

*Full Length Research Paper*

# The inhibitory effect of *Curcuma longa* extract on telomerase activity in A549 lung cancer cell line

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Telomerase is reactivated in lung cancer cells, the most prevalent cancer worldwide, but not normal cells. Therefore, targeting it, preferably with natural compounds derive from medicinal plant such as curcumin, could have important effect on treatment of lung cancer. Curcumin, derived from *Curcuma longa* rhizome, has many anti-cancer activities. Therefore, the main objective of current work was to study inhibitory effect of *C. longa* total extract on telomerase in A549 lung cancer cell line as *in vitro* model of lung cancer. First, total extract of *C. longa* were prepared by n-hexane, methanol and dichloromethane. Then, cytotoxic effect of n-hexane phase was studied on A549 lung cancer cell line with 24, 48 and 72 h MTT assays. Finally, after determination of IC<sub>50</sub>, cells were treated with n-hexane extract and TRAP (Telomeric Repeat Amplification Protocol) assay was done to measure amount of relative telomerase inhibition by the extract. Data analysis showed that n-hexane extract of *C. longa* has dose-dependent cytotoxic effect on A549 lung cancer cell line with IC<sub>50</sub> = 0.23 - 0.28 mg/ml and the extract inhibits telomerase activity with dose-dependent manner. In conclusion, n-hexane extract of *C. longa* has cytotoxic and telomerase-inhibitory effect on cell line A549 and could be exploited as potential source for developing novel drugs against lung cancer.

**Key words:** Telomerase, lung cancer, *Curcuma longa*, A549 lung cancer cell line.

## INTRODUCTION

Telomerase is a ribonucleoprotein reverse transcriptase which maintains telomeric ends of eukaryotic chromosomes during DNA replication. It has been shown that telomerase is active in more than 85% of cancers including lung cancer but not normal cells (Lingner et al., 1995; Kim, 1997). On the other hand, lung cancer is the most prevalent cancer worldwide (Jemal et al., 2007). Therefore, targeting the telomerase in this cancer could be promising step in its treatment. Different agents have been proposed for telo-merase inhibition (Strahl and Blackburn, 1996; Damm et al., 2001; Zou et al., 2006; Massard et al., 2006; Nakamura et al., 2005; Mizushima et al., 2000; Yeo et al., 2005). But, they have undesirable

complications (Mittal et al., 2004) and it is necessary to seek alternative telomerase-inhibiting agents, preferably natural compounds such as curcumin.

*Curcuma longa* is a medicinal plant which its constituents, especially curcumin, has diverse anti-cancer properties including anti-telomerase action (Sanjay et al., 2007; Anand et al., 2008). Anti-telomerase activity of pure curcumin has been studied previously in some cancer cells other than lung cancer cells (Cui et al., 2006; Chakraborty et al., 2006; Ramachandran et al., 2002; Singh and Singh, 2009; Mukherjee et al., 2007). However, to date any study for investigating the effect of curcuma total extract on telomerase in the lung cancer cells remains unperformed. Therefore, the main aim of current work was to study the effect of *C. longa* total extract on telomerase activity in A549 lung cancer cell line as *in vitro* model of lung cancer for its possible therapeutic application in the future.

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## MATERIALS AND METHODS

### Extract preparation

100 g powder of *C. longa* rhizome was dissolved in 200 ml n-hexane and solution was shaken for 4 h at 45°C. Then, supernatant was transferred to a tube. This step was performed three times. In the next step, residue of n-hexane extraction was dissolved in 200 ml dichloromethane instead of n-hexane and same steps repeated using dichloromethane. In the next step, debris of previous extraction step with dichloromethane was dissolved in 200 ml methanol and at the same way supernatant was collected. Finally, solvents of all three phases were dried by rotatory-evaporator and remained powders were stored at -20°C until used.

### Selection of appropriate phase

Based on the fact that Curcuminoids have maximum absorbance at  $\lambda_m = 420 - 430$  nm (Jayaprakasha et al., 2005), the curcuminoids content of n-hexane, dichloromethane and methanol phases of extract was determined spectrophotometrically. Briefly, ODs of 0.038, 0.0304, 0.019, 0.0152, 0.0076 and 0.0019 mg/ml solutions of pure curcumin (sigma) dissolved in 10% DMSO and ODs of 0.0038 mg/ml solutions of extract phases in 10% DMSO were determined in  $\lambda_m = 420$  nm and  $\lambda_m = 430$  nm with three times reading in the spectrophotometer. Then, standard curve was plotted using mean of pure curcumin OD $\times$ 100. Finally, relative curcumin content of phases was determined according to standard curve.

### Cell culture and cytotoxicity

A549 lung cancer cell line (kindly dedicated from pharmaceutical nanotechnology research center, Tabriz University of Medical Sciences, Tabriz, Iran) were cultured in RPMI1640 (Gibco, Invitrogen, UK) supplemented with 10% heat-inactivated fetal bovine serum (FBS) (Gibco, Invitrogen, UK), 2 mg/ml sodium bicarbonate, 0.05 mg/ml penicillin G (Serva co, Germany), 0.08 mg/ml streptomycin (Merck co, Germany) and incubated in 37°C with humidified air containing 5% CO<sub>2</sub>. After culturing sufficient amount of cells, cytotoxic effect of n-hexane extract was studied by 24, 48 and 72 h MTT assays (Carmichael et al., 1987). Briefly, 1000 cell/well were cultivated in a 96 well plate. After 24 h incubation in 37°C with humidified atmosphere containing 5% CO<sub>2</sub>, cells were treated with serial concentrations of n-hexane extract of *C. longa* (0 mg/ml to 0.57 mg/ml) for 24, 48 and 72 h in the quadruplicate manner as cells which received 0 mg/ml extract + 200  $\mu$ l culture medium containing 10% DMSO served as control. After incubation, the medium of all wells of plate were exchanged with fresh medium and cells were leaved for 24 h in incubator. Then, medium of all wells were removed carefully and 50  $\mu$ l of 2 mg/ml MTT (Sigma co, Germany) dissolved in PBS was added to each well and plate was covered with aluminum foil and incubated for 4.5 h. After removing of wells' content, 200  $\mu$ l pure DMSO was added to wells. Then, 25  $\mu$ l Sorensen's glycine buffer was added and immediately absorbance of each well was read in 570 nm using ELx800 Microplate Absorbance Reader (Bio-Tek Instruments) with reference wavelength of 630 nm.

### Cell treatment

After determination of IC<sub>50</sub>,  $1 \times 10^6$  cells were treated with serial concentrations of n-hexane extract (0.028, 0.057, 0.114, 0.142, 0.171 and 0.199 mg/ml). For control cells, the same volume of 10%

DMSO without n-hexane extract was added to flask of control cells. Then, culture flasks were incubated in 37°C containing 5% CO<sub>2</sub> with humidified atmosphere incubator for 24 h exposure duration.

### Telomeric repeat amplification protocol (TRAP) assay

After 24 h, cells were harvested, their total protein were extracted according to instructions of TeloTAGGG Telomerase PCR ELISA<sup>PLUS</sup> package (Cat. No. 12 013 789 001, Roche Applied Science, Germany) and quantity of total protein was determined for each sample by Quick Start™ Bradford Protein Assay (Cat. No. 500-0206, Bio-Rad Laboratories, Inc., USA). Finally, for determining relative telomerase activity in the samples, we used Holt PCR-ELISA based TRAP assay method (Holt et al., 1996) according to instructions of TeloTAGGG Telomerase PCR ELISA<sup>PLUS</sup> kit and our previous work (Zarghami and Asadi, 2007). First, negative control for each sample was constructed by heating at 95°C for 20 min. Then, 50  $\mu$ l reaction mixture (5  $\mu$ g extracted total protein, 5  $\mu$ l dNTPs, 60 ng from each primer, 5  $\mu$ l Internal Standard and 16  $\mu$ l double-distilled nuclease free water) for each sample was added in a new 0.5 ml microtube.

Then, microtubes were transferred to Eppendorf PCR thermal cycler and a combinatory elongation/PCR reaction was performed [1 cycle for 20 min at 25°C (primer elongation); 1 cycle for 20 min at 95°C (telomerase inactivation); 30 cycles for 30 s at 94°C, 30 s at 50°C and 90 s at 72°C and 1 cycle for 10 min at 72°C (PCR)]. Then, PCR amplicons were detected by ELISA-based hybridization. For this purpose, 3  $\mu$ l PCR products were immobilized on streptavidin coated modules and incubated for 2 h at room temperature. Then, module wells were washed three times and horseradish peroxidase-conjugated anti-digoxigenin antibody was added to wells. After that, substrate solution containing 3,3',5,5' tetra methyl benzene (TMB) was added to wells, wells incubated in room temperature for 20 min for color development and more color development was stopped. Finally, absorbance of developed blue color was measured at 450 nm by STAT-FAX 2100 ELISA reader (with a reference wavelength of 630 nm).

### Data analysis

For data analysis of MTT assays, mean OD of each well was calculated. Then, OD of tests and controls was corrected by subtracting mean OD of tests and controls from mean OD of blanks (medium alone). Then, percent of viability of control was calculated according to this formula: percent of viability of control = corrected mean OD of test/corrected control $\times$ 100. Finally, graph of results was plotted using SPSS 16.0 and IC<sub>50</sub> (half maximal inhibitory concentration) of n-hexane extract on A549 was calculated.

Relative telomerase activity percent (%RTA) of each sample was calculated according to instructions of TeloTAGGG Telomerase PCR ELISA<sup>PLUS</sup> kit. Then, %RTA of control cell was calculated for each sample and related graph was plotted using SPSS 16.0.

## RESULTS

Data analysis of spectrophotometric measurement of extracted phases curcuminoids content showed that n-hexane phase has more curcumin than methanol and dichloromethane phases and dichloromethane phase has the lowest content of curcuminoids (Figure 1 and Table 1). Therefore, we investigated the effect of n-hexane extract

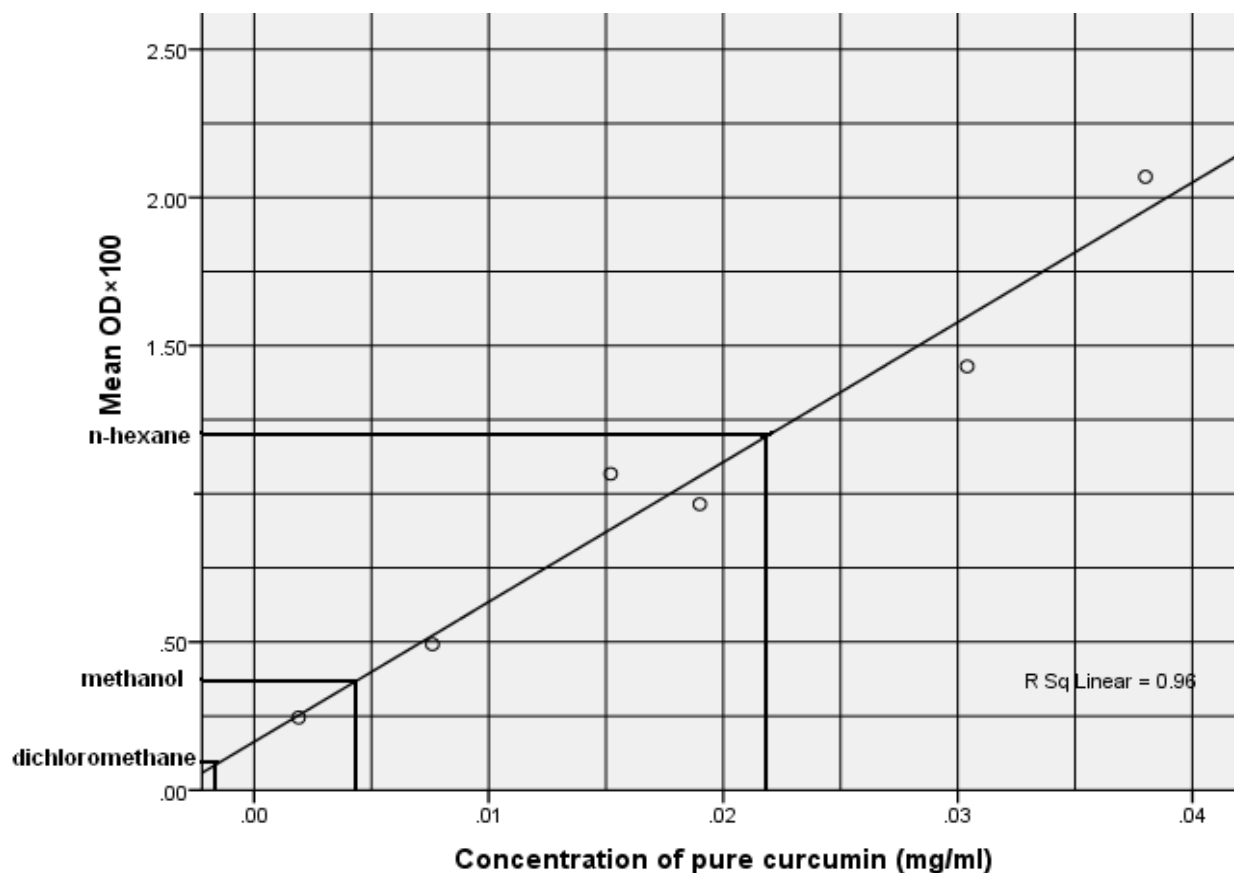


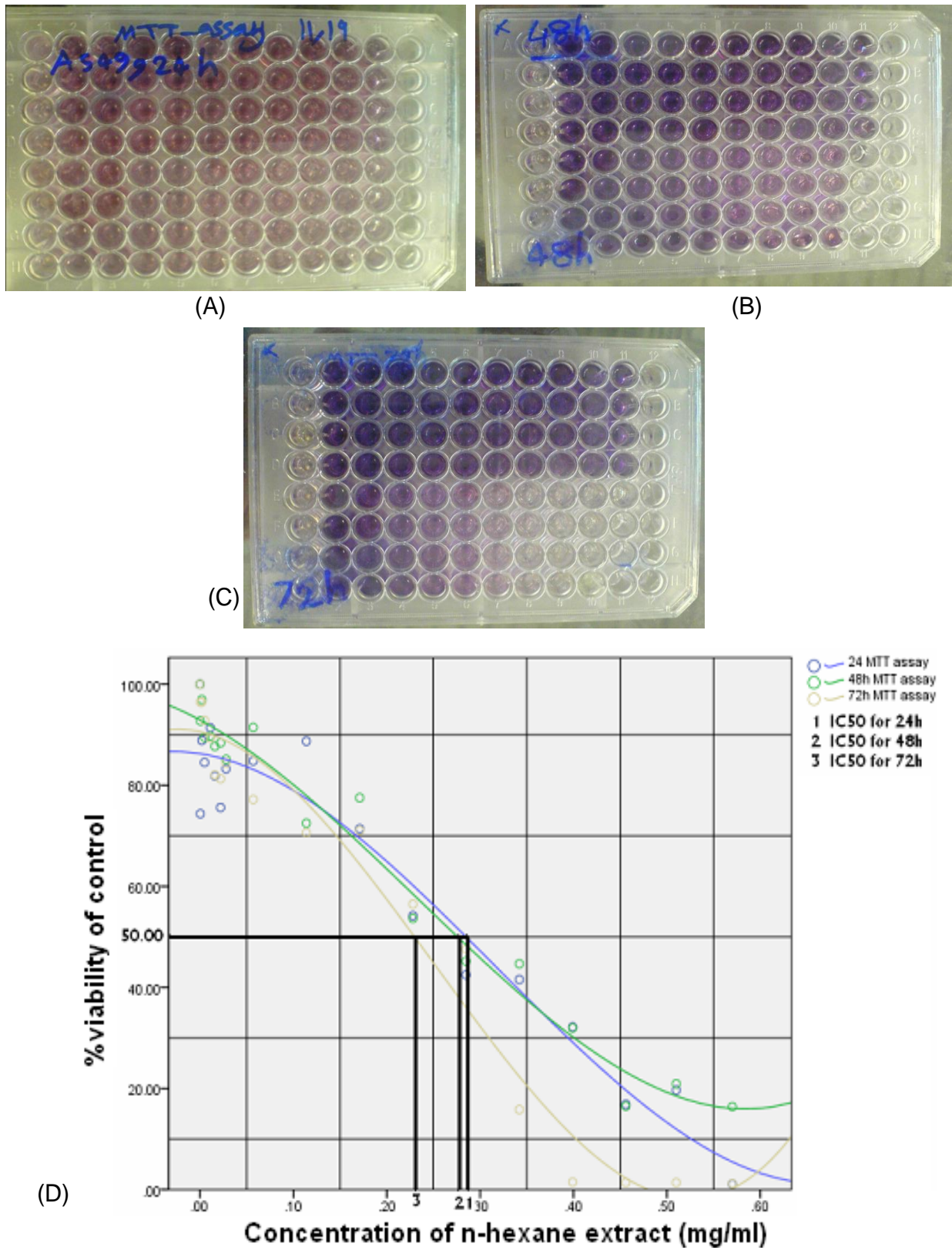
Figure 1. OD<sub>420</sub> and OD<sub>430</sub> of *Curcuma longa* extracts in comparison with pure curcumin (as standard).

Table 1. Mean absorbances of pure curcumin, n-hexane extract, methanol extract and dichloromethane extract solutions in  $\lambda_{\max} = 420$  nm and  $\lambda_{\max} = 430$  nm.

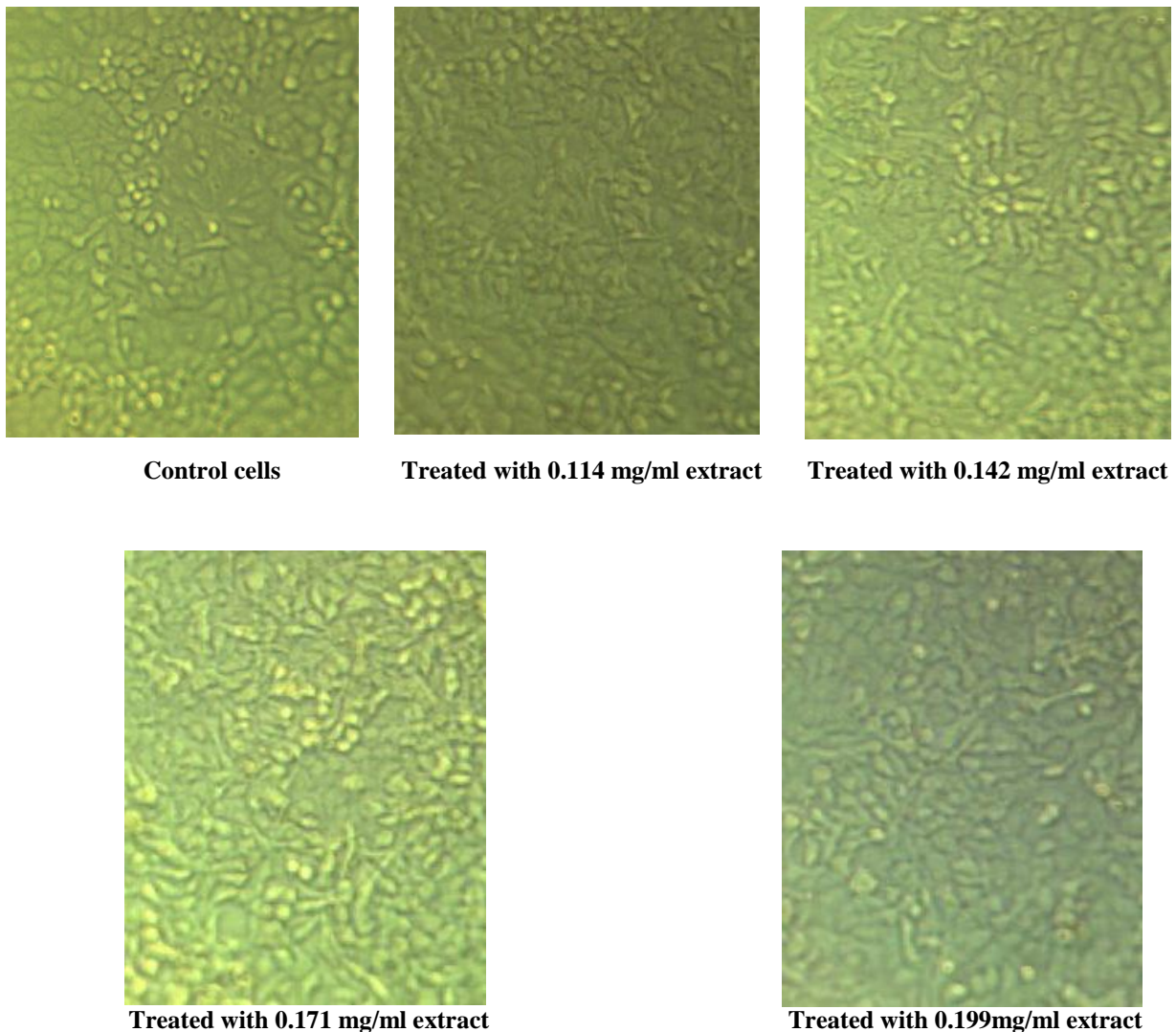
Parameter	Pure curcumin (as standard)						n-hexane extract	Methanol extract	Dichloro-methane extract
	0.0019	0.0076	0.0152	0.019	0.0304	0.038			
Conc. (mg/ml)	0.0019	0.0076	0.0152	0.019	0.0304	0.038	0.0038	0.0038	0.0038
$\lambda_{\max} = 420$ nm	0.0025	0.0049	0.0108	0.0097	0.0143	0.0207	0.0120	0.0038	0.0006
$\lambda_{\max} = 430$ nm	0.0024	0.0050	0.01055	0.0096	0.0143	0.0207	0.0123	0.0037	0.0008
Mean ODx100	0.245	0.492	1.0675	0.965	1.43	2.07	1.215	0.37	0.07

of *C. longa* on telomerase activity in the A549 cell line. Data analysis of cytotoxicity assay showed that IC<sub>50</sub> of n-hexane extract on A549 lung cancer cell line is 0.28, 0.27 and 0.23 mg/ml for 24, 48 and 72 h MTT assays, respectively (Figure 2). Figure 2 shows that IC<sub>50</sub> of n-hexane extract on A549 lung cancer cell line is dose-dependent but not time dependent. Therefore, we investigated telomerase-inhibitory effect of n-hexane extract on A549 for 24 h exposure time (Figure 3). Morphological analysis showed no significant difference between control and treated cells.

Data analysis obtained from TRAP assay showed that n-hexane extract of *C. longa* inhibits telomerase activity dose-dependently and doses higher than 0.114 mg/ml completely inhibit telomerase relative activity (Figure 4, Tables 2 and 3). Table 2 shows that concentration of 0.142 mg/ml is the least effective dose for complete inhibition of telomerase activity in A549 lung cancer cell line. In addition, in this dose telomerase activity was inhibited 46.30 folds in comparison with untreated cells. Therefore, dose of 0.142 mg/ml may be a cut-off point for telomerase inhibition by n-hexane extract in the A549 cell



**Figure 2.** Cytotoxic effect of n-hexane extract on A549 lung cancer cell line after 24 h (A), 48 h (B) and 72 h (C) exposure. (D). IC50 of n-hexane extract of *Curcuma longa* on A549 tumor cell line after 24, 48 and 72 h treatment.



**Figure 3.** Morphological effect of n-hexane extract of *Curcuma longa* on A549 lung cancer cell line after 24 h treatment.

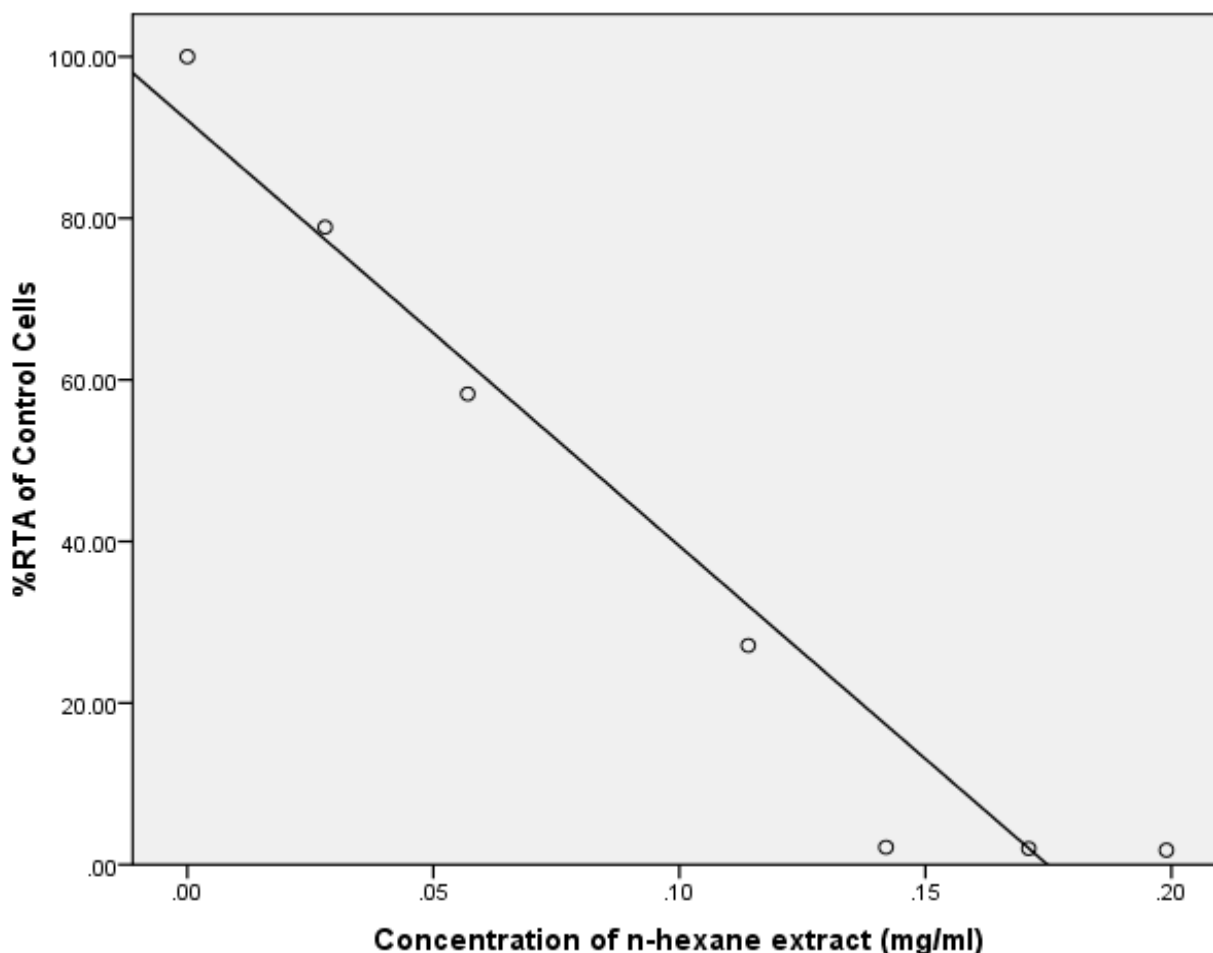
line.

## DISCUSSION

In the present study, total extract used instead of pure curcumin. Because, beside synergistic effects of curcuminoids on each other in a solution (Deshpande and Maru, 1995) and based on Jayaprakasha et al. (2005) report, extraction of pure curcumin from rhizome of *C. longa* is time consumable and needs for using HPLC methods.

In the current work, MTT assay showed that n-hexane extract of *C. longa* has dose-dependent but not time-dependent cytotoxicity on the A549 lung cancer cell line (IC<sub>50</sub> = 0.23 to 0.30 mg/ml). This finding is in accordance

with result of a study (Radhakrishna et al., 2004) in which pure curcumin had dose-dependent but not time-dependent cytotoxicity on A549, but IC<sub>50</sub> of pure curcumin was 0.01841 mg/ml (50  $\mu$ M) for 24 h exposure in that study. In addition, results of same study showed IC<sub>50</sub> = 0.01474 mg/ml (40  $\mu$ M) of pure curcumin on H1299 cells (Radhakrishna et al., 2004). This difference in the IC<sub>50</sub> may be due to purity of curcumin and existence of other substances other than curcumin which may interfere with action of curcumin. Therefore, there is need for separation and further study of n-hexane extraction constituents in the future. Shi et al. (2006) showed that 40  $\mu$ M curcumin inhibits growth of the human epithelium ovarian cancer cell line Ho-8910 in vitro by 52% for 48 h exposure (P < 0.05) (Shi et al., 2006). However, results of current work demonstrated that IC<sub>50</sub> of *C. longa* extract



**Figure 4.** Correlation between different concentrations of n-hexane extract of *Curcuma longa* and telomerase activity in A549 tumor cell line after 24 h treatment.

**Table 2.** Determination of telomerase activity in A549 tumor cells treated with different concentrations of n-hexane extract of *Curcuma longa*.

Dose (mg/ml)	Telomerase activity*
0.0	+
0.028	+
0.057	+
0.114	+
0.142	-
0.171	-
0.199	-

\*The cut off point of being positive (+) and negative (-) was calculated based on following formula:

OD of sample - OD of negative control = A

If  $A > 2 \times$  (OD of negative control), then, telomerase activity considered positive and if not so, activity considered negative.

different finding may be is due to different behavior of two cells in response to curcumin and shows that these cancers may differently response to exposure with curcumin *in vivo*.

Currently there are different types of standard treatment methods for lung cancer patients such as surgery, radiation, chemotherapy, laser therapy, photodynamic therapy (PDT), biologic therapy and gene targeted therapy (Thatcher, 2008). However, there are harsh side effects about all (Geamanu, 2008). Therefore, with regarding to these undesirable effects and result of current study, it seems that *C. longa* may be an appropriate source for developing novel drugs against lung cancer and can be a safer alternative for current treatment regimens.

In conclusion, results of current study showed that n-hexane extract of *C. longa* has inhibitory effect on telomerase in the A549 lung cancer cell line. Inhibition of telomerase by n-hexane extract of *C. longa* in these cells is reported for the first time and even any study, in which

is about 0.26 mg/ml in A549 lung cancer cell line. This

**Table 3.** Comparison Between fold increases of telomerase inhibition by different concentrations of n-hexane extract in A549 lung cancer cell line.

Concentration of n-hexane extract (mg/ml)	RTA of control cells (%)	Fold increase in inhibition of telomerase activity
0.0	100.0	1
0.028	78.9	1.27
0.057	58.26	1.72
0.114	27.13	3.69
0.142	2.16	46.30
0.171	2.0	50
0.199	1.8	55.56

the effect of pure curcumin on the telomerase in the A549 has been investigated, has not been reported to date. However, similar works on other cell lines have been performed with pure curcumin on the other cell lines (Cui et al., 2006; Chakraborty et al., 2006; Ramachandran et al., 2002). Results of these studies have showed that pure curcumin has inhibitory effect on telomerase. Inhibition of telomerase by pure curcumin based on these studies, approve results of current research. Because based on our work n-hexane extract of *C. longa* which contains curcumin, inhibited telomerase dose-dependently in A549 tumor cell lines. Therefore, it can have potentially telomerase-targeting compounds, especially for exploiting in treatment of lung cancer.

Current work demonstrated that n-hexane crude extract of *C. longa* has potent anti-growth effect on A549 and dose-dependently inhibits telomerase in this cell line as in vitro model of NSCLC. Therefore, *C. longa* can be natural potent chemopreventive and chemotherapeutic plant for NSCLC patients and constituents of its n-hexane extract can be appropriate candidate for drug development.

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## REFERENCES

- Anand P, Sundaram C, Jhurani S, Kunnumakkara AB, Aggarwal BB (2008). Curcumin and cancer: an "old-age" disease with an "age-old" solution. *Cancer Lett.* 18(267): 133-64.
- Carmichael J, DeGraff WG, Gazdar AF, Minna JD, Mitchell JB (1987). Evaluation of a tetrazolium-based semiautomated colorimetric assay: assessment of chemosensitivity testing. *Cancer Res.* 47: 936-942.
- Chakraborty S, Ghosh U, Bhattacharyya NP, Bhattacharya RK, Roy M (2006). Inhibition of telomerase activity and induction of apoptosis by curcumin in K-562 cells. *Mutat. Res.* 596(1-2): 81-90.

- Cui SX, Qu XJ, Xie YY, Zhou L, Nakata M, Makuuchi M, Tang W (2006). Curcumin inhibits telomerase activity in human cancer cell lines. *Int. J. Mol. Med.* 18(2): 227-31.
- Damm K, Hemmann U, Garin-Chesa P, Huel N, Kauffmann I, Priepke H, Niestroj C, Daiber C, Enenkel B, Guillard B, Lauritsch I, Müller E, Pascolo E, Sauter G, Pantic M, Martens UM, Wenz C, Lingner J, Kraut N, Rettig WJ, Schnapp A (2001). A highly selective telomerase inhibitor limiting human cancer cell proliferation. *EMBO J.* 20: 6958-6968.
- Deshpande SS, Maru GB (1995). Effects of curcumin on the formation of benzo[a]pyrene derived DNA adducts *in vitro*. *Cancer Lett.* 96(1): 71-80.
- Geamanu A (2008). Efficacy of curcumin in lung cancer: histological evidence. Master of Science thesis, University of Wayne State, Detroit, Michigan.
- Holt SE, Norton JC, Wright WE, Shay JW (1996). Comparison of the telomeric repeat amplification protocol (TRAP) to the new TRAP-eze telomerase detection kit. *Methods Cell Sci.* 18: 237-248.
- Jayaprakasha GK, Jagan L, Mohan R, Sakariah KK (2005). Chemistry and biological activities of *C. longa*. *Trends Food Sci. Tech.* 16: 533-548.
- Jemal A, Siegel R, Ward E, Murray T, Xu J, Thun MJ (2007). *Cancer Statistics, 2007.* *Cancer J. Clin.* 57: 43-66.
- Kim NW (1997). Clinical implications of telomerase in cancer. *Eur. J. Cancer.* 33: 781-786.
- Lingner J, Cooper J, Cech TR (1995). Telomerase and DNA end replication: no longer a lagging strand problem. *Science.* 269: 1533-1534.
- Massard C, Zermati Y, Pauleau AL, Larochette N, Métivier D, Sabatier L, Kroemer G, Soria JC (2006). hTERT: a novel endogenous inhibitor of the mitochondrial cell death pathway. *Oncogene.* 25: 505-514.
- Mittal A, Pate MS, Wylie RC, Tollefsbol TO, Katiyar SK (2004). EGCG down-regulates telomerase in human breast carcinoma MCF-7 cells, leading to suppression of cell viability and induction of apoptosis. *Int. J. Oncol.* 24(3): 703-710.
- Mizushima Y, Ueno T, Oda M, Yamaguchi T, Saneyoshi M, Sakaguchi K (2000). The biochemical mode of inhibition of DNA polymerase beta by alpha-rubromycin. *Biochim. Biophys. Acta.* 1523: 172-81.
- Mukherjee N, Chakraborty S, Ghosh U, Bhattacharyya NP, Bhattacharya RK, Dey S, Roy M (2007). Curcumin-induced apoptosis in human leukemia cell HL-60 is associated with inhibition of telomerase activity. *Mol. Cell. Biochem.* 297(1-2): 31-39.
- Nakamura M, Masutomi K, Kyo S, Hashimoto M, Maida Y, Kanaya T, Tanaka M, Hahn WC, Inoue M (2005). Efficient inhibition of human telomerase reverse transcriptase expression by RNA interference sensitizes cancer cells to ionizing radiation and chemotherapy. *Hum. Gene. Theor.* 16: 859-68.
- Radhakrishna Pillai G, Srivastava AS, Hassanein TI, Chauhan DP, Carrier E (2004). Induction of apoptosis in human lung cancer cells by curcumin. *Cancer Lett.* 208(2): 163-170.
- Ramachandran C, Fonseca HB, Jhabvala P, Escalon EA, Melnick SJ

- (2002). Curcumin inhibits telomerase activity through human telomerase reverse transcriptase in MCF-7 breast cancer cell line. *Cancer Lett.* 184(1): 1-6.
- Sanjay J, Satyaendra S, Satish N, Sumbhate S (2007). Recent trends in Curcuma Longa Linn. *Pharmaco. Rev.* 1(1): 119-128.
- Shi M, Cai Q, Yao L, Mao Y, Ming Y, Ouyang G (2006). Antiproliferation and apoptosis induced by curcumin in human ovarian cancer cells. *Cell Biol. Int.* 30(3): 221-226.
- Singh M, Singh N (2009). Molecular mechanism of curcumin induced cytotoxicity in human cervical carcinoma cells. *Mol. Cell. Biochem.* 325(1-2): 107-119.
- Strahl C, Blackburn EH (1996). Effects of reverse transcriptase inhibitors on telomere length and telomerase activity in two immortalized human cell lines. *Mol. Cell. Biol.* 16: 53-65.
- Thatcher N (2008). First- and second-line treatment of advanced metastatic non-small-cell lung cancer: a global view. *BMC Proc.* 2(2): p. S3.
- Yeo M, Rha SY, Jeung HC, Hu SX, Yang SH, Kim YS, An SW, Chung HC (2005). Attenuation of telomerase activity by hammerhead ribozyme targeting human telomerase RNA induces growth retardation and apoptosis in human breast tumor cells. *Int. J. Can.* 114: 484-489.
- Zarghami N, Asadi J (2007). Effect of Zinc, Selenium and Copper in cell line of breast cancer on telomerase gene expression. *Pharm. Sci. Summer*: pp. 31-38. (Article in Persian).
- Zou L, Zhang P, Luo C, Tu Z (2006). ShRNA-targeted hTERT suppress cell proliferation of bladder cancer by inhibiting telomerase activity. *Cancer Chemother. Pharmacol.* 57: 328-34.