Analgesic and anti-inflammatory activities of propolis obtained from Gboko, Nigeria

Selumun Solomon IPAV¹*, John Ogbaji IGOLI², Terrumun Amom TOR-ANYIIN² and John Vershima ANYAM²

¹Department of Chemical Sciences, University of Mkar, Mkar, Benue State, Nigeria.
²Department of Chemistry, Federal University of Agriculture, Makurdi, Benue State, Nigeria.

Received 7 February, 2022; Accepted 21 June, 2022

The use of propolis for the treatment of various ailments in which inflammation and pain is involved is cross-cultural. This report was designed to investigate the analgesic and anti-inflammatory activities of propolis obtained from Gboko, Benue State using hexane, ethyl acetate, and ethanol solvents. Propolis sample obtained from bee hives was homogenized, extracted and dried at 40°C. The dry extract was prepared in a solution of olive oil for pharmacological and toxicological tests. Acute toxicity of propolis in varying polarity was determined using albino rats. Oedema volume of albino rat’s paw was used to determine the anti-inflammatory activity of the sample. Analgesic property of the sample was investigated by measuring the number of acetic acid induced writhing in albino rats. Hexane, ethyl acetate, and ethanol extracts of propolis were non-toxic at 1,000 mg/kg bodyweight. During anti-inflammatory investigation, sample at 300 and 200 mg/kg bodyweight showed better activity than the standard drug, indomethacin. Best anti-inflammatory activity was observed with ethanol fraction at 300 mg/kg bodyweight. Best analgesic activity (100%) was recorded at 200 mg/kg in hexane. These results show that both non-polar and polar extracts of propolis have anti-inflammatory and analgesic activities at various dosages.

Key words: Propolis, anti-inflammatory, analgesic, acetic acid, diclofenac, indomethacin, Nigeria, polarity.

INTRODUCTION

A natural product is a chemical compound or substance produced by living organisms such as plants, animals, microbes, algae, and marine sponges. The term natural product has also been extended for commercial purposes to refer to cosmetics, dietary supplements, and foods produced from natural sources without added artificial ingredients. Several reasons such as knowledge of composition, structure and bioactivities of natural products necessitate the study of Natural Products (Nande and Igoli, 2017). Increasing evidence suggest that majority of the bioactive phytochemical components in plants impart physiological activities and may offer a variety of health benefits such as antioxidant, antibacterial, anti-inflammatory, and anticancer (Ipav et al., 2018) or anti-venom (Tor-Anyiin et al., 2015). Propolis is a natural product derived from plant resins and collected by honeybees (workers) to be used as glue and as draught-extruder for beehives (Khalil, 2006). Recent foray into
bee propolis research has brought remarkable health benefits thereby making propolis a potential source for development of new drugs (Bankova et al., 2000). It has been reported that propolis contain at least 200 compounds with more than 100 being present in any given sample (Greenaway et al., 1999). These include fatty and phenolic acids and esters, substituted phenolic esters, bioflavonoids (flavones, flavanones, flavonols and others), terpenes, steroids, aromatic aldehydes and alcohols, and derivatives of sesquiterpenes, naphthalene and stilbenes (Marcucci et al., 1996).

Several research studies have corroborated the claim that, chemical composition and bioactivities of propolis depend on the phyto-geographical characteristics of the site of location which then varies with vegetation belts, countries, states and locality. Accordingly, these seasonal variations can influence the biosynthesis of plant metabolites and consequently affect the resin that is secreted by plants, which is used by the honeybees to produce propolis (Teixeira et al., 2008). According to Babatunde et al. (2015) and Khalil (2006), despite its sensitivity to geographical locations and the fact that constituent compositions differ according to each area, most propolis samples contain 50% resin, 30% wax, 10% essential oils, 5% pollen and 5% of other organic compounds in their chemical composition.

Due to the promise propolis holds, it has attracted researchers’ interest in the last decades because of its several biological and pharmacological properties including anti-inflammatory, analgesic, anticancer and antimicrobial activities (Christov et al., 2005). Several reports of the biological activities of propolis have been made in literature including antioxidant (Frozza et al., 2013), antimicrobial (Bispo-Junior et al., 2012), anticancer (Li et al., 2008), anti-inflammatory (Bueno-Silva et al., 2016; Cavendish et al., 2015), cytotoxicity (Alencar et al., 2007), repair of wounds (Batista et al., 2012) and antinociceptive (Cavendish et al., 2015) properties. Few reports are found on the propolis of Nigerian origin and none from Gboko.

In this paper, we report the analgesic and anti-inflammatory effect of extracts of propolis obtained from Gboko, Benue State using solvents of varying polarity; hexane, ethyl acetate and ethanol.

MATERIALS AND METHODS

Collection of propolis sample
Propolis sample was obtained from bee keepers in Gboko and the vegetation surrounding the hives was noted as Maranthes polyandra, Detarium microcarpum and Burkea africana.

Preparation of propolis extract
The homogenized propolis sample was placed in the thimble of the Soxhlet extractor and refluxed for 24 h with hexane, ethyl acetate and ethanol at a maximum temperature of 40°C. The liquid extract thus obtained was evaporated to dryness using a rotary evaporator at 40°C. The dried extracts of hexane, 12.68%, ethyl acetate, 7.46% and ethanol, 3.28% yield were prepared in a solution of olive oil for the pharmacological and toxicological tests.

Analgesic and anti-inflammatory activities of propolis sample

Experimental animals
Albino rats weighing between 70 and 120 g were obtained from the animal house of the Department of Chemical Sciences, University of Mkur, Mkur and used for the study. The experimental animals were kept in standard cages under standard conditions (25±2°C and a 12 h light/dark cycle) and maintained on standard pellets and drinking water ad libitum acclimatize for 5 days prior to commencement of the experiment. The animals were kept under hygienic conditions by constant cleaning and removal of faeces and spilled feeds from cages daily. The use and care of laboratory animals in the study were in accordance with ethical guidelines as contained in the European Convention for the Protection of Vertebrate Animals used for Experimental and Other Scientific Purposes (EEC Directive 86/609/EEC) of 1986.

Acute toxicity studies

Drug: Diclofenac sodium was used as a standard drug for the analgesic activity at a dose of 25 mg/kg body weight.

Acetic acid induced writhing test
Albino mice were induced with pain by intraperitoneal injection using 1% (v/v) acetic acid (23 mL/kg) as described by Santos et al. (1998). Animals were pre-treated with propolis sample prepared in olive oil at doses of 100, 200, 300, 500 and 1000 mg/kg body weight orally, 45 min before acetic acid administration. Control animals received 2 mL volume of olive oil and the positive control animals were treated with the reference analgesic drug; diclofenac sodium (25 mg/kg). The number of abdominal constriction (writhings) was cumulatively counted over a period of 20 min after acetic acid administration. The percentage analgesic activity was calculated using the following formula (Santos et al., 1998).

\[
\text{Analgesic activity} \% = \frac{\text{Mean writhing count (Control group-Treated group)}}{\text{Mean writhing count of control group}} \times 100
\]

Anti-Inflammatory activity assay

Egg albumin induced oedema
The test was conducted using a modified method of Akah and Nwabie (1994). The rats were divided into five groups of three rats of either sex per group and were treated as follows: Group A which served as the negative control received olive oil (2 mL/kg bodyweight), while group B which served as the positive control received 25 mg/kg body weight of indomethacin, Groups C, D and
E received 100, 200 and 300 mg/kg body weight of propolis orally, respectively. After 45 min of propolis sample administration, oedema was induced by sub-plantar injection of 0.1 mL of fresh raw egg albumin in the left hind paw. Oedema volume was determined by measuring the paw volume using a thread. The readings were taken at 20 min intervals, that is, 0, 20, 40, 60, 80, 100, and 120 min after albumin administration.

The average increase in paw size of each group was calculated and compared with the control (olive oil) and indomethacin group.

### Statistical analysis

The results were expressed as mean ± SEM. The statistical analysis was performed by means of Student’s t-test, whereas Analysis of Variance (ANOVA) followed by Dunnett’s multiple comparison test was used in order to compare more than two groups. *P* ≤ 0.05 is indicative of significance.

## RESULTS AND DISCUSSION

The use of propolis for the treatment of various ailments in which inflammation and pain are involved is cross-cultural and widespread. Its effect against diseases such as dermatological, odontological and gynaecological disorders has been widely reported (Castaldo and Capasso, 2002; De Castro, 2001). The mapping of free radicals generated by neutrophils during inflammation is the main mechanism of action for conventional anti-inflammatory drugs. It is also a well-known property of propolis (Paulino et al., 2003; El-Masry et al., 2011; Ichikawa et al., 2002). The process of inflammation includes production and release of mediators from neurons or damaged tissues which lead to various responses including pain (Paulino et al., 2003).

Hexane, ethyl acetate and ethanol extracts of propolis is non-toxic at 1000 mg/kg bodyweight (Table 1). This is in line with several reports of the relatively low toxicity of propolis. Arvouet-Grand et al. (1993) reported the oral LD50 of propolis extract to be greater than 7340 mg/kg in mice (Abozid and Ahmed, 2013). Dobrowolski et al. (1991) administered approximately 700 mg/kg orally to groups of 10 mice (five males and five females) and monitored them for 48 h. No deaths were recorded and the preparations were well tolerated by the test animals. In another study, Havsteen (1983) reports LD50 of flavonoids to range from 8000 to 40000 mg/kg bodyweight of rats. Considering the main component of propolis to be flavonoids, it should be relatively safe since flavonoids are of very low toxicity.

### Anti-inflammatory activity

Figures 1, 2, 3, 4, 5 and 6 indicate that Propolis sample at 300 mg/kg bodyweight showed a steady increase in inhibition from 20 to 100 min. This result is better than the standard drug, indomethacin. The inhibitory effect of propolis at 200 mg/kg weight administration is also better than indomethacin. The best result was observed with 200 mg/kg bodyweight administration of sample. The inhibitory effect observed on the n-hexane extract of propolis is in a dose dependent manner. The higher the dosage, the more the inhibition. Inflammation is presumably initiated by the release of histamine, kinin, fibrinolysin and phospholipase A2 (Parandin and Daroogari, 2019). These intermediaries of inflammation induce oedema by vasodilation and increased vascular permeability (Zhang et al., 2015; Li et al., 2011; Xu et al., 2014; Uwaya et al., 2020). The anti-inflammatory effect of this sample may be due to the decrease in the amount of the induced intermediaries earlier mentioned.

The inhibition of inflammation by ethyl acetate is not dose dependent. No regular pattern is observed with its administration. However, better anti-inflammatory activity is observed at 200 mg/kg bodyweight administration at 60 minutes as seen in figure 4. This too is better than the standard drug, indomethacin.

Figure 6 shows better inhibitory activity of ethanol fraction of propolis at 300 mg/kg bodyweight administration. The activity is time dependent. In the earlier phase, its activity at 300 mg/kg bodyweight administration is not as good as indomethacin. In the later phase; 60-120 min (Figures 4, 5, 6 and 7), the inhibition of inflammation is more pronounced and gets better than indomethacin.

Major constituents of propolis apart from resins and waxes include flavonoids, terpenoids and phenolic acids.

### Table 1. Result of acute toxicity test on Propolis using hexane, ethyl acetate and ethanol solvents.

<table>
<thead>
<tr>
<th>Dose (mg/kg)</th>
<th>Hexane</th>
<th>Ethyl acetate</th>
<th>Ethanol</th>
</tr>
</thead>
<tbody>
<tr>
<td>100</td>
<td>0/3</td>
<td>0/3</td>
<td>0/3</td>
</tr>
<tr>
<td>200</td>
<td>0/3</td>
<td>0/3</td>
<td>0/3</td>
</tr>
<tr>
<td>300</td>
<td>0/3</td>
<td>0/3</td>
<td>0/3</td>
</tr>
<tr>
<td>500</td>
<td>0/3</td>
<td>0/3</td>
<td>0/3</td>
</tr>
<tr>
<td>1000</td>
<td>0/3</td>
<td>0/3</td>
<td>0/3</td>
</tr>
</tbody>
</table>

(0/3); 0=number of deaths, 3=number of mice used per group for the assessment.

Source: Authors
such as cinnamic acid, p-coumaric acid, caffeic acid, ferulic acid and their derivatives (Razmi et al., 2013; Pimenta et al., 2015; Sforcin, 2016). Parandin and Daroogari (2019) reported flavones, flavonols, flavanones, flavanons, chalcones, dihydrochalcones, isoflavones, isodihydroflavones, flavans, isoflavones and...
neoflavonoids as types of flavonoids found in propolis. Several studies show that phytochemicals including flavonoids, phenolic acids and terpenoids have antimicrobial (Inui et al., 2014; Brodowska, 2017; Górniak et al., 2019), antinociceptive, anti-inflammatory and antioxidant properties (Murugan and Parimelazhagan, 2013; Wang et al., 2014; Yamanishi et al., 2014; Fazio et al., 2013; Parandin and Daroogari, 2019; Fernandez et

Figure 3. Comparative anti-inflammatory activities of hexane (HEP), ethyl acetate (EAP) and ethanol (Et) extracts of propolis at 40 min.
Source: Authors

Figure 4. Comparative anti-inflammatory activities of hexane (HEP), ethyl acetate (EAP) and ethanol (Et) extracts of propolis at 60 min.
Source: Authors
Figure 5. Comparative anti-inflammatory activities of hexane (HEP), ethyl acetate (EAP) and ethanol (Et) extracts of propolis at 80 min.
Source: Authors

al., 1998; Yonathan et al., 2006; Xu et al., 2016; Zhao et al., 2016). This is because they have been reported to prevent the production of inflammation intermediaries such as prostaglandins, arachidonic acid, histamine, bradykinins (Amresh et al., 2007; Mirazi and Hosseini, 2013), and cyclooxygenase 2 (Razmi et al., 2013) which are involved in inflammation and pain.

The solvent used during extraction determines the biological activity obtained (Devequi-Nunes et al., 2018). Similar biological activity may be more pronounced with increased polarity as concluded by Wieczynska et al. (2017), who reported a stronger antimicrobial activity of ethanolic extracts than hexane extracts of polished propolis. This improved anti-inflammatory activity of ethanolic extract of propolis obtained from Gboko may be as a result of increased availability of flavonoids and other phenolics due to the high polarity of ethanol.

Analgesic activity

Hexane is highly selective during extraction. Figure 8 shows that hexane extract at 300 and 100 mg/kg bodyweight administration gave an analgesic activity of 78.14%. This result appears better than that of the standard analgesic used (diclofenac sodium). Best analgesic activity was recorded at 200 mg/kg bodyweight dosage which was 100% (Figure 8). This is not dose dependent.

Paulino et al. (2003) reported that the main constituents of propolis in temperate zones are flavonoids, whereas in the tropics other classes such as aromatic derivatives, specific terpenoids and prenylated p-coumaric acids and acetophenones may be found. In spite of these many variations in their chemical composition, they have several biological activities in common (Bankova et al., 1995; Seidel et al., 2008).

Bonvehi et al. (1994) reported poor result when determination and prediction of biological activity of propolis was based on analysis of individual compounds. This further proves the assertion that, biological activity of propolis is based on synergistic activity of individual compounds (Boisard et al., 2015; Chen and Shen, 2008).

Less activity was observed in ethyl acetate fraction. Figure 8 shows the standard drug, diclofenac had better activity than ethyl acetate fraction at 300 and 100 mg/kg dosages. The best analgesic activity was recorded at 200 mg/kg administration as seen in figure 8. The analgesic effect of propolis obtained from Gboko against acetic acid induced writhing may be due to local peritoneal receptors
Figure 6. Comparative anti-inflammatory activities of hexane (HEP), ethyl acetate (EAP) and ethanol (Et) extracts of propolis at 100 min.
Source: Authors

Figure 7. Comparative anti-inflammatory activities of hexane (HEP), ethyl acetate (EAP) and ethanol (Et) extracts of propolis at 120 min.
Source: Authors
Figure 8. Comparative analgesic activities of hexane (HEP), ethyl acetate (EAP) and ethanol (Et) extracts of propolis.
Source: Authors

or by inhibition of prostaglandin synthesis or activity (Adzu et al., 2001; Atta and Alkofahi, 1998).

Figure 8 shows that at 300 mg/kg dosage, ethanol fraction of propolis gave a relatively poor analgesic activity compared to 200 and 100 mg/kg dosage and diclofenac sodium. The component of propolis extracted, are more polar at this stage considering the high polarity of ethanol.

According to Ristivojević et al. (2015), non polar fractions of propolis contain mostly waxes and hydrocarbons which include alkanes, alkenes, alkadienes, monoesters, diesters, aromatic esters, fatty acids and steroids which in most cases do not exhibit any significant pharmacological activity. The observed activity of the n-hexane fraction may be due to a synergistic activity of the non polar components of the propolis. This is in line with the report of Ristivojević et al. (2015) that no specific compound can be associated with the pharmacologic properties of propolis.

Boisard et al. (2015) also reported that different combinations of phenolic compounds involving a complex mechanism of action are essential for biological activity of propolis. For poplar type of propolis, biological activity can be attributed to the synergistic effect of phenolic compounds including cinnamic acid and ester derivatives, including caffeic acid and Caffeic Acid PhenethylEster (CAPE), and flavonoids including pinocembrin, pinobanksin, galangin, chrysins and naringenin (Bankova, 2005; Vardar-Ünlü et al., 2008).

This activity of ethanol fraction of propolis from Nigeria agrees with ethanol extract of propolis from Bulgaria (Paulino et al., 2003), Iran (Parandin and Daroogari, 2019), and China (Sun et al., 2018). It is also in agreement with the result of hydroalcoholic propolis of Brazilian origin (Cavendish et al., 2015), water of Iraqi origin (Abdel Mahdi Kassim Altaee, 2014) and Moroccan origin (Mountassir et al., 2014) which have shown good anti-inflammatory and antinociceptive activities in animal models as reported by Al-Hariri and Abualait (2020).

The findings on analgesic study on propolis are in agreement with studies by Sun et al. (2018) which reported that different fractions from Chinese propolis extracts enriched in polyphenolic constitutions showed central and peripheral antinociceptive effects in animal models. It is also in line with the report of Cavendish et al. (2015) which concluded that the hydroalcoholic extract of Brazilian red propolis exhibited anti-inflammatory and neurogenic pain inhibition without emotional and motor side effects in rodents.

**Conclusion**

The varying polarity of solvents used, has proved that
pharmacological activities of sample fractions of propolis is as a result of synergism rather than single fraction action. This can be seen in the good analgesic activity of hexane, a non-polar solvent and ethanol, a highly polar solvent as reported. The results show that this propolis sample is non-toxic at very high doses. It also shows that both non-polar and polar extracts of propolis obtained from Gboko, Benue State have anti-inflammatory and analgesic activities at various dosages.

CONFLICT OF INTERESTS

The authors have not declared any conflict of interests.

REFERENCES


