academicJournals

Vol. 9(24), pp. 1849-1854, 12 June, 2014 DOI: 10.5897/AJAR11.2236 Article Number: C1EF98C45392 ISSN 1991-637X Copyright © 2014 Author(s) retain the copyright of this article http://www.academicjournals.org/AJAR

African Journal of Agricultural Research

Full Length Research Paper

Nutritive value assessment of three range plants by chemical and *in vitro* gas production techniques

Hamid Reza Gharehshekhlou¹, Behrouz Rasouli¹*, Ali Ahmad Ghotbi¹ and Bahram Amiri²

¹Department of Agriculture, Rasht Branch, Islamic Azad University, Rasht, Iran. ²Department of Agriculture, Firoozabad Branch, Islamic Azad University, Firuzabad, Iran.

Received 28 November, 2011; Accepted 30 May, 2014

This study was conducted in order to measure the nutritional value, as ruminant food, of three range plants (*Dactylic glomerata, Onobrychis sativa* and *Setaria galauca*), that were collected completely random from north of Iran (Guilan). Chemical analysis and *in vitro* gas production technique were used as the base for that evaluation. The chemical composition in term of ash, ether extract (EE), crude protein (CP), acid detergent fiber (ADF), neutral detergent fiber (NDF), non-fibrous carbohydrates (NFC), nitrogen free extract (NFE), crude fiber (CF) and organic material (OM) have significant differences (p < 0.05) among *D. glomerata, O. sativa* and *S. galauca*. CP were ranged from 8 (*S. galauca*) to 24% (*O. sativa*), and ADF from 37.8 (*O. sativa*) to 67.45% (*S. galauca*). Amount of fermentable fraction (b) were ranged from *D. glomerata* (68.53 ml), *O. sativa* (66.09 ml) and *S. galauca* (49.5 ml). Potential gas production (a+b) were ranged from *D. glomerata* (63.89 ml), *O. sativa* (63.39 ml) and *S. galauca* (47.4 ml). This is due to their high content of CP and low content of ADF. Therefore, according to the potential gas production performance, *D. glomerata* was ranked higher than the two other plants. The higher values obtained for the potential gas production in the *D. glomerata* and *O. sativa* indicate a better nutrient availability for rumen microorganisms. Most of chemical compounds have significant correlation with gas production factors such as fermentable part fraction (b) and potential gas production (a+b).

Key words: Range plant, nutritive value, gas production, chemical, correlation.

INTRODUCTION

More than 90 of 164 billion ha of Iran are allocated to rangelands area as an important part of livestock feed (above 80%) (Arzani et al., 2004). It is well known that forages have an important role in ruminant animal in terms of providing energy, protein and minerals as well as fibre for chewing and rumination (Kamlak, 2010). *Dactylic glomerata* and *Setaria galauca* belong to the Gramineae family and *Onobrychis sativa* belongs to the

Leguminosae (Fabaceae) family. They are widely spread in rangelands of Iran and are grazed well by ruminants especially by small ruminants. Holchek et al. (1986) results showed that the amounts of crude protein (CP), acid detergent fiber (ADF), and neutral detergent fiber (NDF) for *O. sativa* are 21.2, 52.3 and 47%, respectively. Jancik et al. (2010) results showed that the amounts of CP, NDF and ADF for *D. glomerata*

*Corresponding author. E- mail: brasooli@gmail.com; rasouli@iaurasht.ac.ir Author(s) agree that this article remain permanently open access under the terms of the <u>Creative Commons Attribution</u> <u>License 4.0 International License</u> are 13.1, 56.3 and 31.4%, respectively. Recently, some researchers have used the *in vitro* gas production technique to evaluate the fermentation kinetics of ruminant feedstuff (Mesgaran and Mohammadabadi, 2010; Chaji et al., 2010; Kamlak, 2010). In the gas method, kinetics of fermentation can be studied by simply reading the increase in gas production at a series of chosen time intervals and using the exponential equation $P = a+b (1 - e^{-ct})$ (Ørskov and McDonald, 1979). The present study was done to evaluate the nutritive value, by the use of *in vitro* incubation techniques with rumen fluid (gas production) and chemical analysis, for *D. glomerata*, *O. sativa* and *S. galauca* range plants.

MATERIALS AND METHODS

Three species of range plants, were grown naturally on the rangeland of Iran, especially north of Iran (Guilan), which receive a total annual precipitation of 500 to 1100 mm (Moghaddam, 2009). Forage samples from each herbaceous range plants were randomly taken, in three repeats (each replication were a compound of 5 samples of each species) and dried at room temperature (25 to 30°C) for 3 weeks, ground to pass through a 1 mm sieve, well mixed and stored frozen at (-20°C) in sealed nylon bags for later analysis and evaluation. Chemical analysis of forage samples were

performed according to AOAC (2005), and contents of NDF and ADF were determined by the method of Van Soest et al. (1991).

The method of Menke et al. (1979) was used to determine the rate of gas production during 96 h incubation (0, 2, 4, 6, 8, 12, 24, 48, 72 and 96 h). Digestion kinetics were calculated according to the exponential equation $P = a + b (1 - e^{-ct})$ of Ørskov and McDonald (1979), where P (ml) were defined as gas production at time (t), a (ml) was the initial gas production, b (ml) was the gas production during incubation, a + b (ml) was the potential gas production and (c) (ml/h) was the fractional gas production. The forage samples (0.200 g dry weight) were incubated in triplicate in rumen fluid, in calibrated 100 ml glass syringes at 39°C following the procedure of Menke and Steingass (1988).

The rumen fluid was collected from three rumen fistulated sheep's (same age and weight) before morning feeding (17 h after the last feed) and was homogenized and strained through 100 μ m nylon cloth into a warm flask (39°C) filled with CO₂. The fistulated sheeps were fed twice daily with a diet containing hay (60%) and concentrate (40%). A total of 30 ml of medium, consisting of 10 ml of rumen fluid and 20 ml of bicarbonate-mineral-distilled water mixture (1:1:2 by vol.), was pumped with an automatic pipette into the warmed syringes containing the samples (200 mg) and into the blank syringes. Gas production from the forage sample was calculated by subtracting the volume of gas produced from the blank with or without the addition of forage, depending on treatment. The difference in gas production as a result of treatment was calculated and expressed as a proportion of that for the untreated sample (that is, % increase).

DMD% = 83.54 - 0.824(ADF%) + 2.626(N%) (Oddy et al., 1983)

NE_(Mcal/lb) = [2.20 + (0.0272 * Gas) + (0.057*CP) + 0.149 * CF)] / 14.64 (Menke et al., 1988)

OMD (%) = 0.9991 (G_{24h}) + 0.0595 (CP) + 0.0181 (CC) + 9 (Menke and Steingass, 1988)

ME (MJ/kaDM) = 0.157 (G_{24h}) + 0.0084 (CP) + 0.022 (EE) - 0.0081(CC) + 1.06 (Menke and Steingass, 1988).

SCFA_(mmol) = 0.0222(G_{24h}) - 0.00425 (Makkar, 2005)

Where: G_{24h} is 24 h net gas production (ml/g DM), CC, CP, EE and CF are crude ash, crude protein, ether extract and fat, respectively (% of DM).

Means of the studied parameters were subjected to an analysis of variance (ANOVA) test, and Duncan test at the 95% confidence level by SPSS soft. Correlation analysis was used to establish the relationship between chemical composition and *in vitro* gas production parameters.

RESULTS

The chemical composition percentage as DM bases and correlation of chemical parameters for three range plants are presented in Tables 1 and 2, respectively. Ash, EE, CP, ADF, NDF, non fibrous carbohydrates (NFC), nitrogen free extract (NFE), crude fiber (CF) and organic material (OM) has significant differences ($p \le 0.05$) among *D. glomerata*, *O. sativa* and *S. galauca*. CP was ranged from 8 (*S. galauca*) to 24% (*O. sativa*) and ADF from 37.8 (*O. sativa*) to 45.67% (*S. galauca*).

In vitro gas production of rumen gas from the three

range plants is presented in Tables 3 and 4. Amount of gas production in all treatments has on an uptrend and it increases. Amount of produced gas (MI/200 mgDM) in three species at all times of incubation has significant differences ($P \le 0.05$). The rate of fermentation fraction (c) was significantly (p < 0.01) higher in *S. galauca* than in *O. sativa* and *D. glomerata*, but fraction (b) and fraction (a+b) were significantly (p < 0.01) higher in *O. sativa* and *D. glomerata*, but fraction (b) and fraction (a-b) were significantly (p < 0.01) higher in *O. sativa* and *D. glomerata* than in *S. galauca* (Table 3).

The rank order in terms of potential gas production performance in 0 to 24 h of incubation are *S. galauca* > *O. sativa* > *D. glomerata* and in 24 to 96 h time of incubation is *D. glomerata* > *O. sativa* > *S. galauca* (Table 4 and Figure 1).

There were significant correlations between the fermentation parameters and the chemical composition of three species (Table 5). Significant correlations were found for all gas parameters, except NE, SCFA and ME with ash. No significant correlation was found, except OMD and NE, with CF content. Negative correlations were detected between all gas parameters, except SCFA

 Table 1. Chemical composition of three range plants as percent DM bases.

Treatment	СР	EE	CF	ASH	ADF	NDF	DMD	NFE	NFC	TDN	ОМ
S. galauca	7.95 ^c	4.06 ^a	31.00 ^a	4.23 ^b	45.67 ^a	73.93 ^a	49.25 ^c	49.7 ^a	38.09 ^a	54.93 ^c	95.77 ^a
D. glomerata	12.16 ^b	3.47 ^a	28.13 ^b	6.09 ^{ab}	41.81 ^b	72.78 ^b	54.19 ^b	45.24 ^b	35.56 ^a	55.71 ^b	93.00 ^b
O. sativa	23.95 ^a	2.51 ^b	12.43 ^c	7.00 ^b	37.78 ^c	53.84 ^c	62.47 ^a	51.16 ^ª	29.667 ^b	68.59 ^a	93.91 ^{ab}

Columns having different superscripts are significantly different (p < 0.05).

Table 2. Correlation coefficient (r) between chemical composition parameters in three range plants.

Pearson correlation	СР	EE	CF	ASH	ADF	NDF	DMD	NFE	NFC	TDN	ОМ
CP	1.00										
EE	-0.87**	1.00									
CF	-0.99**	0.81**	1.00								
ASH	0.37 ^{ns}	-0.44 ^{ns}	-0.29 ^{ns}	1.00							
ADF	-0.96**	0.86**	0.92**	-0.49 ^{ns}	1.00						
NDF	-0.97**	0.82**	0.99**	-0.18 ^{ns}	0.89**	1.00					
DMD	0.991**	-0.871**	-0.97**	0.42 ^{ns}	-0.99**	-0.95**	1.00				
NFE	0.49 ^{ns}	-0.40 ^{ns}	-0.58 ^{ns}	-0.56 ^{ns}	-0.31 ^{ns}	-0.65 ^{ns}	0.41	1.00			
NFC	-0.95**	0.798**	0.936**	-0.56 ^{ns}	0.90**	0.89**	-0.93**	-0.31 ^{ns}	1.00		
TDN	0.97**	-0.82**	-0.99**	0.18 ^{ns}	-0.896**	-1.00**	0.945**	0.65 ^{ns}	-0.89**	1.00	
OM	-0.37 ^{ns}	0.44 ^{ns}	0.29 ^{ns}	-1.00**	0.49 ^{ns}	0.18 ^{ns}	-0.43 ^{ns}	0.56 ^{ns}	0.56 ^{ns}	-0.18 ^{ns}	1.00

*p < 0.05; **p < 0.01; ns = Non significant.

Table 3. Parameters of *in vitro* gas production in three range plants (defined by the equation: $p = a + b (1 - exp^{-ct})$.

Treaturent	Incubation time										
Treatment	а	b	С	a+b	DMD	OMD	SCFA	ME	NE		
S. galauca	-2.08 ^b	49.5 ^b	0.09 ^a	47.42 ^b	55.20 ^b	55.43 ^b	9.38 ^b	7.55 ^b	6.11 ^a		
D. glomerata	-4.68 ^a	68.53 ^a	0.04 ^b	63.85 ^a	69.24 ^a	64.3 ^a	10.65 ^a	8.52 ^a	6.3 ^a		
O. sativa	-2.71 ^b	66.09 ^a	0.04 ^b	63.39 ^a	69.06 ^a	65.92 ^a	9.3 ^b	7.6 ^b	4.62 ^b		
SEM	0.2	0.5	0.27	0.34	0.39	1.36	0.21	0.21	0.12		
Significant	**	**	**	**	**	**	**	*	**		

c = Rate constant of gas production during incubation (ml h^{-1}); a = gas produced from soluble fraction (ml); b = gas produced from insoluble but fermentable fraction (ml); a+b = potential gas production (ml); Columns having different superscripts significantly (p < 0.01).

and ME, with ADF content (P < 0.01) (Table 5).

DISCUSSION

The results showed that there were significant variations in chemical composition and gas production characteristics of three range plants. Amount of CP in *S. galauca, D. glomerata* and *O. sativa* is 7.65, 12.1 and 23.95%, respectively. Theminimal CP content of DM for maintenance of sheep has been indicated by Milford and Haydock (1965) to be 7.2%. However, it was suggested to be at least 8.9% CP in plant material. The CP value in the present study plants were mostly well above the recommended levels by Milford and Haydock (1965) and NRC (1990), suggesting that they might maintain animals. On the other hand, the CP of *S. galauca* is fitted for sustain sheep if used as the only sources of feed. Range land forages are composed of structural and nonstructural constituents. Rezayi (2004) reported that amount of CP belonged to theleaf:stem ratio and causes an increase in plant protein thus this reason amount of CP in *O. sativa* is more than that of *S. galauca* and *D. glomerata*. The leaves of *O. sativa* is phyllde form, but *S. galauca* and *D. glomerata* do not have phyllde leaves (Gramine family) and the leaf:stem ratio is low.

The surface of a leaf will be increased growing rate, DM and protein content as a result of increasing number of photosynthetic organ per unit of leaf surface (Hattab and Harb, 1990). The results showed that the ADF

Treatment	Incubation time									
Treatment	2	4	6	8	12	24	48	72	96	
S. galauca	3.74 ^a	9.67 ^a	18.12 ^a	24.38 ^a	32.83 ^a	39.00 ^b	44.69 ^c	47.94 ^c	49.08 ^c	
D. glomerata	2.12 ^b	4.16 ^c	7.10 ^c	10.69 ^c	21.46 ^b	44.31 ^a	56.40 ^a	59.91 ^a	60.98 ^b	
O. sativa	2.44 ^b	5.11 ^b	8.53 ^b	12.91 ^b	20.63 ^c	38.67 ^b	53.45 ^b	57.68 ^b	61.42 ^a	
SEM	0.21	0.27	0.36	0.03	0.42	0.73	0.89	0.4	0.24	
Significant	**	**	**	**	**	**	**	**	**	

Table 4. Rumen gas production (ml/200 mg DM) from three range plants under different times.

Columns having different superscripts are significantly different (p < 0.05).

Table 5. The correlation coefficients (r) between the chemical composition and gas production parameters.

Pearson correlation	СР	EE	CF	ASH	ADF	NDF
а	0.03 ^{ns}	0.09 ^{ns}	-0.14 ^{ns}	-0.67*	0.22 ^{ns}	-0.27 ^{ns}
b	0.61 ^{ns}	-0.62 ^{ns}	-0.52 ^{ns}	-0.74*	-0.79*	-0.44 ^{ns}
С	-0.70*	0.69*	0.61 ^{ns}	-0.72*	-0.86**	0.54 ^{ns}
a+b	0.68*	-0.67*	-0.59 ^{ns}	0.72*	-0.85**	0.54 ^{ns}
OMD	0.79*	-0.75*	-0.72*	0.68*	-0.92**	-0.66 ^{ns}
DMD	0.69*	-0.68*	-0.60 ^{ns}	0.72*	-0.85**	-0.53 ^{ns}
SCFA	-0.31 ^{ns}	0.16 ^{ns}	0.42 ^{ns}	0.54 ^{ns}	0.07 ^{ns}	0.50 ^{ns}
ME	-0.22 ^{ns}	0.08 ^{ns}	0.33 ^{ns}	0.59 ^{ns}	-0.03 ^{ns}	0.41 ^{ns}
NE	-0.93**	0.76**	0.96**	-0.07 ^{ns}	0.82**	0.99**

c = Rate constant of gas production during incubation (% h^{-1}); a = gas produced from soluble fraction (ml/0.200 g OM); b = gas produced from insoluble but fermentable fraction (ml/0.2 g OM); a+b = potential gas production (ml/0.200 g OM); *P < 0.05, **P < 0.01***P < 0.001; ns = non significant.

content was ranged from 37.8 (*O. sativa*) to 67.45% (*S. galauca*). The leaf:stem ratio decreased in the fibre content and an increased level of protein in the plants (Aaron et al., 2005). Amount of CP, NDF and ADF of *O. sativa* is 25, 53.8 and 37%, respectively. This result is in agreement with findings by Holchek et al. (1986) who reported 21.2, 52.3 and 47%, respectively. Arzani et al. (2006) reported that CP of legume family is more than gramine family and ADF is lower. However, in contrast to our results, Kaplan (2011) showed lesser CP (16.9%) and same ADF and NDF according to chemical compounds for the *O. sativa*. Jancik et al. (2010) detected that amount of CP (13.1%),

NDF (56.3%) and ADF (31.4%) contents are in agreement with present findings in the *D. glomerata*. Results of Bostan et al. (2010), in *D. glomerata* showed that the amount of ADF (31%) is lesser than that of present findings, CP (18%) is more than that of present findings and NDF (71%) is similar with that of present findings. The highest amount of NDF and ADF are caused to decreasing dry matter digestibility (DMD) and increasing plant fiber (van Soest et al., 1991; Mohanty et al., 2000; Beakou et al., 2008). The result showed that the highest level of gas production occurred after 16 to 24 h incubation. This stage of incubation in ruminants showed that fermentation of forage is maximal. It related

to the ration and its constituents, and for more readily digestible carbohydrates is 12 to 16 h and for less digestible carbohydrates is 24 to 96 h (Kinan and Krishnamoorthy, 2007; Vanic et al., 2008).

The most of fermentable fraction (b) and the potential gas production (a+b) in 0 to 20 h of incubation time belonged to S. galauca and after 20 h of incubation time, belonged to O. sativa, D. glomerata and S. galauca, respectively. It may be due to their content of ADF, NDF and protein, whereas the potential gas production (a+b) is associated with degradability of feed (Kamalak et al., 2005). Therefore, the higher values obtained for the potential gas production in the O. sativa indicated a better nutrient availability for rumen microorganisms. ADF and EE correlation with fraction (b) and (a+b) were negative but ash and CP correlation with fraction (b) and (a+b) were positive. This result is consistent with findings of Frutos et al. (2002). ADF were negatively correlated with most of the estimated parameters. This result is in agreement with the findings of Abdulrazak et al. (2000) and Kamalak et al. (2004).

The negative correlation between potential gas production and ADF may be due to the reduction of microbial activity from increasingly adverse environmental conditions as incubation time progress. CP was positively correlated with the rate of fermentation

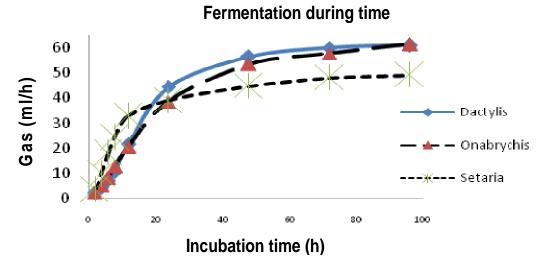


Figure 1. Cumulative gas production of three range plants.

fraction (c), fraction (b) and fraction (a+b). These findings were supported by Kamalak et al. (2004). Correlation relationship of chemical composition with gas production kinetics and some estimated parameters are in agreement with findings of Kaplan (2011). The chemical characteristics correlation showed that CP was negatively correlated with EE, ADF, CF, and NDF. The high amount of a leaf could be increased growing rate, DM and protein content and decreased ADF, CF, and NDF as a result of increasing number of photosynthetic organ per unit of leaf (Hattab and Harb, 1990).

Conclusion

The results revealed that the *O. sativa, D. glomerata* and *S. galauca* could be used such as forage for livestock, respectively.

REFERENCES

- Aaron J, Schwart L, Gibson R, Douglas L, Karlen ML, Jean-Luc J
(2005). Planting date effect on winter triticale dry matter and nitrogen
accumulation.Agron.J.97:1333-1341.http://dx.doi.org/10.2134/agronj2005.0010
- Abdulrazak SA, Fujihara T, Ondiek JK, Ørskov ER (2000). Utritive evaluation of some Acacia tree leaves from Kenya. An. Feed Sci. Technol. 85:89-98. http://dx.doi.org/10.1016/S0377-8401(00)00133-4
- AOAC (2005). Official Methods of Analysis, AOAC International.18th ed. Gaithersburg, USA.
- Arzani H, Zohdi M, Fisher E, Zaheddi Amiri GH, Nikkhan A, Waster D (2004). Phenological effects on forage quality of five grass species, J. Range. Manage. 57:624-630. http://dx.doi.org/10.2307/4004019
- Arzani H, Basiri M, Khatibi F, Ghorbani G (2006). Nutritive value of some zagros mountain rangeland species. Small Rum. Res. 65:128-135. http://dx.doi.org/10.1016/j.smallrumres.2005.05.033
- Beakou A, Ntenga R, Lepetit J, Ateba JA, Ayin LO (2008). Physicochemical and micro Structural characterization of Rhectophyllum

amerunense, Composites. Part A. Appl. Sci. Manuf. 39(1):67-74. http://dx.doi.org/10.1016/j.compositesa.2007.09.002

- Bostan C, Moisuc A, Radu F, Cojocariu L, Sarateanu V (2010). Study of the action of *Poa pratensis* L. vegetal extract on the chemical Composition of some perennial grasses. Res. J. Agric. Sci. 42(1).
- Chaji M, Mohammadabadi T, Mamouei M, Tabatabaei S (2010). The effect of processing with high steam and sodium hydroxide on nutritive value of sugarcane pith by *in vitro* gas production. J. Anim. Vent. Advan. 9:1015-1018. http://dx.doi.org/10.3923/javaa.2010.1015.1018
- Frutos P, Hervas G, Ramos G, Giraldez FJ,Mantecon AR (2002). Condensed tannin content several shrub species from a mountain area in northern Spain and its relationship to various indicators of nutritive value. An. Feed Sci. Technol. 95:215-226. http://dx.doi.org/10.1016/S0377-8401(01)00323-6
- Hattab AH, Harb MY (1990). Effect of planting date and nitrogen levels on forage yield and quality in sorghum sodan grasses hybrid in the central valley of Jordan. Dirasat Univ. Jordan, Ser. B 18:70-92.
- Holchek JL, Wofford H, Artgun D, Galyean ML, Wallace JD (1986). Evaluation of total fecal collection for measuring cattle forages. J. Range. Manage. 1:39.
- Jancik F, Koukolová V, Homolka P (2010). Ruminal degradability of dry matter and neutral detergent fiber of grasses. Czech J. Anim. Sci. 55(9):359-371.
- Kamalak A, Canbolat O, Gurbuz Y, Ozay O, Ozkan CO, Sakaray M (2004). Chemical composition and *in vitro*, gas production characteristics of several tannin containing tree leaves. Livestock research for rural development

http://www.cipav.org.co/lrrd/lrrd16/6/kam http://dx.doi.org/10.1080/09712119.2010.9707157

- Kamalak A, Canbolat O, Gurbuz Y, Ozay O, Ozkose E (2005). Chemical composition and its relationship to *in vitro* gas production of several tannin containing trees and shrub leaves. Asian-Aust. J. Anim. Sci.18:203-208.
- Kamlak A (2010). Determination of potential nutritive value of *Polygonum aviculare* hay harvested at three maturity stages. J. Appl. Anim. Res. 38:69-71.
- Kaplan M (2011).Determination of potential nutritive value of
(Onobrychis sativa) hays harvested at flowering stage.J. Anim.Veter.Advan.10(15):2028-2031.http://dx.doi.org/10.3923/javaa.2011.2028.2031
- Kinan D, Krishnamoorthy U (2007). Rumen fermentation and microbial biomass synthsis indices of tropical feedstuffs determined by the *in vitro* gas production technique. Anim. Feed Sci. Technol. 134(1-2):170-179. http://dx.doi.org/10.1016/j.anifeedsci.2006.05.017

- Menke KH, Steingass H (1988). Estimation of the energetic feed value obtained from chemical analysis and gas production using rumen fluid. Ani. Res. Develop. 28:7-55.
- Menke K, Raab L, Salewski A, Steingass H, Fritz D, Schneider W (1979). The estimation of digestibility and metabolizable energy content of ruminant feeding stuffs from the gas production when they are incubated with rumen liquor *in vitro*. J. Agric. Sci. Camb. 3:217-222. http://dx.doi.org/10.1017/S0021859600086305
- Mesgaran MD, Mohammadabadi M (2010). The effect of fat content of chemically treated sunflower meal on *in vitro* gas production parameters using isolated rumen microbiota. J. Anim. Vet. Adv. 9:2466-2471. http://dx.doi.org/10.3923/javaa.2010.2466.2471
- Milford R, Haydock KP (1965).The nutritive value of protein in subtropical pasture species grown in southeast Queensland. Aust. J. Exp. Agric. Anim. Husb. 5:13-17. http://dx.doi.org/10.1071/EA9650013
- Moghaddam MR (2009). Rangeland Management, 2rd Edition, Publication of Tehran University of Natural Resources.
- NRC (1990). National Research Council: Nutrient requirements of domestic animals goats: National Academy Press. Washington DC.
- Mohanty AK, Misra M, Hinrichsen G (2000). Biofibers, An. Overview. Macromol. Matter Eng. 276-277:1-24. http://dx.doi.org/10.1002/(SICI)1439-2054(20000301)276:1<1::AID-MAME1>3.0.CO;2-W
- Ørskov ER, McDonald P (1979). The estimation of protein degradability in the rumen from incubation measurements weighed according to rate of passage. J. Agric. Sci. 92:499-503. http://dx.doi.org/10.1017/S0021859600063048

- Rezayi A (2004). Investigation of different phenology and chemical composition and nutritive value of *Onobrychis sativa*, Bo Ali University.
- Van Soest PJ, Robertson JD, Lewis BA (1991).Methods for dietary fibre, neutral detergent fibre and non-starch polysaccharides In relation to animal nutrition. J. Dairy Sci. 74:3583-3597. http://dx.doi.org/10.3168/jds.S0022-0302(91)78551-2
- Vanic M, Knezevic M, Bosnjak K, Leto J, Perculija G, Matic I (2008). Effects of replacing grass silage harvested at two maturity stages with maize silage in the ration upon the intake, digestibility and N retention in weather sheep. J. Livest. Sci. 114(1):84-92. http://dx.doi.org/10.1016/j.livsci.2007.04.011