ghane, 2011). The plant-water relationship during drought stress is an essential component for modulating the antioxidant defense mechanism. With a basic physiological and biochemical knowledge, the present study was an effort to determine the role of antioxidant defense system, in response to applied drought stress.

MATERIALS AND METHODS

Plant material and growth conditions

Niger seeds (RCR-18 variety) were procured from University of Agricultural Sciences, Dharward, India. All chemicals used were of analytical grade. Seeds were surface sterilized with 0.1% (w/v) HgCl_2 for 30 s, rinsed immediately with large volume of distilled water. The seeds were sown in plastic trays containing vermiculite and acid-washed sand (1:1 w/w) and irrigated twice a day with distilled water. The germination was carried out under natural greenhouse conditions; day/night temperature and relative humidity were 30/25°C and 75/70%, respectively. The average photoperiod was 12 h light/12 h dark.

Drought stress and experimental design

Drought stress was applied by withholding water for 10 days after germination (DAG) of seedlings. Leaf samples were collected at 24, 48 and 72 h and assayed for various parameters. The experimental design was carried out employing random factorial scheme, with 3 evaluation points (24, 48 and 72 h). Each experiment comprised 6 experimental units (leaf samples of control and stressed plants) and in triplicate. Seedlings watered twice a day were used as control.

Determination of relative water content (RWC)

The relative water content was determined by following the method

of Turner and Kramer (1980), using the equation: $\text{RWC} = (\text{FW} - \text{DW}) \times 100 / (\text{TW} - \text{DW})$. Leaf discs of 6 mm diameter were weighed to determine the fresh weight (FW); they were soaked in distilled water at 25°C for 4 h to determine the turgid weight (TW), and then oven dried at 80°C for 24 h to determine the dry weight (DW).

Determination of antioxidants and stress markers

Estimation of ascorbate (ASC) and glutathione (GSH) in control and drought stressed leaves was carried out according to Sadasivam and Manickam (1997) and Beutler et al. (1963), respectively. Further, H_2O_2 levels in control and stressed samples were determined according to Velikova et al. (2000). Lipid peroxidation was determined by estimating the malondialdehyde (MDA) content in 1 g fresh tissue according to Heath and Packer (1968). The MDA content was calculated using extinction coefficient of $155 \text{ mM}^{-1} \text{ cm}^{-1}$. Free proline was extracted from 0.5 g of fresh tissue and estimated according to the method of Bates et al. (1973).

Enzyme extraction

Fresh leaf plants were homogenized with pre-chilled 50 mM Na_2PO_4 buffer (pH 7.0), containing 5 mM β -mercaptoethanol and 1 mM EDTA. Homogenate was centrifuged at 4°C for 15 min at 12,000 $\times g$. Supernatant was used to determine the enzyme activity and protein content. Total soluble protein content was determined according to Lowry et al. (1951), using BSA as standard.

Assay of antioxidant enzymes

Guaiacol peroxidase (POX) activity was determined according to Chance and Maehly (1955) by measuring increase in absorbance at 470 nm ($\epsilon = 26.6 \text{ mM}^{-1} \text{ cm}^{-1}$) due to formation of tetraguaiacol. The reaction mixture contained 3 ml of 50 mM phosphate buffer (pH 7), 20 mM guaiacol, 10 mM H_2O_2 and 100 μl enzyme extract. One unit of enzyme activity is defined as the quantity of enzyme required to convert 1 μmol of $\text{H}_2\text{O}_2 \text{ min}^{-1}$ at 25°C.

Catalase activity was measured according to Aebi (1984) by following decline in absorbance at 240 nm ($\epsilon = 39.4 \text{ M}^{-1} \text{ cm}^{-1}$). The reaction mixture consisted of 50 mM Na_2PO_4 buffer (pH 7.0) containing 50 μl of enzyme extract and 10 mM H_2O_2 . One unit of activity is defined as the amount of enzyme that catalyzes the oxidation of 1 μmol of $\text{H}_2\text{O}_2 \text{ min}^{-1}$ under the assay conditions.

Ascorbate peroxidase activity assay was based on the method of Allen (1968) which measures an increase in the absorption at 290 nm ($\epsilon = 2.8 \text{ mM}^{-1} \text{ cm}^{-1}$). One unit of APX is defined as the amount of enzyme required to convert μmol of ascorbate min^{-1} at 25°C. The reaction mixture contained 50 mM HEPES buffer (pH 7), 1 mM EDTA, 1 mM H_2O_2 , 0.5 mM sodium ascorbate and 50 μl of enzyme extract.

GR activity was measured according to Carlberg and Mannervik (1985) by measuring oxidation of NADPH at 340 nm ($\epsilon = 6220 \text{ M}^{-1} \text{ cm}^{-1}$). The assay mixture contained 50 mM Tris-HCl buffer (pH 7.5), 3 mM MgCl_2 , 0.2 mM NADPH and 37 μl of enzyme extract. The reaction was initiated by the addition of 0.5 mM GSSG. One unit of activity is defined as the amount of enzyme that catalyzes the oxidation of 1 μmol of NADPH min^{-1} under the assay conditions.

Assay of metabolic enzymes

The activity of β -amylase was determined by the method of Bernfeld (1955). Reaction mixture contained 500 μl of 2% starch solution in 50 mM phosphate buffer (pH 7.0) and 500 μl of enzyme extract. The number of μmoles of maltose released min^{-1} is defined

as one unit of β -amylase. Acid phosphatase (AP) activity was determined by measuring the release of *p*-nitrophenol at 410 nm according to the method of Hoerling and Svensmark (1976). Each unit of activity is defined as 1 μ mole of *p*-nitro phenol released min^{-1} .

Statistical analysis

All data are expressed as a means of triplicate experiments unless mentioned otherwise and experiments were performed using randomized design. Data were subjected to analysis of variance (ANOVA) using GraphPad Prism version 6.0 and the mean differences were compared by lowest standard deviations test (LSD). Comparisons with $P \leq 0.05$ were considered significantly different.

RESULTS AND DISCUSSION

Growth parameters

Water cycling through the soil-plant-atmosphere continuum can be estimated by water potential, which also suggests the energy status of plant water. Physiological consequences leading to water deficit in cell can be appropriately measured by determining RWC. Plants exhibit varied RWC values in species specific manner and extent of drought. Some drought tolerant species have been shown to possess a range of RWC from 60 to 70%, while severely desiccated and dying leaves in drought sensitive species was about 30 to 40% (Barrs and Weatherley, 1962). Niger exhibited RWC of 75 and 50%, respectively at 48 and 72 h of drought stress (Figure 2), but 72 h stressed plants failed to revive on re-watering. This suggests the detrimental effects of drought beyond 48 h. A similar relationship between RWC and survival was observed in faba bean under salinity (Tavakkoli and Rengasamy, 2010). These effects of reduced RWC were reflected in fresh weight of drought stressed seedlings (Figure 3). However, dry weight of the seedlings remained unchanged relative to control, emphasizing primacy of physiological mechanisms in water balance under drought stress.

Stress markers

Tolerance of plants to an applied stress involves many complex and multifaceted process. Drought in plants induces oxidative stress, and ability of plants to tolerate depends upon unfolding of the genetic plasticity in order to induce specific antioxidant defense mechanism. Quantitative estimation of antioxidant components would indicate the prevalence of an adverse condition, severity of the stress and strength of antioxidant system. Oxidative stress creates imbalance in electron transport and metabolic utilization of reducing power, thereby increasing ROS levels in the cell. Progressive reduction in PS-II and β -oxidation of lipids during drought results in

generation of H_2O_2 , most stable intermediate circumventing the other ROS species (Wang et al., 2011). A 0.74-fold increase in H_2O_2 after 48 h of applied stress in niger suggested the onset of oxidative stress, as observed in *Azolla* exposed to different levels of paraquat (Sood et al., 2011). A decrease in H_2O_2 level during extended drought for 72 h suggested the onset of enzymatic and non-enzymatic antioxidative defense system to control the oxidative damage.

Level of MDA is indicative of the extent of lipid peroxidation in plant under oxidative stress induced by various environmental factors. Almost exponential increase in MDA with extended duration of drought indicated the severity of stress leading to suppression of antioxidant defense systems. While similar negative correlation between lipid peroxidation and antioxidant system has been reported in wheat (Selote and Khanna-Chopra, 2010) and maize genotypes subjected to drought stress (Vishal et al., 2013), Li and Zhang (2013) had reported lower MDA content in *Zoysia japonica* pretreated with 5 and 10 mM CaCl_2 under drought conditions. This suggests that pretreatment with the appropriate CaCl_2 concentration reduces oxidative damage that results from drought.

A number of metabolic pathways are synchronized to alleviate induced oxidative stress. Synchronized operation of ascorbate-glutathione (ASC-GSH) pathway known to establish a balance between generation and metabolism of ROS and its reaction products is an essential plant stress tolerance mechanism (Vivancos et al., 2013). GSH, a multifunctional water soluble tripeptide contributes to cell protection against detrimental effects of free radical by recycling ascorbate in its reduced form. GSH either forms adducts directly with reactive electrophiles, or acts as a proton donor in the presence of ROS, yielding GSSG; thus, providing a mechanism to protect biological macromolecules (Shao and Kang, 2008). A 1.2- fold increase in GSH level during 48 h and further decline at 72 h of drought indicated the operation of GSH-ASC cycle during early drought stress and its inefficiency during extended drought. A similar response was reported in drought stressed Hyacinth bean (Myrene and Devaraj, 2011). ASC acts as a specific electron donor to reduce H_2O_2 to water with the concomitant generation of monodehydroascorbate. As a potent reducing power, ASC maintains metalloenzyme activity and chloroplastic α -tocopherol (Pang and Wang, 2010). ASC levels in stressed seedlings of niger showed a 0.48-fold increase over 72 h of exposure time. These results are in conformity with drought stressed almond (Sorkheh et al., 2011). Parallel increase in ASC and GSH levels suggested efficient operation of ASC-GSH cycle during early period of drought stress.

Accumulations of osmoregulatory molecules such as proline, glycine betaine and sugars in response to oxidative stress had been reported in many plants (Deinlein et al., 2014). In addition to being compatible

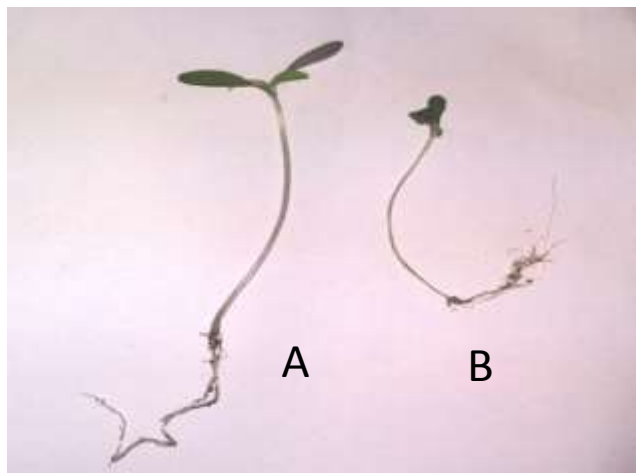


Figure 1. Effect of drought on niger seedlings: A: Control seedlings, B: stressed seedlings after 48 h.

solute and enzyme protectant, proline is also known to render structural stability to macromolecules and organelles. Increase in proline levels ~20%, as compared to control during the entire period of applied stress suggested effective osmotic adjustment contributing to osmotic stress tolerance in niger. The observed proline levels in niger were in consonance with those of drought stressed sugar cane (Abbas et al., 2014) and corn cultivars (Sinay and Karuwal, 2014) (Table 1).

Seedlings of niger were drought stressed and individual seedlings were analyzed for determination of H_2O_2 , ASC, GSH, PRO and MDA. Results are mean \pm SD, obtained from three replicates.

Antioxidant enzymes

In many plant species which exhibit stress tolerance, both enzymatic and non-enzymatic antioxidant systems operate simultaneously. Guaiacol specific POXs are known to play a vital role in various biosynthetic processes like ethylene and auxin metabolism, maintain redox homeostasis in plasma membrane, lignification and suberization of cell wall and several other developmental and defense mechanisms (Lepeduš et al., 2004) thereby, making it an indispensable component of the antioxidant system. Elevated levels of GPOX up to 48 h of applied stress, and its decline beyond 48 h suggested the enzymes potential ROS quenching during 48 h of drought stress, and its insufficiency beyond 48 h of drought (Figure 1), similar to observations made in wheat species under drought stress (Sheoran et al., 2015).

The levels of APX and GR in leaves of Niger at 48 h of drought stress showed increase in both control and stressed seedlings. Ascorbate peroxidases are class I heme-peroxidases, which utilize ASC as specific electron donor and catalyze the reduction of H_2O_2 into H_2O . APX

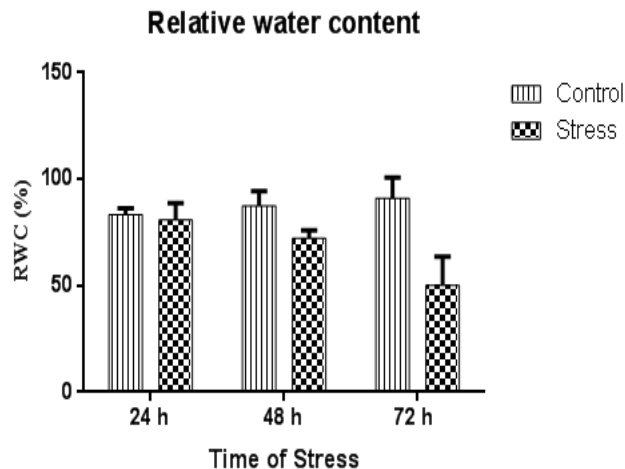


Figure 2. RWC in leaf tissues of niger seedlings at 24, 48, 72 h of drought stress.

activity in response to environmental stress generally increases along with other enzymes activities, such as CAT, POX and GR (Shigeru et al., 2002). APX showed 2.5-fold enhancements after 48 h of stress (Figure 3), which coincided with relatively lower H_2O_2 levels. Such changes had been reported in drought stressed *Solanum melongena* L. and *Piper longum* L. (Caverzan et al., 2012) and suggested to contribute to drought tolerance. GR evokes cell defense by detoxification of ROS and regeneration of GSH from GSSG with the accompanying oxidation of NADPH. Detoxification of ROS and maintenance of redox potential via NADP production by GR had been shown to contribute to abiotic stress tolerance. A 3.5-fold enhancement in GR levels in niger under drought indicated its role in stress tolerance (Figure 4). Similar activation in GR levels had been demonstrated in cowpea under water stress (Torres-Franklin and Zuily-Fodil, 2008) and tobacco under heat stress (Tan et al., 2011). Enhanced levels of APX and GR showed that they are essential components of ASC-GSH cycle. Contrary to the current findings, Chugh et al. (2010) reported inhibition of GR activity in maize (Paras, sensitive genotype) under drought stress, rendering it susceptible to drought.

Catalase showed a progressive decline during drought stress with 2.2-fold decline after 72 h (Figure 2). CATs are ubiquitous enzymes known to contain tetrameric-heme moiety catalyzing dismutation of two molecules of H_2O_2 into oxygen and water. Although, an enzyme with high turnover number CATs lower affinity towards H_2O_2 and photo-inactivation distinguishes it from other alternative H_2O_2 scavenging systems. Consequently, these inhibiting conditions lower the steady state levels of the enzyme as observed in Niger. CATs are known to be efficient tools for gross removal of high H_2O_2 levels, but they are less suited for fine tuning of sensitive redox balances with low H_2O_2 concentrations (Nicholls and

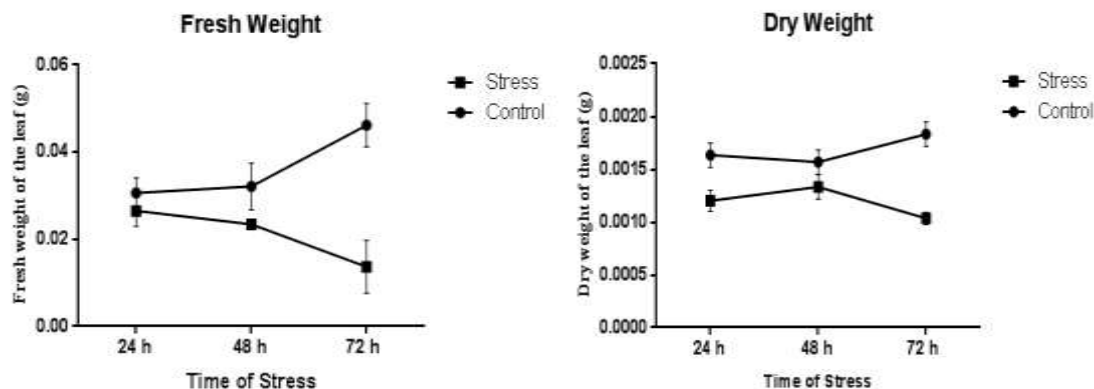


Figure 3. Fresh and dry weight of niger leaves at 24, 48 and 72 h of applied drought stress. Values for FW and DW are \pm SE (± 0.001).

Table 1. Levels of stress markers in leaves of niger (*Guizotia abyssinica*) subjected to drought stress*.

Tissue	Time	Stress marker	Control	Stress
Leaf	24 h	H ₂ O ₂ ^a	29.10664 \pm 1.902405	38.78818 \pm 0.748218
		Proline ^b	410.9 \pm 6.38	505.3 \pm 15.39976
		MDA ^c	3.460347 \pm 0.37537	4.9717 \pm 0.539988
		GSH ^a	80.605 \pm 7.230614	164.15 \pm 2.76101
		Ascorbate ^b	24.885 \pm 0.518965	33.12 \pm 0.476235
	48 h	H ₂ O ₂ ^a	33.91708 \pm 1.301969	59.12682 \pm 2.062253
		Proline ^b	488.2667 \pm 12.76784	576.3333 \pm 12.95411
		MDA ^c	3.347 \pm 0.266551	7.6508 \pm 0.447347
		GSH ^a	97.755 \pm 3.771727	195.51 \pm 6.407581
		Ascorbate ^b	29.7 \pm 1.091192	44.01 \pm 1.169134
	72 h	H ₂ O ₂ ^a	20.70311 \pm 2.352013	31.29817 \pm 2.50181
		Proline ^b	543.2667 \pm 9.648201	629.0267 \pm 9.16527
		MDA ^c	2.503267 \pm 0.109233	13.6808 \pm 0.673692
		GSH ^a	104.615 \pm 3.65863	171.5 \pm 6.095529
		Ascorbate ^b	30.555 \pm 0.273724	40.85167 \pm 0.837269

*^a μ g/g fresh weight tissue; ^bmg/g fresh weight tissue; ^cm moles/g fresh weight tissue.

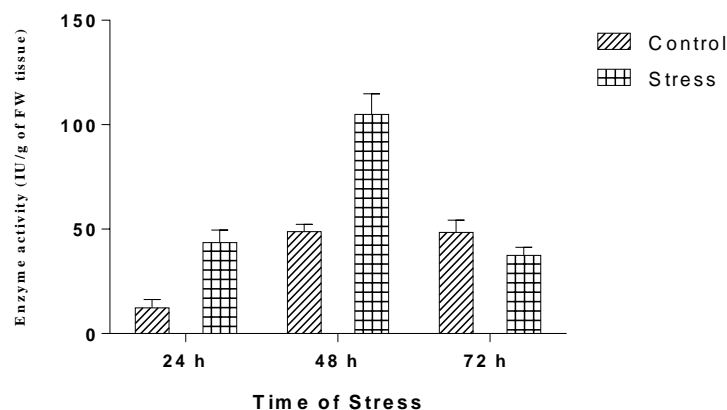


Figure 3. Effect of drought stress on APX activity in leaves of niger. Results are mean \pm SE ($P \leq 0.05$), obtained from three replicates.

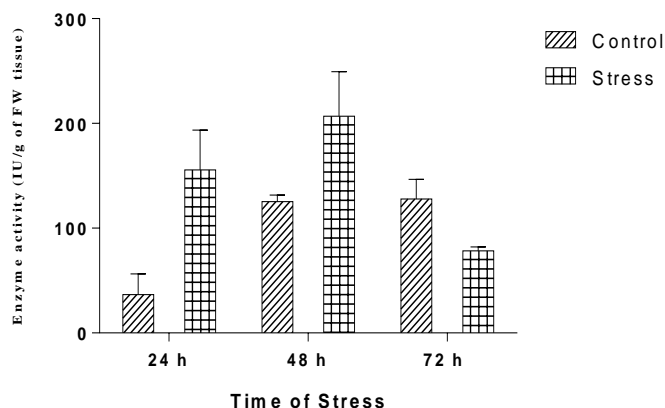


Figure 4. Effect of drought stress on GR activity in leaves of niger. Results are mean \pm SE ($P \leq 0.05$), obtained from three replicates.

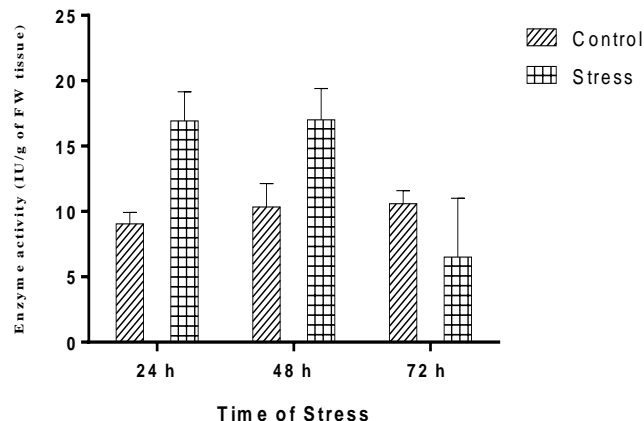


Figure 5. Effect of drought stress on AMY activity in leaves of Niger. Results are mean \pm SE ($P \leq 0.05$), obtained from three replicates.

Ferguson, 2001). On the other hand, APX is known to have greater affinity for H_2O_2 (Sofa et al., 2015), and play a pivotal role in H_2O_2 detoxification. The results are in agreement with previous studies carried out in bentgrass species (Michelle and Bingru 2007) and wheat cultivars (Chakraborty and Pradhan, 2012) subjected to drought stress, exhibiting countervailing CAT activity by APX. However, the extent of stress and susceptibility of plant species may lead to significant increase in the levels of CAT activity as was reported in oats (Islam et al., 2010) and maize (Chugh et al., 2010) under drought stress.

Though antioxidant enzymes play a pivotal role in stress tolerance, there are reports of induction of metabolic enzymes like acid phosphatase and β -amylases (Yang et al., 2007) during abiotic stress. In response to water stress, levels of inorganic phosphate are maintained by acid phosphatase, which is accomplished by co-transporting it with H^+ along the gradient of proton motive force. AP activity in niger showed 1.4-fold increase after 48 h and declined with extended exposure to stress (Figure 6). A similar observation was noted in alfalfa (*Medicago sativa* L.) explants subjected to salt and drought stress (Ehsanpour and Amini, 2003) and pigweed leaves under drought stress (Cunhua et al., 2010). β -Amylase plays a major role in starch degradation and in the daily turnover of transitory starch in photosynthetic organs. β -Amylase catalyzes the breakdown of glucans into maltose, which in cytosol is converted to glucose and ultimately leading to formation of sucrose and fructose. β -Amylase contributes to stress tolerance by increasing maltose and other soluble starch that can act as emergency compatible solutes (Krasensky and Jonal, 2012). Enhanced β -amylase levels in niger during 48 h of applied stress (Figure 5) suggested increased accumulation of sugars, which act as osmoregulant to maintain cell turgidity and maintain membrane integrity in response to the applied stress. Increase in β -amylase

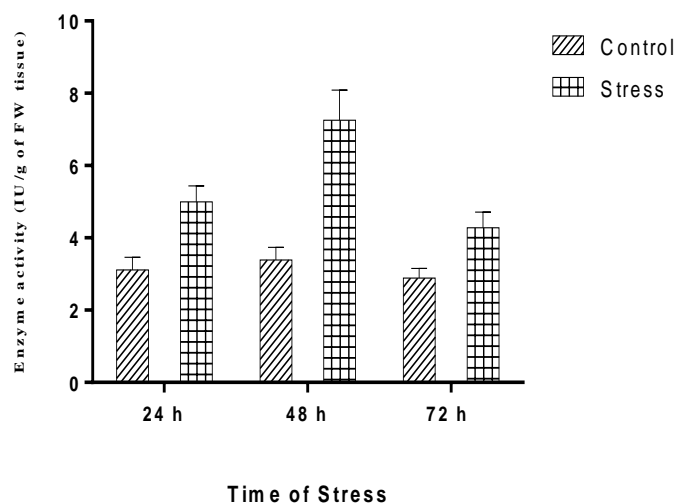


Figure 6. Effect of drought stress on AP activity in leaves of Niger. Results are mean \pm SE ($P \leq 0.05$), obtained from three replicates.

activity was seen in *Populus nigra* L. clones subjected to drought stress (Regier and Streb, 2009), which is interpreted as a tolerance mechanism.

Conclusion

Effects of different abiotic stresses such as drought on plants involve overproduction of ROS leading to oxidative stress. Plants exert a positive adaptation to drought by strategizing potent antioxidant defense systems and efficiently combat by non-enzymatic components, ASC, GSH and proline and enzymatic components, GPX, APX, GR, AMY and AP. From induction of antioxidant enzymes, enhanced antioxidants and revival of plants

upon re-watering, it is concluded that Niger is tolerant to drought stress of up to 48 h. Thus, the drought response in Niger measured in terms of antioxidant and antioxidant enzymes levels suggested participation of both components in tolerance mechanism. However, the antioxidant system employed by Niger to overcome/tolerate drought appears to be sufficient to protect against short term (up to 48 h) drought stress.

CONFLICT OF INTERESTS

The authors have not declared any conflict of interests.

ACKNOWLEDGEMENT

H. Kavya Naik acknowledges the Department of Science and Technology, Government of India for financial support with reference No. SR/WOS-A/LS-1167/2014 (G).

REFERENCES

- Abbas SR, Ahmad SD, Sabir SM, Shah AH (2014). Detection of drought tolerant sugarcane genotypes (*Saccharum officinarum*) using lipid peroxidation, antioxidant activity, glycine - betaine and proline contents. *J. Soil Sci. Plant Nutr.* 14(1):233-243.
- Aebi H (1984). Catalase *in vitro*. *Method Enzymol.* 105:121-126.
- Allen MM (1968). Simple conditions for growth of unicellular blue-green algae on plates. *J. Phycol.* 4:1-4.
- Andréia C, Gisele P, Sílvia BR, Carolina WR, Fernanda L, Márcia MP (2012). Plant responses to stresses: Role of ascorbate peroxidase in the antioxidant Protection. *Genet. Mol. Biol.* 35(4):1011-1019.
- Barrs HD, Weatherley PE (1962). A re-examination of the relative turgidity techniques for estimating water deficits in leaves. *Aust. J. Biol. Sci.* 15:413-428.
- Bates LS, Waldren RP, Teare ID (1973). Rapid determination of free proline for water stress studies. *Plant Soil.* 39:205-207.
- Bernfeld P (1955). Amylase α and β . *Method Enzymol.* 1:149-158.
- Beutler E, Duron O, Kelly BM (1963). Improved method for determination of blood glutathione. *J. Lab. Clin. Med.* 61:882.
- Boyer JS (1988). Cell enlargement and growth-induced water potentials. *Physiol. Plant* 73:311-316.
- Caverzan A, Passaia G, Rosa SB (2012). Carolina Werner Ribeiro, Fernanda Lazzarotto, Márcia Margis-Pinheiro, Plant responses to stresses: Role of ascorbate peroxidase in the antioxidant Protection. *Genet. Mol. Biol.* 35(4):1011-1019.
- Chakraborty U, Pradhan B (2012). Wheat varieties under drought stress. *Braz. J. Plant Physiol.* P 24.
- Chance B, Maehly AC (1955). Assay of catalases and peroxidases. -In: Colowick, S.P., Kaplan, N.O. (ed.): *Methods in Enzymology*. Academic Press, New York 2:764-775.
- Chugh V, Kaur N, Gupta KA (2011). Evaluation of oxidative stress tolerance in maize (*Zea mays* L.) seedlings in response to drought, *Indian J. Biochem. Biophys.* 48(1):47-53.
- Cunhua S, Wei D, Xiangling C, Xinna X, Yahong Z, Dong S, Jianjie S (2010). The effects of drought stress on the activity of acid phosphatase and its protective enzymes in pigweed leaves. *Afr. J. Biotechnol.* 9(6):825-833.
- Deinlein U, Stephan AB, Horie T, Luo W, Xu GH, Schroeder JI (2014). Plant Stress Tolerance Mechanism. *Trends Plant Sci.* 19:371-379.
- Diaz-Vivancos P, Faize M, Barba-Espin G, Faize L, Petri C, Hernández JA, Burgos L (2013). Ectopic expression of cytosolic superoxide dismutase and ascorbate peroxidase. *Plant Biotechnol. J.* 8:976-85.
- Ehsanpour AA, Amini F (2003). Effect of salt and drought stress on acid phosphatase activities in alfalfa (*Medicago sativa* L.) explants under *in vitro* culture. *Afr. J. Biotechnol.* 2(5):133-135.
- Ghane SG, Lokhande VH, Nikam TD (2012). Differential growth, physiological and biochemical responses of niger (*Guizotia abyssinica* Cass.) cultivars to water-deficit (drought) stress. *Acta Physiol. Plant* 34:215-225.
- Heath RL, Packer L (1968). Photoperoxidation in isolated chloroplasts, Kinetics and stoichiometry of fatty acid peroxidation. *Arch. Biochem. Biophys.* 125:189-198.
- Hoerling N, Svensmark O (1976). Carboxyl esterase with different substrate specificity in human brain extracts. *J. Neurochem.* 27:523-528.
- Islam MR, Xue X, Mao S, Ren C, Enejie AE, Hua Y (2010). Effects of water-saving superabsorbent polymer on antioxidant enzyme activities and lipid peroxidation in oat (*Avena sativa* L.) under drought stress. *J. Sci. Food Agric.* 15, 91(4):680-686.
- Krasensky J, Jonak C (2012). Drought, salt, and temperature stress-induced metabolic rearrangements and regulatory networks. *J. Exp. Bot.* 63(4):1593-1608.
- Lepeduš H, Cesar V, Kršnik-Rasol M (2004). Guaiacol Peroxidases in Carrot Root. *Food Technol. Biotechnol.* 42(1):33-36.
- Lowry OH, Rosebrough NJ, Farr AR, Randoll RJ (1951). Protein measurement with Folin- Phenol reagent. *J. Biol. Chem.* 193:265-275.
- Mannervik B, Carlberg I (1985). Glutathione reductase. *Methods in Enzymology* 113:484-490.
- Michelle D, Bingru H (2007). Changes in Antioxidant Enzyme Activities and Lipid Peroxidation for Bentgrass Species in Response to Drought Stress. *J. Amer. Soc. Hort. Sci.* 132(3):319-326.
- Myrene DR, Devaraj VR (2011). Specific and non-specific response to Hyacinth bean (*Dolichos lablab*) to drought stress. *India J. Biotechnol.* 10:130-139.
- Nezhadahmadi A, Prohdan ZH, Faruq G (2013). Drought Tolerance in Wheat, Hindawi Publishing Corporation. *Sci. World J.* 610721:12.
- Nicholls DG, Ferguson S (2001). Plant Responses to Drought Stress: From Morphological to Molecular Features. *Bioenergetics Academic Press, London* 3:109-111.
- Pang Q, Chen S, Dai S, Chen Y, Wang Y, Yan X (2010). Comparative proteomics of salt tolerance in *Arabidopsis thaliana* and *Thellungiella halophila*. *J. Proteome Res.* 7, 9(5):2584-2599.
- Regier N, Streb S, Coccozza C, Schaub M, Cherubini P, Zeeman CS, Frey B (2009). Drought tolerance of two black poplar (*Populus nigra* L.) clones: contribution of carbohydrates and oxidative stress defence. *Plant Cell Environ.* 32:1724-1736.
- Rybka K, Nita Z (2015). Physiological requirements for wheat ideotypes in response to drought threat. *Acta Physiol. Plant* 37:97.
- Sadasivam S, Manickam A (1997). Vitamins. In: *Biochemical methods*. New Age International (P) Limited, New Delhi, 2nd Edition pp. 185-186.
- Selote DS, Khanna- Chopra R (2010). Antioxidant response of wheat roots to drought acclimation. *Protoplasma* 245:153-163.
- Shao HB, Chu LY, Lu ZH, Kang CM (2008). Primary antioxidant free radical scavenging and redox signalling pathways in higher plant cells. *Int. J. Biol. Sci.* 4:8-14
- Sheoran S, Thakur V, Narwal S, Turan R, Mamrutha HM, Singh V, Tiwari V, Sharma I (2015). Differential Activity and Expression Profile of Antioxidant Enzymes and Physiological Changes in Wheat (*Triticum aestivum* L.) Under Drought. *Appl. Biochem. Biotechnol.* 177:1282-1298.
- Shigeru S, Takahiro I, Masahiro T, Yoshiko M, Toru Takeda, YY, Kazuya Y (2002). Regulation and function of ascorbate peroxidase isoenzymes. *J. Exp. Bot.* 53(372):1305-1319.
- Silva MA, Santos CM (2015). Physiological and biochemical responses of sugarcane to oxidative stress induced by water deficit and paraquat. *Acta Physiol. Plant* 37:172.
- Sinay H, Karuwal RL (2014). Proline and total soluble sugar content at the vegetative phase of six corn cultivars from Kisar Island Maluku, grown under drought stress conditions. *Int. J. Adv. Agric. Res.* 2:77-82.
- Sofa A, Scopa A, Nuzzaci M, Vitti A (2015). Ascorbate peroxidase and catalase activities and their genetic regulation in plants subjected to

- drought and salinity stresses. *Int. J. Mol. Sci.* 16(6):13561-1378.
- Sood A, Pabbi S, Uniyal PL (2011). Effect of paraquat on lipid peroxidation and antioxidant enzymes in aquatic fern *Azolla microphylla* Kuhl. *Russ J. Plant Physiol.* 58:667-673.
- Sorkheh K, Shiran B, Rouhi V, Khodambashi M, Sofo A (2011). Regulation of the ascorbate-glutathione cycle in wild almond during drought stress. *Russ J. Plant Physiol.* 58(1):76-84.
- Tana W, Menga QW, Brestic M, Olsovskab K, Yanga X (2011). Photosynthesis is improved by exogenous calcium in heat-stressed tobacco plants. *J. Plant Physiol.* 168:2063-2071.
- Tavakkoli E, Rengasamy P, McDonald GK (2010). High Concentration of Na^+ and Cl^- Ions in Soil Solution Have Simultaneous Detrimental Effects on Growth of Faba Bean under Salinity Stress. *J. Exp. Bot.* 61:4449-4459.
- Torres-Franklin ML, Contour-Ansel D, Zuily-Fodil Y, Pham-Thi AT (2008). Molecular cloning of glutathione reductase cDNAs and analysis of GR gene expression in cowpea and common bean leaves during recovery from moderate drought stress. *J. Plant Physiol.* 165:514-521.
- Turner NC, Kramer PJ (1980). Adaptation of plant to water and high temperature stress. Wiley Interscience Pub, New York. pp. 207-230.
- Velikova V, Yordanov I, Edreva A (2000). Oxidative stress and some antioxidant system in acid rain treated bean plants: Protective role of exogenous polyamines. *Plant Sci.* 151:59-66.
- Vishal C, Narinder K, Grewal MS, Anil GK (2013). Differential antioxidative response of tolerant and sensitive maize (*Zea mays* L.) genotypes to drought stress at reproductive stage. *Indian J. Biochem. Biophys.* 50:158-158.
- Wang YC, Lee CM, Lee LC, Tung LC, Hsieh-Li HM, Lee-Chen GJ, Su MT (2011). Mitochondrial dysfunction and oxidative stress contribute to the pathogenesis of Spinocerebellar Ataxia Type 12 (SCA12). *J. Biol. Chem.* 286(24):21742-21754.
- Yang H, Yang T, Baur JA, Perez E, Matsui T, Carmona JJ, Lamming DW, Souza-Pinto NC, Bohr VA, Rosenzweig A (2007). Nutrient-sensitive mitochondrial NAD^+ levels dictate cell survival. *Cell* 130:1095-1107.
- Yerranguntla RR, Zubaidha PK, Jakku NR, Kondhare D, Deshmukh S, Saiprakash SP (2012). Production of Biodiesel from *Guizotia abyssinica* seed oil using crystalline Manganese carbonate (MnCO_3) a Green catalyst. *Catal. Sustain. Energy* pp. 22-27.