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Vol. 16(17), pp. 953-961, 26 April, 2017 DOI: 10.5897/AJB2017.15890 Article Number: D6FDB4463972 ISSN 1684-5315 Copyright © 2017 Author(s) retain the copyright of this article http://www.academicjournals.org/AJB

African Journal of Biotechnology

Full Length Research Paper

# Phenotypic diversity in physic nut (*Jatropha curcas* L.) *in vivo* germplasm bank for superior parent selection

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#### Received 12 January, 2017; Accepted 11 April, 2017

Jatropha curcas is an interesting alternative for biodiesel production due to the high oil content in its seeds, its ability to grow in a wide range of climate and soil conditions as well as low cost of production. However, the species is considered to be in domestication and there are no defined cultivars. Therefore, it is extremely important to understand the genetic diversity of the species for selection and characterization of promising genotypes to initiate breeding programs. The objective of this study is to evaluate the phenotypic diversity of physic nut in order to select the most divergent and superior genotypes to compose future breeding programs, using multivariate analysis. Eleven agronomic characters were evaluated in 165 J. curcas genotypes from the in vivo germplasm bank, which were: Plant height, stem diameter, number of primary branches, fruit length, width, weight and shape, seed length, width and weight plus the oil content. The data were analyzed by principal component analysis (PCA), cluster analysis by Ward and k-means methods. The character fruit shape was removed from the multivariate analysis as the only one with qualitative character. The PCA resulted in 4 main components (PC), which explained 71.62% of total variance. The characters selected in PC1 were seed weight, fruit width, fruit length and fruit weight. There were 22 promising genotypes highlighted, with potential to be exploited in breeding programs. Cluster analysis by Ward and k-means methods generated 9 groups influenced by all analyzed characters, of which five groups of genotypes had advantageous characters. Regarding fruit shape, 13 genotypes had an ellipsoid lanceolate shape and the others had an ellipsoid spherical shape. Multivariate analyses allowed genotype characterization, indicating good strategies used for the selection in genetic breeding programs.

Key words: Agronomic characters, Jatropha curcas, oleaginous, multivariate analysis.

#### INTRODUCTION

Jatropha curcas, also known as physic nut, is a shrub, perennial and monoecious species that belongs to the

Euphorbiaceae family. It is in the same family with castor bean, cassava and rubber tree. It is believed that this

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Author(s) agree that this article remains permanently open access under the terms of the <u>Creative Commons Attribution</u> <u>License 4.0 International License</u> species originated from Central America; however, it may have adequate development in many tropical and subtropical countries (Laviola and Dias, 2008). *Jatropha* genus is entomophilous (Saturnino et al., 2005), which increases the genetic variability probability within species. The height of physic nut plant ranges from 2 to 5 m; it can live up to 35 years, and produces fruit with three seeds of 20 mm long, 11 mm wide and 9 mm thick. They are oblong and black. Its young leaves are reddish, and become dark-green during maturity; they have three to five lobes, petiolate, alternate. Its flowers are small and greenish-yellow (Drummond et al., 1984).

Due to the global energy crisis from environmental and climatic impacts caused by the high fossil energy consumption, the demand for renewable fuels that produce lower pollutants is increasing. In this context, physic nut is an alternative source because of some of its advantages: the high oil content of its seed, between 22 and 42% (Sunil et al., 2008; Achten et al., 2008), its low production cost, it is an edible crop and meets environmental demands, since its oil contains sulfur in insignificant amounts. The physic nut can reach between 500 and 3000 kg ha<sup>-1</sup> depending on genotype and environment (Wani et al., 2016), producing about 2000 kg ha<sup>-1</sup> of oil in the fourth year of cultivation, depending on the spacing (Laviola and Dias, 2008). However, the cultivation depends on its domestication in order to achieve higher productivity and production uniformity (Fairless, 2007). This potential can be surpassed with breeding programs along with production system improvement.

Since there are no defined cultivars and descriptors for physic nut, its genetic diversity exploration and characterization is of great importance in the species genetic breeding programs (Achten et al., 2010). Therefore, studies concerning genetic diversity among the genotypes around the world have been done at the molecular level (Pazeto et al., 2015; Pecina-Quintero et al., 2014; Sinha et al., 2016) as well as the phenotypic level (Freitas et al., 2015; Oliveira et al., 2016; Priyanka et al., 2015) stating that there is a large genetic variability, making physic nut a prosperous species for domestication and breeding.

Multivariate techniques are important tools in predicting genetic diversity, germplasm classification, accession variability ordering and analysis of genetic relationships between characteristics and existing genetic material (lqbal et al., 2008). Among these techniques, it can be highlighted the principal component analysis (PCA) and cluster analysis (Cruz et al., 2004; Gonçalves et al., 2008). The principal component analysis is a useful tool used to identify characters containing more information for the germplasm characterization, even as to inform which characters contribute less to the total variation available (Cruz et al., 2004). Cluster analysis aims to gather, by some classification criteria, the sample units in groups, for there to homogeneity within the group and heterogeneity among them (Neto and Moita, 1998). The PCA technique was applied for physic nut by Singh et al. (2016) to distinguish parental accessions for plant improvement, Nietsche et al. (2015) to evaluate the variability in reproductive traits and by Tripathi et al. (2015) to study the genetic diversity of Indian accessions. Different methods of cluster analysis have also been widely used for the species aiming to study its genetic diversity (Noor Camellia et al., 2012; Silva Junqueira et al., 2016; Reddi et al., 2016). Due to the importance of physic nut genotypes characterization for domestication and the need to obtain superior genotypes for future use in breeding programs, the present study aimed to evaluate phenotypic diversity to select the most divergent and superior genotypes from a physic nut germplasm bank using multivariate analysis strategies.

#### MATERIALS AND METHODS

We evaluated 165 physic nut plants from 50 accessions from four Brazilian states: Paraíba, Pernambuco, Tocantins and São Paulo (Table 1). The genotypes belong to the *in vivo* Germplasm Bank of the College of Agricultural and Veterinary Sciences of the Universidade Estadual Paulista - UNESP, Jaboticabal, SP, located in the Plant Production Department experimental area. Harvesting and agronomic characters were evaluated from December 2014 to May 2015.

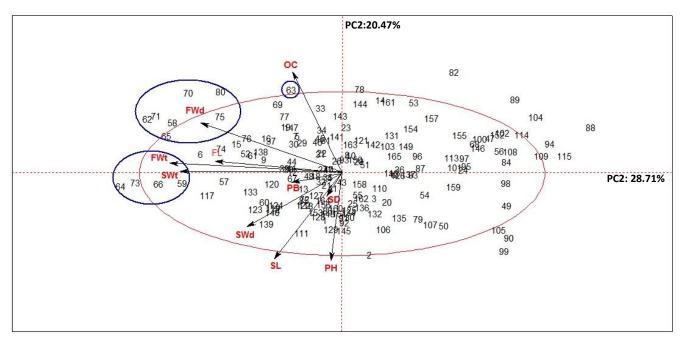
To define the characters evaluated, some descriptors used for castor bean (Ricinus communis L.) were used (MILANI, 2008), since the two species are from the same family. Eleven agronomic traits were evaluated: Plant height - measured from the plant collar to the apex (m); stem diameter - measured using a digital caliper (ZAAS brand) at 10 cm ground level interval (cm); number of primary branches - the primary branches of each genotype were observed; fruit length - length of 10 fruits per genotype were measured using a digital caliper (cm); fruit width - widths of 10 fruits per genotype were measured using a digital caliper (cm); fruit weight - 10 fruits per genotype were weighed using a digital scale (g); fruit shape - 10 fruits per plant were evaluated, classified as ellipsoid spherical or ellipsoid lanceolate, according to Laviola et al. (2011); seed length - 10 seeds per genotype were measured using a digital caliper (cm); seed width - 10 seeds per genotype were measured using a digital caliper (cm); seed weight - 10 seeds per genotype were weighed using a digital scales (g); oil content extraction was performed with a soxhlet extractor, the method includes leaching of the oil in the material via contact with a particular solvent in a series of cycles, according to the AOCS official method (AOCS, 2003) (%). The process was performed in duplicate for each genotype and subsequently the average was calculated for each plant, with results presented in g/100 g.

All fruits were harvested when their color was brown and at random from each plant over the previously mentioned period by multiple harvests. To perform the agronomic traits analyses, seeds and fruits evaluated were randomly chosen from each genotype. For fruit length, width and weight characters, as well as seed oil content, the mean values for each genotype were considered.

Principal Components Analysis and Cluster Analysis were performed using Statistica software, version 10 (STATSOFT, 2010) for all agronomic traits except fruit shape since it has a qualitative characteristic. Data were standardized resulting in mean zero and variance one for all the variables analyzed. Hierarchical clustering was performed by the Ward method and dissimilarity estimates were generated using the Euclidean distance procedure.

Origin	Coordinates	Accession identification	Number of plants		
Tocantinópolis – TO	06°19'S 47°24'W	1	3		
Garanhuns - PE	08°56'S 36°30'W	2/3/4/5/6/7/8/9/10/11/12/13/14	2/2/5/2/2/3/4/5/1/2/3/2/3		
Mundo Novo – PE	07°35'S 37°11'W	15 / 16 / 17 / 18 / 19 / 20 / 21 / 22	3/6/5/3/2/8/5/7		
Pugmil - TO	06°24'S 37°48'W	23 / 24 / 25 / 26 / 27	2/2/3/1/3		
Alagoinha - PB	06°57'S 35°32'W	28 / 29 / 30 / 31	9/6/6/6		
Oliveira de Fátima - TO	10°42'S 48°54'W	32 / 33 / 34 / 35 / 36 / 37 / 38	2/3/3/2/2/3/1		
Marizópolis - PB	09°47'S 49°39'W	39 / 40 / 41 / 42	5/1/6/3		
Ituverava - SP	20°20'S 47°46'W	43 / 44 / 45 / 46 / 47 / 48 / 49 / 50	3/2/3/2/2/2/2/2		

Table 1. Physic nut origin, identification and number of plants used in this study.



**Figure 1.** Biplot graph with dispersion of 165 physic nut genotypes according to the principal components PC1 x PC2 and vectors projection of the agronomic traits: plant height (PH), stem diameter (SD), number of primary branches (PB), seed weight (SWt), seed width (SWd), seed length (SL), fruit width (FWd), fruit length (FL), fruit weight (FWt) and oil content (OC).

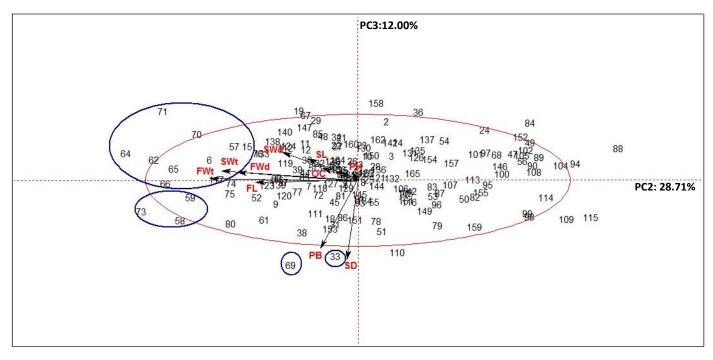
#### **RESULTS AND DISCUSSION**

The first four principal components explained 71.62% of the variance contained in the original ten variables, and the first principal component (PC1) retained 28.71% of the original variance (Figure 1). The principal characters that explained this variance retention (PC1) were the production components: seed weight, fruit weight, width and length (Table 2). The second main component (PC2) retained 20.47% of the variance, explained by different characters of the plants such as plant height, seed length and oil content (Table 2). The third principal component (PC3) retained 12.00% of the variance, which was explained by the plant proportions: stem diameter and number of primary branches (Table 2). The fourth principal component (PC4) retained 10.42% of the variance and was contributed by fruit length and seed width; loads presenting absolute value greater than 0.5 were considered relevant (Table 2).

The 165 genotypes were distributed along the axis of the principal components. That means the closer a genotype is to the other; the more similar they will be, while the genotypes that are further away from the axis of the principal components are the most discrepant. The two-dimensional plane formed by PC1 (28.71%) and PC2 (20.47%) components retained altogether 49.18% of the original variance. It can be observed in Figure 1, that 58, 59, 62, 63, 64, 65, 66, 70, 71, 73, 75 and 80 genotypes are located to the left of PC2, indicating negative correlations, differentiated by the seed weight, fruit width,

Table 2. Correlation between the variables and the principal component (PC) of J. curcas
genotypes for agronomic traits: Plant height (PH), stem diameter (SD), number of primary
branches (PB), seed weight (SWt), seed width (SWd), seed length (SL), fruit width (FWd),
fruit length (FL), fruit weight (FWt) and oil content (OC).

Parameter	PC1	PC2	PC3	PC4
PH	-0.061	-0.733	0.081	-0.229
SD	-0.079	-0.216	-0.793	-0.169
PB	-0.236	-0.084	-0.680	0.465
SWt	-0.824	-0.031	0.070	0.029
SWd	-0.489	-0.437	0.219	0.592
SL	-0.341	-0.725	0.188	0.042
FWd	-0.717	0.370	0.039	-0.013
FL	-0.638	0.078	-0.072	-0.567
FWt	-0.886	0.072	-0.013	-0.111
OC	-0.246	0.767	0.075	0.239



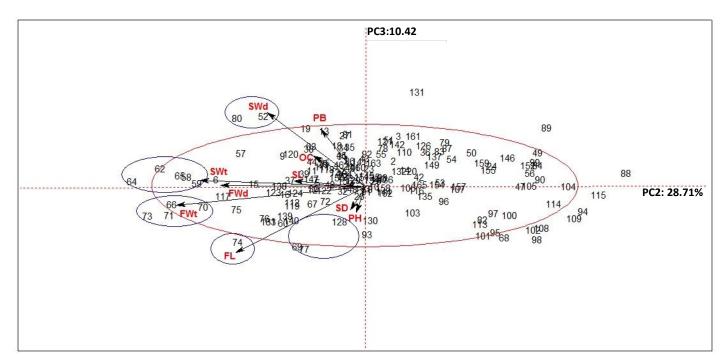
**Figure 2.** Biplot graph with dispersion of 165 physic nut genotypes according to the principal components PC1 x PC3 and vectors projection of the agronomic traits: Plant height (PH), stem diameter (SD), number of primary branches (PB), seed weight (SWt), seed width (SWd), seed length (SL), fruit width (FWd), fruit length (FL), fruit weight (FWt) and oil content (OC).

length, and weight, plant height and oil content variables. The two-dimensional plane formed by PC1 and PC3 (12.00%) components retained 40.71% of the variance and was characterized by seed weight, fruit width, length and weight, stem diameter and number of primary branches variables, differentiating genotypes 6, 15, 33, 57, 58, 62, 64, 65, 66, 69, 70, 71, 73 and 117, located to the left of PC3 (Figure 2).

The two-dimensional plane formed by PC1 and PC4 (10.42%) components retained 39.13% of the variance,

and was characterized by the seed weight, fruit width, length and weight, plus seed width variables, differentiating genotypes 13, 17, 19, 52, 58, 59, 62, 64, 65, 66, 69, 70, 71, 73, 74, 80 and 128, located to the left of CP4 (Figure 3).

The seed weight and oil content characters also had great importance in the principal components analysis of Malaysian physic nut (Shabanimofrad et al., 2013) and castor accessions (Anjani, 2010). According to the report of Reis et al. (2015) in their study on physic nut



**Figure 3.** Biplot graph with dispersion of 165 physic nut genotypes according to the principal components PC1 x PC4 and vectors projection of the agronomic traits: plant height (PH), stem diameter (SD), number of primary branches (PB), seed weight (SWt), seed width (SWd), seed length (SL), fruit width (FWth), fruit length (FL), fruit weight (FWt) and oil content (OC).

accessions, oil production per plant showed coefficient of variation of 60.00%, indicating that this is a character with high phenotypic variability. On the other hand, characteristics related to the seed presented the lowest coefficients of variation, with values below 10%. Seed morphological characters of the wild accessions are considered to be the first step in ascertaining genetic variability of the population. Large seeds, for example, may be favoured because they generate larger and more vigorous seedlings with better chances of survival than small seeds, on the other hand, the small seeds may have a selection advantage thanks to its wider and more effective dispersal (Eriksson, 1999).

The variables that have the same sign act in the same direction; that is, when there is an increase in one variable, it also occurs in the other, and those with opposite signs act in opposite directions; when the value of one increases, the other decreases. Thus, in accordance with the correlations indicated in Table 2, in PC1 and PC3, variables considered with higher discriminatory power act directly. In PC2, plant height and seed length characters act directly, but indirectly to the oil content. In turn, in PC4, seed width and fruit length characters act indirectly.

Carvalho (2010) reported that the number of primary branches was a character that had high contribution in the third principal component in physic nut, and may be the variable discarded according to Cruz et al. (2004) and Pereira et al. (2003)'s criteria, showing similar results presented in our study. Thus, seed width variable could also be discarded, since it showed a correlation only in the fourth principal component. However, considering the economic potential of the understory crop in the initial years of establishment, the characters plant height and number of branches are considered important for major selection indices when the objective is to incorporate physic nut in an agroforestry system wherein balanced trade off can be made on yield (Rao et al., 2008).

By observing the agronomic traits values for each genotype selected by PCA (Table 3), genotypes 63 and 53 were selected only due to their high seed-oil content, genotypes 75 and 15 for their high fruit width values, genotype 69 for the greatest number of primary branches and high stem diameter value, genotypes 64, 66 and 59 for their high seed and fruit weights and genotype 73 for higher seed weight and fruit length and weight. Genotypes 71, 65, 62 and 117 had higher seed weight, fruit width and weight, 57 had high seed weight, genotype 6 had higher seed weight and fruit weight, 58 had higher fruit width, length and weight values. Genotype 33 was selected due to its high stem diameter value. Genotypes 17 and 128 were selected due to the high plant height and larger stem diameter, genotype 52 had high seed width value and genotype 74 had the longest fruit length.

Laviola et al. (2011) found that the stem diameter and plant height traits contributed 12 and 11%, respectively, to the genetic diversity of physic nut accessions. Considering that physic nut is a bushy plant that can

Table 3. Physic nut genotypes averages	selected by principal	component analysis fo	r 10 agronomic traits: F	Plant height (PH), stem
diameter (SD), number of primary branche	s (PB), seed weight (	SWt), seed width (SWd),	, seed length (SL), fruit v	vidth (FWd), fruit length
(FL), fruit weight (FWt) and oil content (OC)	1.			

Accession	Genotype	PH (m)	SD (cm)	PB	SWt (g)	SWd (mm)	SL (mm)	FWdh (mm)	FL (mm)	FWt (g)	OC (%)
21	75	3.00	21.00	4.00	0.79	10.64	17.83	23.06	30.93	3.62	60.89
20	69	2.50	28.00	5.00	0.75	10.51	16.98	21.67	34.54	3.48	57.37
19	64	3.30	18.00	4.00	0.82	11.23	19.24	23.83	30.72	3.65	58.60
21	73	3.00	28.00	4.00	0.82	11.07	18.75	23.10	32.07	3.84	56.00
19	66	3.00	27.00	3.00	0.86	11.17	18.86	23.17	30.97	3.45	56.80
19	59	3.00	26.00	4.00	0.86	11.08	18.74	22.81	29.68	3.53	56.87
20	71	2.50	12.00	2.00	0.83	10.99	18.75	23.06	33.08	3.61	60.56
20	70	2.60	16.00	2.00	0.80	10.96	17.92	23.13	31.55	3.74	62.09
19	65	3.00	18.00	4.00	0.84	11.14	17.90	23.70	30.17	3.57	58.70
19	62	2.70	16.00	4.00	0.85	11.16	18.04	23.97	30.54	3.54	59.17
6	15	2.55	16.00	3.00	0.69	11.07	18.73	23.34	29.79	3.84	56.62
18	57	3.00	15.00	4.00	0.78	11.42	18.77	22.23	29.75	3.55	58.20
3	6	3.00	15.00	4.00	0.78	11.12	18.04	23.10	29.78	3.93	56.24
22	80	2.15	26.00	5.00	0.83	11.25	17.59	22.71	28.19	3.53	62.28
18	58	2.50	30.00	4.00	0.80	11.16	18.38	22.76	31.25	3.65	62.61
11	33	2.60	31.00	5.00	0.68	10.66	17.83	22.77	29.10	3.26	61.94
19	63	2.15	12.00	5.00	0.76	11.00	17.36	22.87	29.95	3.22	57.50
32	117	3.20	21.00	4.00	0.75	11.06	18.77	23.49	29.92	3.65	54.01
7	17	3.20	22.00	3.00	0.71	11.19	17.82	22.40	29.08	3.24	57.17
21	74	2.95	18.00	4.00	0.78	11.48	18.58	22.66	33.13	3.52	57.38
16	52	2.65	18.00	6.00	0.76	11.23	18.62	22.94	28.28	3.30	58.82
16	53	2.60	19.00	4.00	0.68	10.45	17.49	22.49	28.41	2.90	59.00
36	128	3.20	24.00	2.00	0.75	11.10	18.76	22.20	30.26	3.02	52.98

reach up to 5 m in height (Saturnino et al., 2005) and its harvest is mainly performed manually, selecting smaller genotypes will improve the harvesting process. In addition, for commercial purposes, those genotypes that have taller trees, low oil content, and low productivity are not feasible.

Genotypes 70 and 80 were considered the most promising and with potential to be used in genetic plant breeding programs, as they presented higher seed weight, fruit weight and width, oil content and also lower plant height. In a study of physic nut seeds using genotypes from Suriname, Ethiopia, Nigeria, Brazil and China, Vaknim et al. (2011) found that the oil content varied between 39 and 62%, Aguilera-Cauich et al. (2015) verified an average of 50.52% oil content in American physic nut accessions, whereas in the present work values observed vary between 50.81 and 62.89%.

Cluster analysis was performed using the Ward method, which generated the dendrogram shown in Figure 4. The dendrogram allowed the formation of nine groups from a cutoff level where abrupt changes were observed, as recommended by Cruz et al. (2004). Cluster analysis by the non-hierarchical k-means method (Figure 5) allowed the characterization of the nine groups formed, according

to the generated dendrogram. It is possible to observe that five within nine groups had oil content above average, and these same groups presented below average for plant height; that is, they were considered groups with genotypes that presented good results to be considered in genetic breeding programs. Group 1 was considered the best group, especially for presenting the highest values for seed weight, fruit width, length and weight and oil content, plus the low plant height. Group 2 presented the largest stem diameter and higher number of primary branches, besides high oil content and low plant height. Groups 3, 5 and 6 were classified as having the worst performance, as they had low oil content and high plant height values, therefore not considered relevant. Group 4 is composed of genotypes with high oil content, but low number of primary branches. Groups 5 and 6 were characterized as having high plant height values and low oil content. Group 7, despite presenting low values for all traits related to seed and fruit, had high oil content. Group 8 was also considered good due to the low plant height and high oil content. In contrast, Group 9 was not considered interesting since it was characterized by having low values for traits related to fruit and seed, and low oil content.

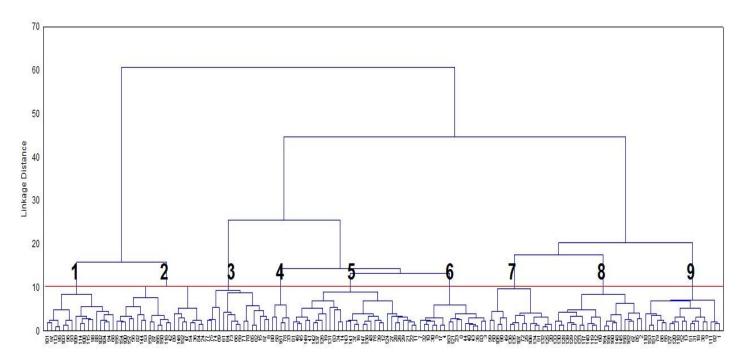
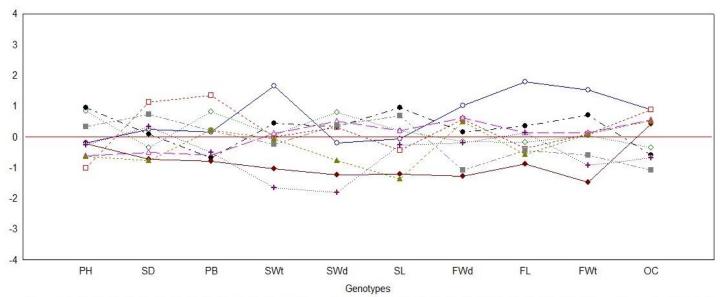


Figure 4. Hierarchical cluster analysis dendrogram using the Euclidean distance and the link between the groups by Ward method for agronomic traits: Plant height, stem diameter, number of primary branches, seed weight, seed width, seed length, fruit width, fruit length, fruit weight and oil content.



--- Group 1: 6; 58; 59; 61; 62; 64; 65; 66; 69; 70; 71; 73; 74; 75; 76; 77 --- Group 2: 9; 18; 31; 33; 38; 45; 51; 52; 78; 80; 81; 110; 149; 161 ---- Group 3: 1; 2; 3; 4; 13; 17; 35; 40; 43; 44; 54; 55; 57; 83; 85; 86; 87; 92; 111; 118; 120; 125; 127 ---- Group 4: 5; 10; 11; 15; 16; 19; 21; 22; 26; 27; 28; 29; 30; 34; 36; 37; 39; 46; 48; 137; 138; 141; 142; 143; 147; 150 - Group 5: 12; 32; 60; 67; 72; 112; 117; 119; 122; 123; 124; 128; 130; 133; 139; 140; 148; 156; 158; 160; 162; 164 --- Group 6: 20; 25; 41; 42; 49; 50; 79; 90; 91; 99; 105; 106; 107; 116; 126; 129; 132; 134; 135; 136; 145; 151; 153; 159 - Group 7: 24; 47; 56; 68; 84; 88; 89; 94; 100; 102; 104; 109; 146; 152; 155 --- Group 8: 7; 8; 14; 23; 53; 63; 82; 121; 131; 144; 154; 157; 163; 165 -+- Group 9: 93; 95; 96; 97; 98; 101; 103; 108; 113; 114; 115

Figure 5. Profile graph of group centroid distributions in the cluster analysis by k-means, formed by the variables: Plant height (PH), stem diameter (SD), number of primary branches (PB), seed weight (SWt), seed width (SWd), seed length (SL), fruit width (FWd), fruit length (FL), fruit weight (FWt) and oil content (OC).

According to groups formed by Ward's dendrogram, hybridizations between groups 1 x 4, 1 x 7, 1 x 8, 2 x 4, 2

x 7, 2  $\times$  8, 4 x 7 and 4 x 8 can be recommended, due to the distance between the groups, thus having

heterogeneity between them. Also they have genotypes with attractive characteristics for the physic nut production system, such as a low plant height and high oil content. It is noteworthy that, within groups, the genotypes belonging to different accessions had similar values in some characters. Rao et al. (2008) and Spinelli et al. (2010) observed positive correlation between the number of branches and plant height, and productivity character as well. This is a very important information that can be used to facilitate the selection of promising genotypes for this crop which still has a lot of genetic variability to be exploited.

Cluster analysis indicated that there is variability within the accessions, as the genotypes belonging to the same accession are in different groups and their characteristics do not always resemble each other. In a study with phenotypic diversity of physic nut, Aguilera-Cauich et al. (2015) found variability within the accessions and concluded in their study that are greater diversity among the American physic nut accessions evaluated in comparison with reports on diversity for India and Malaysia. Likewise, Trebbi et al. (2015) verified an increased genetic variability and heterozygosity in physic nut accessions of Mexico and Guatemala. These results can be explained by the fact that Central America is the center of origin of the species. Genetic variability in physic nut population was also found by Brasileiro et al. (2013), as well as higher estimates of heritability, in which it was possible to obtain genetic gains for growth and production traits. Reinforcing this information, Abreu et al. (2009), in physic nut accessions, obtained high heritability coefficients for plant height, first leaf height, stem diameter and number of leaves, due to the wide genetic variability among accessions. The higher the heritability of a characteristic, the better is the prediction of genetic value by individual performance and the faster the response to selection for this trait (Oliveira et al., 2007).

In a study with multivariate analysis for resistant peanut genotypes selection, Pitta et al. (2010) concluded that Ward and K-means clustering methods were efficient and complementary to the principal components analysis, also presented in this study. Due to the presented variability within the accessions, the generated dendrogram did not reveal a pattern with similar geographic regions, and it is explained by the fact that each group brings together different accessions within it. Similar results with physic nut were obtained by Tripathi et al. (2015) that used the k-means method to group accessions from different parts of India, Jun-ling et al. (2010) that studied by UPGMA method 38 accessions from different regions of China and Indonesia, and Kaushik et al. (2007) that analyzed accessions from India by non-hierarchical Euclidian cluster analysis, concluding that geographical diversity need not obligatorily be related to genetic diversity.

For fruit shape, genotypes 29, 65, 66, 67, 68, 69, 70,

71, 72, 73, 74, 75 and 77 presented an ellipsoid lanceolate, and most of them were in Group 3. All the other 152 genotypes had an ellipsoid spherical fruit shape. These results corroborate with that of Laviola et al. (2011)'s study where among 195 physic nut accessions evaluated, 190 had fruit in an ellipsoid spherical shape, four had ellipsoid lanceolate shape and one had ellipsoid ovoid fruit shape. It can be concluded, then, that fruit shape is a qualitative trait that contributes little to the variance among accessions.

#### Conclusions

The study showed that there is genetic variability for the physic nut accessions evaluated for the traits assessed and the results are very important information to be exploited in a genetic breeding program. Multivariate analyses allowed genotype characterization and also indicated those that are different from each other, allowing the targeting of crossings. All agronomic traits allowed genotype discrimination and characterization.

#### **CONFLICT OF INTERESTS**

The authors have not declared any conflict of interests.

#### REFERENCES

- Abreu FB, Resende MDV, Anselmo JL, Saturnino HM, Brenha JAM, Freitas, FB (2009). Variabilidade genética entre acessos de pinhãomanso na fase juvenil. Magistra 21(1):36-40.
- Achten WMJ, Nielsen LR, Aerts R, Lengkeek AG, Kjaer ED, Trabucco A, Hansen JK, Maes WH, Graudal L, Akinnifesi FK, Muys B (2010). Towards domestication of *Jatropha curcas*. Biofuel 1(1):91-107.
- Achten WMJ, Verchot L, Franken YJ, Mathijs E, Sing VP, Aerts R, Muys B (2008). Jatropha bio-diesel production and use. Biomass Bioenergy 32(12):1063-1084.
- Aguilera-Cauich EA, Pérez-Brito D, Yabur AN, López-Puc G, Najera GC, Rivero JCS, Atoche CR, Uc-Várguez A, Góngora-Canul C, Mijangos-Cortes JO (2015). Assessment of phenotypic diversity and agronomic contrast in American accessions of *Jatropha curcas* L. Ind. Crops Prod. 77:1001-1003.
- Anjani K (2010). Pattern of genetic diversity among *Fusarium* wilt resistant castor germplasm accessions (*Ricinus communis* L.). Elect. J. Plant Breed. 1(2):182-187.
- AOCS (2003). Official methods and recommended practices of the American Oil Chemistry Society. 5th ed. Champaign.
- Brasileiro BP, Silva SA, Souza DR, Santos PA, Oliveira RS, Lyra DH (2013). Genetic diversity and selection gain in the physic nut (Jatropha curcas). Genet. Mol. Res.12(3):2341-2350.
- Carvalho DS (2010). Comportamento genético de progênies de meioirmãos de pinhão manso no recôncavo baiano, Brasil. Thesis (Master's degree) - Universidade Federal do Recôncavo da Bahia, Cruz das Almas, 51 p.
- Cruz CD, Regazzi AJ, Carneiro PCS (2004). Modelos biométricos aplicados ao melhoramento genético. In. Cruz CD, Regazzi, JA, Carneiro PCS (Eds.), Viçosa. pp. 377-413.
- Drummond OA, Purcino AAC, Cunha LHS, Veloso JM (1984). Cultura do pinhão-manso. Belo Horizonte: EPAMIG. 131:99.
- Eriksson O (1999). Seed size variation and its effect on germination and seedling performance in the clonal herb *Canvallaria majalis*. Acta

Oecol. 20:61-66.

- Fairless D (2007). Biofuel: the little shrub that could-maybe. Nature 449:652-655.
- Freitas RG, DIAS LAS, Cardoso PMR, Evaristo AB, Silva MF, Araújo NM (2015). Diversity genetic parameter estimates for yield and its components in *Jatropha curcas* L. Genet. Mol. Res. 1:1-10.
- Gonçalves LSA, Rodrigues R, Amaral ATJ, Karasawa M, Sudré CP (2008). Comparison of multivariate statistical algorithms to cluster tomato heirloom accessions. Genet. Mol. Res. 7(4):1289-1297.
- Iqbal Z, Arshad M, Mahmood T, Waheed A (2008). Evaluation of soybean [*Glycine max* (L.) Merrill] germplasm for some important morphological traits using multivariate analysis. Pak. J. Bot. 40(6):2323-2328.
- Jun-Ling S, Xiang-Nan J, Hui-Qun N, Pei-Guang S, Shi-Hui N, Xiao-Yang C (2010). AFLP analysis of genetic diversity of Jatropha curcas grown in Hainan, China. Trees 24(3):455-462.
- Kaushik N, Kumar K, Kumar S, Kaushik N, Roy S (2007). Genetic variability and divergence studies in seed traits and oil content of Jatropha (*Jatropha curcas* L.) accessions. Biomass Bioenergy 31:497-502.
- Laviola BG, Dias LAS (2008). Teor e acúmulo de nutrientes em folhas e frutos de pinhão-manso. Rev. Bras. Ciênc. Solo 32:1969-1975.
- Laviola BG, Bhering LL, Mendonça S, Rosado TB, Albrecht JC (2011). Caracterização morfo-agronômica do banco de germoplasma de pinhão-manso na fase jovem. Biosci. J. 27(3):371-379.
- Lima RLS, Severino LS, Sampaio LR, Freire MAO, Beltrão, NEM, Arriel NHC (2009). Crescimento e teor foliar de nutrientes em mudas de pinhão-manso (*Jatropha curcas* L.) em substratos contendo cinco materiais orgânicos e fertilizante mineral. Rev. Bras. Ol. Fibros. 13(1):29-36.
- MILANI M (2008). Descritores de mamona utilizados pela Embrapa Algodão. Embrapa Algodão. Documentos.
- Neto JMM, Moita GC (1998). Uma introdução à análise exploratória de dados multivariados. Quím. Nova 21(4):467-469.
- Nietsche S, Vendrame WA, Crane JH, Pereira MC, Costa A, Reis ST, (2015). Variability in reproductive traits in *Jatropha curcas* L. accessions during early developmental stages under warm subtropical conditions. Glob. Change Biol. Bioenergy 7:122-134.
- Noor Camellia NA, Thohirah Lee A, Abdullah NAP (2012). Genetic relationships and diversity of *Jatropha curcas* accessions in Malaysia. Afr. J. Biotechnol. 11(13):3048-3054.
- Oliveira JPM, Silva JF, Oliveira BS, Santos PGF, Silveira PS, Matos FS (2016). Phenotypic characterization of physic nut populations. Afr. J. Agric. Res. 11(45):4559-4566.
- Oliveira MM, Rota EL, Dionello NJL, Aita MF (2007). Herdabilidade e correlações genéticas do perímetro escrotal com características produtivas em bovinos de corte: Revisão. R. Bras. Agrociência 13:141-146.
- Pazeto MSR, Unêda-Trevisoli SH, Corrêa AAP, Vianna VF, Leite DC, Di Mauro AO (2015). Genetic diversity in *Jatropha* species from different regions of Brazil based on morphological characters and interssimple sequence repeat (ISSR) molecular markers. Afr. J. Biotechnol. 14(25):2066-2079.
- Pecina-Quintero V, Anaya-López JL, Zamarripa-Colmenero A, Núñez-Colín CA, Montes-García N, Solís-Bonilla JL, Jiménez-Becerril MF (2014). Genetic structure of *Jatropha curcas* L. in Mexico and probable centre of origin. Biomass Bioenergy 60:147-155.
- Pereira FHF, Puiatti M, Miranda GV, Silva DJH, Finger FL (2003). Divergência genética entre acessos de taro utilizando caracteres morfoqualitativos de inflorescência. Hortic. Bras. 21(3):520-524.
- Pitta MR, Boiça Jr AL, Jesus FG, Tagliari SRA (2010). Seleção de genótipos resistentes de amendoinzeiro à Anticarsia gemmatalis hübner (Lepidoptera: Noctuidae) com base em análises multivariadas. Neotrop. Entomol. 39(2):260-265.

- Priyanka, Ram D, Shiva N, Anurag K, Dalbeer (2015). Assessment of variability parameters and character association for quantitative traits in Jatropha. Trends Biosci. 8(15):3896-3899.
- Rao GR, Korwar GR, Shanker AK, Ramakrishma YS (2008). Genetic associations, variability and diversity in seed characters growth, reproductive phenology and yield in *Jatropha curcas* L. accessions. Trees 22(5):697-709.
- Reddi KR, Satya AK, Ramesh P, Lekkala SP, Narendra K, Reddy PCO, Reddy CVCM, Sekhar AC (2016). Molecular genetic assessment of *Jatropha curcas* L. germplasm of diverse origin along with its wild relatives for various early growth and establishment related traits. Int. J. Pharma Bio Sci. 7(4):(B)261-271.
- Reis MVM, Junior PCD, Campos TO, Diegues IP, Freitas, SC (2015). Variabilidade genética e associação entre caracteres em germoplasma de pinhão-manso (*Jatropha curcas* L.). Rev. Ciênc. Agron. 46(2):412-420.
- Saturnino HM, Pacheco DD, Kakina J, Tominaga N, Gonçalves NP (2005). Cultivation of *Jatropha curcas* L. Rev. Inf. Agropecuário 26(229):44-78.
- Shabanimofrad M, Rafii MY, Megat Wahab PE, Biabani AR, Latif MA (2013). Phenotypic, genotypic and genetic divergence found in 48 newly collected Malaysian accessions of *Jatropha curcas* L. Ind. Crops Prod. 42:543-551.
- Silva Junqueira SV, Peixoto ALD, Laviola GB, Bhering LL, Mendonça S (2016). Bayesian multi-trait analysis reveals a useful tool to increase oil concentration and to decrease toxicity in *Jatropha curcas* L. Plos One 11(6):1-10.
- Singh S, Prakash A, Chakraborty NR, Wheeler C, Wheeler PK, Ghosh A (2016). Trait selection by path and principal component analysis in *Jatropha curcas* for enhanced oil yield. Ind. Crops Prod. 86:173-179.
- Sinha P, Islam MDA, Negi MS, Tripathi SB (2016). First identification of core accessions of *Jatropha curcas* from India based on molecular genetic diversity. Plant Genet. Resour. 14(1):77-80.
- Spinelli VM, Rocha RB, Ramalho AR, Marcolan AL, Vieira JR, Fernandes CF, Militão JSLT, Dias LAS (2010). Primary and secondary yield components of the oil in physic nut (*Jatropha curcas* L.). Ciênc. Rural 40(8):1752-1758.
- Statsoft Inc. (2010). STATISTICA, versão 10. http://statistica.io/.
- Sunil N, Varaprasad KS, Sivaraj N, Kumar TS, Abrahan B, Prasad RBN (2008). Assessing *Jatropha curcas* L. germplasm *in situ*: a case study. Biomass Bioenergy 32(3):198-202.
- Trebbi D, Papazoglou EG, Saadaoui ES, Vischi M, Baldini VM, Stevanato P, Cettul E, Sanzone AP, Gualdi L, Fabbri A (2015). Assessment of genetic diversity in different accessions of *Jatropha curcas*. Ind. Crops Prod. 75:35-39.
- Tripathi A, Mishra DK, Shukla JK (2015). Genetic diversity and trait association between growth, yield and seed component of *Jatropha curcas* (L.) source collection from Indian sub-continent. J. Plant Breed. Crop Sci. 7(5):143-157.
- Vaknim Y, Ghanim M, Samra S, Dvash L, Hendelsman E, Eisikowitch D, Samocha Y (2011). Predicting *Jatropha curcas* seed-oil content, oil composition and protein content using near-infrared spectroscopy A quick and non-destructive method. Ind. Crops Prod. 34(1):1029-1034.
- Wani S, Garg KK, Chander G (2016). Water needs and productivity of *Jatropha curcas* in India: myths and facts. Biofuels Bioprod. Bioref. 10:240-254.