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Full Length Research Paper

Effect of ethanolic leaf extract of *Vinca major* L. on biochemical parameters and glucose level of alloxan induced diabetic rats

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In view of World Health Organization (WHO) recommendation on the use of medicinal plants for the management of diabetes mellitus, this study evaluated the effect of ethanolic leaf extract of Vinca major L. on biochemical parameters and glucose level of alloxan induced diabetic rats as objective. Sixty-four male Wistar albino rats were induced with diabetes by a single intraperitoneal injection and were separated into four groups (1-4) of sixteen rats each. Group 1 served as the control while groups 2 to 4 served as the test groups. Apart from the control, the test groups were treated with different concentrations of the leaf extract, and four rats from each group were sacrificed every seven days for assessment. The treatment period lasted for 28 days. Haemoglobin (Hb) levels in groups 3 and 4 rats significantly increased (p<0.05) against the control throughout the number of days of the study. Red blood cell (RBC) levels in groups 2 and 4 increased significantly (p<0.05) against the control on the 14th day, while all the test group rats had significantly increased (p<0.05) RBC levels on the 21st and 28th days of the study. The observed trend followed by electrolyte ions, urea and creatinine in test rats against the control in the present study, may be attributed to V. major leaf extract trying to salvage the excretory organs of alloxan battered diabetic rats. Glucose level, aspartate aminotransferase (AST), alanine aminotransferase (ALT) and alkaline phosphatase (ALP) liver enzymes reduced significantly (p<0.05) in test rats against the control. Diabetic rats treated with V. major leaf extract in this study, showed significant improvements on those associate problems of diabetes which include anaemia, dyslipidaemia, and hepatic necrosis and inflammation. Rats treated with the leaf extract also showed reduced glucose level (Hypoglycemia). From the observations of this study, extract from leaf of V. major may be effective against diabetes and some of its associate problems.

Key words: diabetes, biochemical parameters, Vinca major, leaf extract, Wistar rats.

INTRODUCTION

Diabetes is among the diseases that plagued the existence of man on this planet Earth. According to Demoz et al., (2015), diabetes is a complex and chronic illness characterized by hyperglycemia resulting from defects in insulin secretion, insulin action, or both. Edem (2009) and Ogbonnia et al., (2008) noted that diabetes is a major degenerative disease in the world today, affecting millions of people and having complications which include hypertension, atherosclerosis and microcirculatory disorders. Demoz et al., (2015) reported that over 3 million deaths worldwide are attributed to diabetes every year. Over 380 million people were reportedly living with diabetes in 2014, and the number is projected to rise by more than double by 2035 without any intervention (Shrivastava et al., 2013). Type 2 diabetes mellitus (T2DM) is among the existing types of diabetic disorder with rapidly increasing incidence in the world (Demoz et al., 2015; Islam et al., 2017). According to Kesari et al., (2007), Type 2 diabetes mellitus accounts for about 90% of diabetic population. Islam et al., (2017) and Bastaki (2005) noted that among all the endocrine disorders, diabetes mellitus is the most prevalent one and almost three hundred million will suffer from the disease by 2025. Ethnopharmacological studies revealed that more than 1200 plants are utilized in traditional medicine for their alleged hypoglycemic activity (Marles and Farnsworth, 1995; Nole et al., 2016; Tsabang et al., 2016). Such plants are known as medicinal plants and are the main source of organic compounds such as polyphenols. tannins. alkaloids. carbohvdrates. terpenoids, steroids, flavonoids and etcetera (Marles and Farnsworth, 1995). These compounds are collectively addressed as phytochemicals (Marles and Farnsworth, 1995; Amadi et al., 2012; Duru et al., 2018) and represent a source for the discovery and development of new types of antidiabetic molecules (Firdous, 2014). Nawrot et al. (2003) and Agomuo et al., (2017) noted that natural compounds found in plants or their synthetic forms are the basis of modern pharmacopeia.

Vinca major L. an Apocynaceae also known as periwinkle, is one of such medicinal plants with some of these organic compounds addressed as phytochemicals. It is an evergreen perennial trailing vine plant that roots along its stems to form dense masses of groundcover. It can grow up to 25 cm (10 in) high while spreading indefinitely (Blamey and Grey-Wilson, 1989). The leaves of *V. major* can grow up to 9 cm long and 6 cm broad. Its flowers are hermaphrodite in nature (Blamey and Grey-Wilson, 1989). The plant is found along river banks and grows well in full sun and shade at an altitude between 0 and 800 metres (0-2,625 ft) above sea level (Blamey and

Grey-Wilson, 1989). Extract of the plant is used against diseases such as malaria, leukemia and Hodgkin's disease. The leaf juice of V. major is used in the treatment of treat wasp stings, and sore throats. The flower extract is used for infants' eyewash. The plant is used in the preparation of periwinkle tea, which is used in the treatment of cough (Blamey and Grey-Wilson, 1989). Due to adverse effects and other factors militating against the use of many synthetic anti-diabetic agents, medicinal plants continue to play very vital role in the management of diabetes mellitus in developing country. It has been reported that plants products that are effective against diabetic disease are cost effective and less toxic than synthesised drugs (Jung et al., 2006; Patel et al., 2012). In view of World Health Organization (WHO) recommendation on the use of medicinal plants for the management of diabetes mellitus, there is need to expand the frontiers of scientific research to discover more plants with anti-diabetic properties, especially now that recent findings have shown a rise in number of new cases of type 2 DM with an earlier onset and associated complications in developing countries. Hence upturning the earlier believe that diabetes is mainly found in developed countries of the world.

Through *in vivo* studies using animal models, a number of plants with anti-diabetic activity have been discovered (Bnouham et al., 2006; Kayarohanam et al., 2015; Mussie et al., 2015; Tsabang et al., 2017), but there is paucity of studies on *V. major* in relation to diabetic activity and other biochemical parameters. The present study investigated the effect of ethanolic leaf extract of *Vinca major* L. on biochemical parameters and glucose level of alloxan induced diabetic rats.

MATERIALS AND METHODS

Plant collection, identification and preparation

Fresh leaves of *V. major* L. were collected from the plant between the months of February and March 2018, in Choba, Rivers State, Nigeria. Taxonomic identification was done and voucher specimen was deposited at the Herbarium unit of Rhema University, Nigeria. The identified leaves were air dried for two weeks and then blended into powder.

Plant extraction and toxicity study

The method as described by Adebayo et al., (2006) was used for preparation of plant extract. The powdered leaves of *V. major* (800 g) were soaked in 8 L of 70% ethanol for four days, after which the extract was filtered using a Whatman no. 1 filter paper and a cotton

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Author(s) agree that this article remain permanently open access under the terms of the <u>Creative Commons Attribution</u> <u>License 4.0 International License</u> wool. It was further concentrated at 50°C using a rotary evaporator and further concentrated using water bath at 48°C. The weight of the extract obtained was 80 g giving a percentage yield of 10%. The toxicity study (LD_{50}) was carried using the method as described by Lorke (1983), and the toxic dosage (LD_{50}) for the leaf extract was found to be above 6,000 mg/kg.

Experimental design, induction of diabetes and treatment of experimental animals

A total of sixty four male Wistar albino rats were procured from the animal colony of Department of Biochemistry, University of Port Harcourt, Nigeria for the present study. The rats were housed in plastic cages covered with wire mesh with facilities for food and water in them. The water and feeds were given *ad libitium*. The rat feed was a brand of commercial grower freshly obtained from a feed dealer along Abayi road, Aba, Nigeria. After acclimatization period, the experimental rats were made diabetic by Induction with alloxan monohydrate which was prepared by dissolving 10 g in 100 mL sterilized water. By single intraperitoneal injection, diabetes was induced in fasting rats by administration of 150 mg/kg body weight alloxan to each rat. Rats with blood glucose level above 200 mg/dL (after three days of post induction) were considered diabetic and chosen for the study.

The induced diabetic rats were then separated into four (1-4) groups of sixteen rats each. Group 1 served as the control, while groups 2 to 4 served as test groups. The treatments given to the groups are shown below

Group 1(Control): Diabetic rats treated normal saline (0.9% (w/v) NaCl)

Group 2: Diabetic rats treated with 100 mg/kg body weight of extract Group 3: Diabetic rats treated with 250 mg/kg body weight of the extract

Group 4: Diabetic rats treated with 450 mg/kg body weight of the extract

The extract was given orally daily. The groups were placed on the same feed and water for twenty-eight days. The floors of the cages were constantly cleaned at interval of two days, their feed consumption and body weights were taken at interval of seven days.

Experimental handling of animals was in accordance with international guidelines on animal care and uses (NIH, 1985).

Samples collection for analysis

Four rats from each group were sacrificed weekly (7, 14, 21 and 28 days) after subjecting them to overnight fast. The rats were subsequently anaesthetized with diethyl-ether and blood samples were collected by cardiac puncture into clean tubes for liver enzymes, electrolyte ions, urea, creatinine and glucose studies. Blood samples for haematological indices were collected with anticoagulant tubes. The tubes were properly labelled and used for analysis.

Haematology analysis

The autoanalyzer machine was used for hematological analysis. The blood samples contained in the anticoagulant tubes were swirled/rolled on the blood roller each for five seconds and then opened and put under the probe of the autoanalyzer. The probe then collected the blood from the tubes for about 10 s and entered back into the haematology machine. The result was then printed a few seconds later, giving the parameters haemoglobin (Hb), red blood cells (RBC), white blood cells (WBC), lymphocytes, monocytes, Basophils, mean corpuscular volume (MCV), and mean corpuscular haemoglobin concentration (MCHC). Haematocrit (PCV) level was estimated using microhaematocrit methods as described by Alexander and Griffiths (1993).

Biochemical assays

All the biochemical parameters investigated were measured using BUCK 211 spectrophotometer, England. The following biochemical parameters were carried out in serum; urea was analysed using the Bethlot Searcy's method (Searcy et al., 1967); creatinine was determined by the method described by Larsen (1971). Sodium ion (Na⁺) and chloride (Cl⁻) ion levels were determined according to the instructions on their diagnostic kits purchased from Randox laboratories (UK). Potassium ion (K⁺) was determined by direct spectrophotometric method. Bicarbonate (HCO₃) was determined using Forrester et al., (1979) method. Urea estimation was done using Urease-Berthlot method. Creatinine was estimated using the method described by Heinegård and Tiderström (1973). Total cholesterol, high density lipoprotein cholesterol (HDL-cholesterol), and triglyceride lipid profiles were assayed enzymatically with their test kits (Randox Laboratories, England). The relations as described by Friedwald et al. (1972) were used for low density lipoprotein cholesterol (LDL-cholesterol) estimation. Liver enzyme such as alanine aminotransferase (ALT) and aspartate aminotransferase (AST) were estimated using Reitman and Frankel (1957) methods; alkaline phosphatase (ALP) was carried out using the phenolphthalein monophosphate method (Babson et al., 1966); total bilirubin was calculated using Doumas et al., (1973) method; whereas glucose level was obtained using the enzymatic GOD-PAP method as described by Trinder (1969).

Statistical analysis

Results are presented as mean and standard deviations of triplicate determinations. Group comparisons were done using least significant difference (LSD). Significant difference was established at 5% level as described by Onu and Igwemma (2000). Bars of the same day with different letters of alphabet are statistically significant at p<0.05.

RESULTS

Figures 1 to 9 show haematology result of rats given *V. major* leaf extract at different concentrations. From the Figures, Hb ranged from 9.11 to 12.85 g/dl (Figure 1); haematocrit or PCV ranged from 25.80 to 38.00% (Figure 2); RBC ranged from 2.75-5.48×10⁹ cell/L (Figure 3); WBC ranged from 2.10-4.53×10⁹ cell/L (Figure 4); lymphocytes ranged from 35.10-66.00% (Figure 5); monocytes ranged from 14.10-19.95% (Figure 6); basophil ranged from 1.50-4.68% (Figure 7); mean cell volume (MCV) ranged from 71.70-98.30 fl (Figure 8); and mean corpuscular haemoglobin concentration (MCHC) ranged from 26.60-36.58 pg/dl (Figure 9).

Results of electrolyte ions, urea and creatinine as presented in Figures 10 to 15 show that sodium ion (Na⁺) ranged from 71.00 to 107.55 mEq/L (Figure 10); potassium ion (K⁺) ranged from 4.11 to 6.36 mEq/L) (Figure 11); Chloride ion (CI⁻) ranged from 60.71 to 92.73 mEq/L (Figure 12), bicarbonate ion (HCO₃⁻) ranged from



Figure 1. Hb level of the diabetic rats treated with V. major leaf extract for 28 days.



Figure 2. Haematocrit level of diabetic rats treated with V. major leaf extract for 28 days.

16.54 to 21.35 mmol/L (Figure 13); urea ranged from 38.12 to 56.56 mg/dL (Figure 14) and creatinine raged from 0.01 to 4.36 mg/dL (Figure 15).

The lipid profiles as presented in Figures 16 to 19 reveal that total cholesterol ranged from 69.67 to 123.10 mg/dL (Figure 16), triglyceride ranged from 98.38 to 147.18 mg/dL (Figure 17); HDL cholesterol ranged from 16.34 to 24.01 mg/dL (Figure 18) and LDL cholesterol ranged from 30.60 to 83.53 mg/dL (Figure 19). Liver enzymes, total bilirubin, protein and glucose levels as presented in Figures 20 to 24 show that AST was between 16.17 to 27.15 U/L (Figure 20); ALT ranged 46.93 to 62.10 U/L (Figure 21); ALP ranged 12.71 to 25.31 U/L (Figure 22); total bilirubin ranged from 0.14 to 0.25 mg/dl (Figure 23); and glucose ranged from 224.76 to 306.37 mg/dl (Figure 24).

DISCUSSION

Assessment of haematological parameters can be used to determine the extent of deleterious effect of foreign compounds in the body (Mohammed et al., 2009; Duru et

al., 2012b; Duru et al., 2018). Both plant extracts and free radical from alloxan, on the blood constituents of an animal are amongst the foreign compounds (Mohammed et al., 2009). According to Lebovitz (1994) and Andreoli et al., (1990), blood relating functions are amongst the aberration of diabetes mellitus. Haemoglobin (Hb) levels in groups 3 and 4 rats significantly increased (p<0.05) against the control throughout the number of days of the study. However, Hb in group 2 rats reduced significantly (p<0.05) when compared to the control (Group 1) on the 7th and 14th days, but increased insignificantly (p>0.05) against the control on the 21st and 28th days of the study (Figure 1). Sheela and Augusti (1992) noted that diabetic rats form glyscosylated haemoglobin in results in decreased total haemoglobin. The increase in Hb levels of test rats in the present study could be attributed to the ability of V. major leaf extract to induce Hb production in diabetic condition. Haematocrit levels in rats of test groups 3 and 4 increased significantly (p<0.05) when compared to the control (Group 1). Haematocrit in group 2 increased insignificantly (p>0.05) in test group 2 rats against the control. However, the observed increase



Figure 3. RBC level of diabetic rats treated with V. major leaf extract for 28 days.



Figure 4. WBC level of diabetic rats treated with V. major leaf extract for 28 days.



Figure 5. Lymphocyte level of diabetic rats treated with V. major leaf extract for 28 days.



Figure 6. Monocyte level of diabetic rats treated with V. major leaf extract for 28 days.



Figure 7. Basophils level of diabetic rats treated with V. major leaf extract for 28 days.



Figure 8. Mean cell volume (MCV) level of diabetic rats treated with V. major leaf extract for 28 days.

became significant (p<0.05) on the 28^{th} day of the study (Figure 2). Saliu et al., (2012) attributed decrease in haematocrit in diabetic condition to cellular damage on the erythrocyte membrane as a result of oxidative stress

by agent of induction. Haematocrit measures the percentage by volume of packed red blood cells (RBCs) in a whole blood sample after centrifugation. Only group 2 rats had significantly increased (p < 0.05) RBC level



Figure 9. Mean corpuscular haemoglobin concentration (MCHC) level of diabetic rats treated V. major leaf extract for 28 days.







Figure 11. K⁺ level of diabetic rats treated with *V. major* leaf extract for 28 days.



Figure 12. Cl⁺ level of diabetic rats treated with V. major leaf extract for 28 days.



Figure 13. HCO₃⁻ (mmol/L) of the rats treated with V. major leaf extract for 28 days.



Figure 14. Urea level of diabetic rats treated with V. major leaf extract for 28 days.

when compared to the control on the 7th day. RBC levels in groups 2 and 4 increased significantly (p<0.05) against the control on the 14th day, while all the test group rats had significantly increased (p<0.05) RBC levels on the 21st and 28th days of the study (Figure 3). According to Rao et al., (2003), reactive O₂ species generated during alloxan metabolism is implicated in red cell damage. The increase in RBC levels of test rats could give credence to the increased haematocrit, which may be linked to the

influence of *V. major* leaf extract on Hb production in diabetic condition. Edet et al., (2011) noted that alloxan diabetogenesis may cause perturbation in the bone marrow stem cells. This observation was experienced in the present study where the level of WBC in all the test rats significantly increased (p<0.05) when compared to the control throughout the study (Figure 4). White blood cell differentials are indicators of the ability of an organism to eliminate infection (Duru et al., 2012a, b;



Figure 15. Creatinine level of diabetic rats treated with V. major leaf extract for 28 days.



Figure 16. Total cholesterol level of diabetic rats treated with V. major leaf extract for 28 days.



Figure 17. Triglyceride level of diabetic rats treated with V. major leaf extract for 28 days.



Figure 18. HDL-cholesterol level of diabetic rats treated with V. major leaf extract for 28 days.



Figure 19. LDL-cholesterol level of diabetic rats treated V. major leaf extract for 28 day.



Figure 20. AST level of diabetic rats treated with V. major leaf extract for 28 days.

Amadi et al., 2013; Ugbogu et al., 2016). Levels of lymphocytes, monocytes and basophils increased significantly (p<0.05) in test rats when compared to the

control (Figure 5 to 7). The observed increase in WBC differential could be indication of improved immune defence in test rats, and could be attributed to the



Figure 21. ALT level of diabetic rats treated with V. major leaf extract for 28 days.



Figure 22. ALP level of diabetic rats after treated with V. major leaf extract for 28 days.



Figure 23. Total bilirubin level of diabetic rats treated with V. major leaf extract for 28 days.

presence *V. major* leaf extract. Mean cell volume (MCV) and Mean corpuscular heamoglobin concentration (MCHC) are related to individual red blood cells (Adebayo et al., 2005, 2010, Duru et al., 2018). MCV level of test

rats in groups 3 and 4 increased significantly (p<0.05) against the control on the 7th and 14th days. MCV of group 2 rats reduced significantly (p<0.05) when compared to the control on the 7th and 14th days



Figure 24. Glucose level of diabetic rats treated with V. major leaf extract for 28 days.

(Figure 8). The trend however reversed on the 21st and 28th days of the study. All the test rats had significantly reduced (p<0.05) MCV level on the 21st day while only test groups 3 and 4 rats reduced significantly (p<0.05) against the control on the 28th day (Figure 8). MCHC increased insignificantly (p>0.05) in rats test groups 3 and 4 on the 7th day against the control. MCHC was also insignificantly affected (p>0.05) in all the test rats against the control on the 14th and 28th days of the present study (Figure 9). It has been noted that several haematological changes affecting the red blood cells (RBCs), white blood cells (WBCs) and the coagulation factors are directly associated with diabetes mellitus (Wong and Lin, 1998; Bunza and Dallatu, 2017). Mansi and Lahham (2008) revealed that various hematological parameters and the immune system were reported to be altered during the course of diabetes. Hence, the observed all improvements on haematological parameters of the test rats against the control could be attributed the treatment with V. major leaf extract.

Electrolyte ions, urea and creatinine are indices for evaluation of excretory organ (Duru et al., 2012b, 2013). Pecoits-Filho et al., (2016) noted that diabetic kidney disease is one of the most frequent and dangerous complications of diabetes mellitus. High level of blood sugar, genetics and blood pressure are among the factors that facilitate renal organ disease in diabetic condition. Na⁺ reduced significantly (p<0.05) in test rats against the control on the 7th and 28th days; while on the 14th and 21st days, only rats in test group 4 had insignificant reduction when compared to the control (Figure 10). K⁺ in test group 2 rats reduced significantly (p<0.05) against the control on the 7th day; all the test group rats had significantly reduced K⁺ when compared to the control on the 14th day (Figure 11); whereas only test groups 3 and 4 rats had significantly (p<0.05) reduced K⁺ against the control on the 21st and 28 days of

the study (Figure 11). Cl⁻ in the test rats reduced (p<0.05) significant against the control (Figure 12). HCO₃ significantly reduced (p<0.05) in test group 4 rats on the 7th and 28 davs against the control (Figure 13); those of test group 2 and 4 reduced significantly (p<0.05) on the 4th day against the control; on the 21st day, all the test rats had significantly reduced (p<0.05) HCO₃ when compared to the control (Figure 13). Urea and creatinine are both important parameters of renal function. The levels of urea and creatinine in test rats reduced significantly (p<0.05) against the control (Figures 14 and 15). The observed trend followed by electrolyte ions, urea and creatinine in test rats against the control in the present study, may be attributed to V. major leaf extract trying to salvage the excretory organs of alloxan battered diabetic rats.

Diabetic dyslipidaemia is a common experience in diabetic condition. It is normally the aetiology of premature coronary heart disease and atherosclerosis. There is increasing evidence that dyslipidaemia in diabetes is associated with increased risk of cardiovascular disease, which is the leading cause of death in patients with type 2 diabetes (Ronald, 2004). Levels of total cholesterol (Figure 16), triglyceride (Figure 17), and LDL-cholesterol (Figure 19) reduced (p<0.05) significantly in test rats when compared to the control for the number days of study. Cholesterol is needed by the body to maintain healthy cell status (Njoku et al., 2017). However, its high level in the body leads to coronary artery disease (Njoku et al., 2017). Triglyceride is a type of fat found in the blood and its high levels are related to higher risk of heart and blood vessel disease (Duru et al., 2014). Decrease in LDL-cholesterol, the bad cholesterol has been linked to reduced risk of coronary heart diseases (Glew, 2006; Shen, 2007; Duru et al., 2017); while increase in high levels of HDL-cholesterol are linked to a reduced risk of heart and blood vessel disease

(Shen, 2007). HDL-cholesterol of diabetic treated rats increased significantly (p<0.05) against the control on the 7th and 21 days of the study (Figure 18). The observations made on lipid profile parameters in this study could be indication of the salvage power of *V. major* leaf extract on diabetic dyslipidaemia.

Different authors have noted that hepatic dysfunction is abnormalities of diabetic condition among the (Yakhchalian et al., 2018). There are increasing evidence on occurrence of hepatic necrosis and inflammation induced by diabetes mellitus pathogenesis (West, 2000; Kyle et al., 2002). Inflammation and liver injuries that lead to damage of hepatocytes are also ascribed to agent of diabetic induction in experimental animals such as alloxan and streptozotocin (Jacobs et al., 2004; Zafar et al., 2009; Pagana and Pagana, 2013). AST, ALT and ALP liver enzymes reduced significantly (p<0.05) in test rats against the control (Figures 20 to 22). Total bilirubin reduced insignificantly (p>0.05) in test rats against the control on the 7th day, the observed reduction became significant (p<0.05) in test groups 2 and 4 rats on the 14th and 21st days whereas the significant reduction in total bilirubin manifested in all the test rats on the 28th day (Figure 23). Level of glucose in test rats reduced significantly p<0.05) when compared to the control (Figure 24). Many studies have reported that single diabetogenic dose of alloxan or streptozotocin showed an increase in glucose, ALT and AST levels (Zafar et al., 2009, Zafar and Nagvi 2010; Ahmed et al., 2012). The observed reduction in AST, ALT, ALP and glucose in test rats in this could be attributed to the ability of V. major leaf extract to improve on the hepatic necrosis and inflammable associated with diabetic condition.

Conclusion

Diabetic rats treated with *V. major* leaf extract in this study, showed significant improvements on those associate problems of diabetes which include anaemia, dyslipidaemia, and hepatic necrosis and inflammation. Rats treated with the leaf extract also showed reduced glucose level (Hypoglycemia). From the observations of this study, extract from leaf of *V. major* may be effective against diabetes and some of its associate problems. There is need to urgently extend the scope of the present study to accommodate the isolation of the active ingredients or compounds that could be responsible for these actions in *V. major* leaf.

CONFLICT OF INTERESTS

The Authors declare no conflict of interests and are all aware that the article has been submitted to African Journal of Biotechnology. The Authors also declare that extra authors may be added as the case maybe.

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REFERENCES

- Adebayo AH, Abolaji AO, Opata TK, Adegbenro IK (2010). Effects of ethanolic leaf extract of *Chrysophyllum albidum* G. on biochemical and haematological parameters of albino wistar rats. African Journal of Biotechnology9(14):2145-2150.
- Adebayo AH, Aliyu R, Gatsing D, Garba IH (2006). The effects of ethanolic leaf extract of *Commiphora africana* (Burseraceae) on lipid profile in rats. International Journal of Pharmacology 2(6):618-622.
- Adebayo JO, Adesokan AA, Olatunji LA, Buoro DO, Soladoye AO (2005). Effect of ethanolic extract of *Bougainvillea spectabilis* leaves on haematological and serum lipid variables in rats. BIOKEMISTRI 17(1):45-50.
- Agomuo E, Duru M, Amadi B, Amadi P, Ugwokaegbe P (2017). Effect of Caffeine on Some Selected Biochemical Parameters Using Rat Model. https://doi.org/10.1155/2017/9303276
- Ahmed O, Mahmoud A, Abdel-Moneim A, Ashour M (2012). Antidiabetic effects of hesperidin and naringin in type 2 diabetic rats. Diabetologia Croatica 41(2):53-67.
- Amadi BA, Agomuo EN, Duru MKC (2013). Toxicological studies of *Asmina triloba* leaves on haematology, liver, kidney using rat model. International Science Research Journal 4(2):11-17.
- Amadi BA, Duru MKC, Agomuo EN (2012). Chemical profiles of leaf, steam, and flower of *Ageratum conyzoides*. Asian Journal of Plant Science and Research 2(4):428-432.
- Alexander RR, Griffths JM (1993). Haematocrit in Basic Biochemical Methods, John Wiley & Sons, New York, NY, USA, 2nd Edition.
- Andreoli TE, Carpenter CC, Plum F, Smith LH (1990). Diabetes mellitus. In: Cecil Esssential of Medicine, Dyson, J. (Ed.). 2nd Edn. W. B. Saunders, Philadelphia, PA., USA pp. 496-505.
- Babson LA, Greeley SJ, Coleman CM, Philips GD (1966). Serum alkaline phosphatase determination. Clinical Chemistry 12:482-490.
- Bastaki S (2005). Review: Diabetes mellitus and its treatment. International Journal of Diabetes and Metabolism. 13(3):111-134
- Blamey M, Grey-Wilson C (1989). Flora of Britain and Northern Europe. Hodder and Stoughton.
- Bnouham M, Ziyyat A, Mekhfi H, Tahri A, Legssyer A (2006). Medicinal plants with potential anti-diabetic activity- A review of ten years of herbal research (1990-2000). International Journal of Diabetes and Metabolism 14(1):1-25.
- Bunza FU, Dallatus MK (2017). Hematological indices in alloxan-Induced diabetic rats: effect of supplementation with the antioxidant dimethyl sulfoxide. Asian Journal of Medicine and Health 8(1):1-7.
- Demoz MS, Gachoki KP, Mungai KJ, Negusse BG (2015). Evaluation of the anti-diabetic potential of the methanol extracts of *Aloe camperi*, *Meriandra dianthera* and a Polyherb. Journal of Diabetes Mellitus 5(4):267-276
- Doumas BT, Perry BW, Sasse EA, Straumfjord JV (1973). Standardization in bilirubin assays: evaluation of selected methods and stability of bilirubin solutions. Clinical Chemistry 19(9):984-93.
- Duru M, Amadi B, Amadike U, Adindu E (2014). Éffect of "udu", an antimalarial herbal preparation on visceral organ weight and blood lipid profiles in wistar rats. JPC 8:1-7. www.arpapress.com/Volumes/JPCS/Vol8/JPCS_8_01.pdf
- Duru MKC, Agomuo EN, Amadi BA (2012b). Biochemical studies on "udu" an antimalarial concoction used in Umunchi village, Isiala Mbano L.G.A of Imo State, Nigeria. Continental Journal of Pharmacology and Toxicology Research 5(2): 28-34.
- Duru MKC, Amadi BA, Amadi CT, Lele KC, Anudike JC, Chima-Ezika OR, Osuocha K (2012a). Toxic effect of *Carica papaya* bark on body weight, haematology, and some biochemical parameters. Biokemistri 24(2):67-71.

- Duru M, Nwadike C, Ozougwu J, Eboagwu I (2017). Bioactive constituents of *Pleurotus squarrosulus* (Mont.) singer and effect of its dietary incorporation on body/organ weights and lipid profile levels of rats placed on high cholesterol diet. Academic Journal Chemistry 2(4):28-37.
- Duru M, Constance N, Ahamefula E, Caleb N, Ijeoma E, Odika P, Samuel N, Chieme C (2018). Evaluation of nutritional, anti-nutritional and some biochemical studies on *Pleurotus sqarrosulus* (Mont) singer using rats. African Journal of Biochemistry Research 12(20):7-27.
- Duru M, Amadike U, Amadi B (2013). Effect of *Solanum macrocarpon* fruit on haematology, hepatic and renal function. Advances in Biochemistry 1(2):28-32.
- Edem OD (2009). Hypoglycemic effects of ethanolic extract of alligator pear seed (*Persea americana* Mill) in rats. European Journal of Scientific Research 33(4):669-678.
- Edet EE, Akpanabiatu MI, Uboh FE, Edet TE, Eno AE, Itam EH, Umoh IB (2011). *Gongronema latifolium* crude leaf extract reverses alterations in hematological indices and weight loss in diabetic rats. Journal of Pharmacology and Toxicology 6(2):174-181.
- Firdous SM (2014). Phytochemicals for treatment of diabetes EXCLI Journal 13:451-453.
- Forrester RL, Watafe LJ, Silverman DA, Pierre KJ (1979). Enzymatic method for the determination of CO2 in serum. Clinical Chemistry 22(2):243–245.
- Friedwald TW, Federickson DS, Levy RJ (1972). LDL cholesterol estimation. Clinical Chemistry 18:499-501.
- Glew RH (2006). Lipid metabolism II: Pathways of metabolism of special lipids. In: Devlin TM (ed) Textbook of biochemistry with clinical correlations, 6th edn. Wiley Liss, New Jersey pp. 695-741.
- Heinegård D, Tiderström G (1973). Determination of serum creatinine by a direct colorimetric method. Clinica Chimica Acta. 43(3):305–310.
- Islam D, Huque A, Mohanta LC, Lipy EP, Rahman MN, Sultana SA, Prodhan UK (2017). Studies on the hypoglycemic and hypolipidemic effects of *Nelumbo nucifera* leaf in long-evans rats. Journal of Diabetes Mellitus 7(3):55-70.
- Jacobs SD, Demott RW, Oxley KD (2004). Laboratory test handbook. 3ed; Lexi comp.
- Jung M, Park M, Lee HC, Kang YH, Kang ES, Kim SK (2006). Anti-Diabetic Agents from Medicinal Plants. Current Medicinal Chemistry 13(10):1203-1218.
- Kayarohanam S, Kavimani S (2015). Current Trends of Plants Having Antidiabetic Activity: A Review. Journal of Bioanalysis & Biomedicine 7(2):55-65.
- Kesari AN, Kesari S, Singh SK, Gupta RK, Watal G (2007). Studies on the glycemic and lipidemic effect of *Murraya koenigii* in experimental animals. Journal of Ethnopharmacology 112(2):305-311.
- Kyle R, Katzmann J, Lust J, Dispenzieri A (2002). Clinical indications and applications of electrophoresis and immunofixation. In Manual of Clinical Laboratory Immunology. 6ed Washington DC; ASM Press.
- Larsen K (1971). Creatinine assay by a reaction-kinetic principle. Clinica Chimica Acta 41:209-217.
- Lebovitz HE (1994). Oral anti-diabetic agents. In: Joslins diabetes mellitus, Khan, C.R. and G.C. Weir (Eds.). 13th Edn. Lea and Febiger, Philadelphia, PA., USA pp. 508-524.
- Lorke D (1983). A new approach to practical acute toxicity test. Institute Fur Toxicologie, Bayer AG, Friedrich – Ebert- Strasse; 217, D-5600.
- Mansi K, Lahham J (2008). Effects of Artemisia sieberi Besser (a. herba-alba) on heart rate and some hematological values in normal and alloxan-induced diabetic rats. Australian Journal of Basic and Applied Sciences 4(2):57-62.
- Marles RJ, Farnsworth NR (1995). Antidiabetic plants and their active constituents. Phytomedicine 2(2):137-189.
- Mohammed A, Adelaiye AB, Bakari AG, Mabrouk MA (2009). Antidiabetic and some hematological effects of ethyl acetate and nbutanol fractions of *Ganoderma lucidum* aqueous extract in alloxaninduced diabetic wistar rats. International Journal of Medicine and Medical Sciences 1(12):530-535.
- Mussie SD, Kareru PG, Keriko JM, Berhane GN (2015). Ethnobotanical survey and preliminary phytochemical studies of plants traditionally used for diabetes in Eritrea. European Journal of Medicinal Plants 9(2):1-11.

- National Institute of Health (1985). Guide for the care and use of laboratory animals. U.S. Department of health education and welfare. Washington, D.C.: NIH Publication.
- Nawrot P, Jordan S, Eastwood J, Rotstein J, Hugenholtz A, Feeley M (2003). Effects of caffeine on human health. Food Additives and Contaminants 20(1):1-30.
- Njoku S, Duru M, Akubugwo E, Ozougwu J, Nwadike C (2017). The influence of cigarette smoking and moderate beer consumption on selected biochemical indices in male volunteers. Journal of Biology and Nature 7(4):177-189.
- Nole T, Lionel TDW, Cedrix TFS, Gabriel AA (2016). Ethnomedical and Ethnopharmacological Study of Plants Used For Potential Treatments of Diabetes and Arterial Hypertension by Indigenous People in Three Phytogeographic Regions of Cameroon. Diabetes Case Reports 1(2):1-9. doi: 10.4172/2572-5629.1000110.
- Ogbonnia SO, Odimegwu JI, Enwuru VN (2008). Evaluation of hypoglycemic and hypolipidemic effects of ethanolic extract of *Treculia Africana* Decne and *Bryophyllum pinnatum* Lam. and their mixture on streptozotocin (STZ)-induced diabetic ratsAfrican Journal of Biotechnology 7(15):2535-2539.
- Onu M, Igwemma AA (2000). Applied statistical techniques for business and basic sciences, Skillmark Media, 2nd edition.
- Pagana K, Pagana T (2013). Mosby's manual of diagnostic and laboratory tests. Elsevier Health Sciences.
- Patel DK, Prasad SK, Kumar R, Hemalatha S (2012). An overview on anti-diabetic medicinal plants having insulin mimetic property. Asian Pacific Journal of Tropical Biomedicine 2(4):320-330.
- Rao G, Kamath U, Raghothama C, Pradeep KS, Rao P (2003). Maternal and fetal indicators of oxidative stress in various obstetric complications. Indian Journal of Clinical Biochemistry 18(2):80-86.
- Reitman S, Frankel S (1957). A Colorimetric Method for the Determination of Serum Glutamic Oxalacetic and Glutamic Pyruvic Transaminases. American Journal of Clinical Pathology 28(1):56-63.
- Ronald MK (2004). Lipids and lipoproteins in patients with type 2 diabetes. Daibetes Care 27(6):1496-1504.
- Pecoits-Filho R, Abensur H, Betônico CCR, Machado AD, Parente EB, Queiroz M, Salles JEN, Titan S, Vencio S (2016). Interactions between kidney disease and diabetes: dangerous liaisons. Diabetology & Metabolic Syndrome 8:50.
- Searcy RL, Reardon JE, Foreman JA (1967). A new photometric method for serum urea nitrogen determination. American Journal of Medical Technology 33(1):15-20.
- Saliu JA, Elekofehinti OO, Komolafe K, Oboh G (2012). Effects of some green leafy vegetables on the haematological parameters of diabetic rats. Journal of Natural Product and Plant Resources 2(4):482-485.
- Sheela CG, Augusti KT (1992). Antidiabetic effects of S-allyl cysteine sulphoxide isolated from garlic *Allium sativum* Linn. Indian Journal of Experimental Biology 30(6):523-526.
- Shen GX (2007). Lipid disorders in diabetes mellitus and current management. Current Pharmaceutical Analysis 3(1):17-24.
- Shrivastava SR, Shrivastava PS, Ramasamy J (2013). Role of self-care in management of diabetes mellitus. Journal of Diabetes & Metabolic Disorders 12(1):14.
- Trinder P (1969). Determination of glucose in blood using glucose oxidase with an alternative oxygen acceptor. Annals of Clinical Biochemistry 6:24-27.
- Tsabang N, Fongnzossié E, Keumeze V, Jiofack R, Njamen D, Sonwa DJ Nguelefack BT (2017). Ethnomedical and Ethnopharmacological Study of Plants Used by Indigenous People of Cameroon for The Treatments of Diabetes and its Signs, Symptoms and Complications. Journal of Molecular Biomarkers and Diagnosis 8:310. doi: 10.4172/2155-9929.1000310
- Tsabang N, Ngah N, Estella FT, Agbor GA (2016). Herbal medicine and treatment of diabetes in Africa: case study in Cameroon. Diabetes Case Reports 1(2):112. doi: 10.4172/2572-5629.1000112.
- Ugbogu AE, Okezie E, Uche-Ikonne C, Duru M, Atasie OC (2016). Toxicity evaluation of the aqueous stem extract of Senna alata in wistar rats. American Journal of Biomedical Research 4(4):80-86.
- West I (2000). Radicals and oxidative stress in diabeties. Diabetic Medicine 17(3):171-180.
- Wong CK, Lin CS (1998). Remarkable response of lipoid proteinosis to oral dimethyl sulphoxide.. British Journal of Dermatology 119(4):541-

544.

- Yakhchalian N, Mohammadian N, Hatami K, Nosrati H, Yousofvand N (2018). Hematological and serum biochemical analysis of streptozotocin-induced insulin dependent diabetes mellitus in male adult wistar rats. https://doi.org/10.1101/359844
- Zafar M, Naqvi S (2010). Effects of STZ-induced diabetes on the relative weights of kidney, liver and pancreas in albino rats: a comparative study. International Journal of Morphology 28(1):135-142.
- Zafar M, SN-u-H N, Ahmed M, Khani ZAK (2009). Altered liver morphology and enzymes in streptozotocininduced diabetic rats. International Journal of Morphology 27(3):719-25.