

Full Length Research Paper

***Streptomyces noboritoensis* isolated from rhizosphere soil and its use in controlling banana-tissue culture contaminants**

Mohamed, SH^{1,3}, El-Helafiy, SS², Ismail Mona, A² and Sadik, AS^{1,4}

¹Department of Biology, Faculty of Science, Taif University, P.O. Box 888, Taif, KSA.

²Department of Biotechnology, Faculty of Science, Taif University, P.O. Box 888, Taif, KSA.

³Soil, Water and Environmental Research Institute, Agricultural Research Center, 9 Gamaa st., P.O. Box 12619, Giza, Egypt.

⁴Department of Agricultural Microbiology, Faculty of Agriculture, Ain Shams University, P.O. Box 68 Hadayek Shubra, 11241, Cairo, Egypt.

Accepted 13 May, 2013

In this study, the role of *Streptomyces noboritoensis* (14) isolated from the rhizosphere of banana plant and having antagonistic activity against the bacterial- and fungal-tissue culture contaminants was determined *in vitro*. Results show that the filtrate was more effective against the fungal-tissue culture contaminants than the bacterial-tissue culture contaminants. This was indicated when jars were fungi-free and bacteria-free after one month and 21 days from incubation, respectively. Results of *in vitro* application show that the filtrate of *S. noboritoensis* (14) as a bio-control agent could be used for controlling the contaminants in banana. Data showed that treatment F (sterilized shoots treated with *Streptomyces* filtrate and cultivated on *Streptomyces*-inoculated medium) was the most effective followed by treatment E (sterilized shoots untreated with *Streptomyces* filtrate and cultivated on *Streptomyces*-inoculated medium). Therefore, the study suggests conducting further studies towards the use of streptomycetes in the biological control in a large scale production.

Key words: *Streptomyces*, tissue culture contaminants, antagonistic activities, bio-control.

INTRODUCTION

The nutrient media in which the plant tissue was cultivated is a good source of nutrient for microbial growth. These microbes compete adversely with plant tissue culture for nutrient. The presence of these microbes in these plant cultures usually resulted in increased culture mortality, the presence of latent infections can also resulted in variable growth, tissue necrosis, reduced shoot proliferation and reduced rooting (Kane, 2003).

Studies on biological control of plant diseases, particularly using antibiotic metabolites of microbial origin,

have been expanded and such materials may supplement to be an alternative to chemical disease control (Fischer et al., 1992). *Streptomyces* sp. is gram-positive filamentous bacteria that produce and secrete a wide array of biologically active compounds including antibiotics, hydrolytic enzymes and enzyme inhibitors. They are resistant to desiccation and nutrient stress, by their ability to produce spores (Dhanasekaran et al., 2005). These characteristics make streptomycetes attractive candidates for biological control agents against soil-borne plant pathogens (Samac and Kinkel, 2001; Hassan et al.,

2011). Attempts have been made to develop *Streptomyces* species as fungal root disease control agents, since *Streptomyces* spp. are capable of producing a remarkably wide spectrum of antibiotics as secondary metabolites (Yuan and Crawford et al., 1995).

El-Tarabily et al. (2009) evaluated the potential of *Actinoplanes campanulatus*, *Micromonospora chalcone* and *Streptomyces spiralis* endophytic in cucumber roots, to promote plant growth and to protect seedlings and mature plants of cucumber from diseases caused by *Pythium aphanidermatum*, under greenhouse conditions. Sadeghi et al. (2009) studied the effectiveness of two disease-suppressive *Streptomyces* spp. to control sugar beet *Rhizoctonia solani* damping off under field conditions. *Streptomyces* seed treatments reduced seedling damping off in naturally and artificially infested soils. Evaluation of final harvest revealed that the root yield of the biocontrol agents increased compared to untreated control in these years. Soil-actinomycetes particularly *Streptomyces* spp. have antagonistic activity against a wide range of plant pathogens. In recent decades, they have attracted high interests as biological control agents (Zarandi et al., 2009). In this study, the role of *Streptomyces noboritoensis* (14) isolated from the rhizosphere of banana plant and having antagonistic activity against the bacterial- and fungal-tissue culture contaminants was determined *in vitro*.

MATERIALS AND METHODS

Source of *Streptomyces* strain

In this study, the *S. noboritoensis* 14 strain isolated from banana-rhizosphere and having antagonistic activities against five of each of fungal strains and bacterial isolates recorded as tissue-culture contaminants was used. The strain was obtained from Department of Biology, Female Branch, Faculty of Sciences, Taif University, KSA.

***In vitro* evaluation of the effective substances of the *S. noboritoensis* 14 to control the growth of microbial contaminants**

Banana shoots were previously surface sterilized by immersion in a 2.5% (v:v) sodium hypochlorite solution containing 0.1% (v:v) and rinsed four times with d.H₂O (Dornelas and Vieira, 1994). Sterilized plant materials were then placed in a clean box constructed from a clear 60 x 30 x 30 cm plastic box. All shoots were subcultured on MS medium (Murashige and Skoog, 1962) supplemented with 3% sucrose, 5 mg L⁻¹ 6-benzyl aminopurine (BAP) and 0.8% Difco-Bacto agar (Becerra et al., 2004). The jars were formulated to contain no antimicrobial agent, or contained the filtrate of *S. noboritoensis* 14 prepared as described by Mohamed et al. (2012) (1 ml for each box). The jars were incubated in a clean box at cabinet temperature (25°C) under constant illumination. Type and time of appearance of contaminants in tissue culture banana jars treated with the filtrate of *S. noboritoensis* 14 as a bio-control agent for controlling the contaminants in banana tissue cultures were

measured up to 60 days using banana tissue culture shoots. Level and time of appearance of contaminants in tissue culture banana jars *via* six treatments (A, B, C, D, E and F) with the filtrate of *S. noboritoensis* 14, were also measured up to 19 weeks post incubation.

RESULTS AND DISCUSSION

Today, there are strict regulations on chemical pesticide use, and there is political pressure to remove the most hazardous chemicals from the market. Additionally, the spread of plant diseases in natural ecosystems may preclude successful application of chemicals, because of the scale to which such applications might have to be applied. Consequently, some pest management researchers have focused their efforts on developing alternative inputs to synthetic chemicals for controlling pests and diseases. Among these alternatives are those referred to as biological controls (Pal and McSpadden Gardener, 2006).

Raudales and McSpadden Gardener (2008) reviewed examples of bactericides and fungicides for controlling some plant pathogens. *Streptomyces* strains are gram-positive, filamentous soil bacteria that are well known for their abilities to produce antibiotics and other secondary metabolites. These organisms have been implicated in the antagonism of a wide variety of plant pathogenic fungi, bacteria, and nematodes and are currently under investigation for their potential use as biological disease control agents (Ara et al., 2012). *Magnaporthe oryzae*, the causal agent of rice blast disease was *in vitro* suppressed with 100 *Streptomyces* isolates (Zarandi et al., 2009). They showed that soil-actinomycetes particularly *Streptomyces* spp. have attracted high interests as biological control agents. Results in Table 1 show the type and time of appearance of contaminants in control-tissue culture banana jars (untreated with *Streptomyces* filtrate). No bacterial contaminants were found 4 days post culturing while both of bacterial and fungal contaminants were increased by time of incubation, as the maximum growth of contaminants appeared after two months of incubation. Results are shown in Figure 1.

Type and time of appearance of contaminants in tissue culture banana jars treated with the filtrate of *S. noboritoensis* 14 as a bio-control agent for controlling the contaminants in banana tissue cultures are recorded in Table 2. Data appeared that the filtrate of the applied *Streptomyces* strain was effective in inhibiting the growth of fungal contaminants, as the jars were fungi-free and bacteria-free after one month and 21 days from incubation, respectively. In other words, the growth of fungal and bacterial contaminants was weak in the Jars of banana-tissue culture treated with *Streptomyces* filtrate till 60 days from incubation. As a conclusion, the filtrate was more effective against the fungal-tissue culture

Table 1. Type and time of appearance of contaminants in control-tissue culture banana jars (untreated with *Streptomyces* filtrate).

Type of contaminant	Jars of banana tissue culture untreated with <i>Streptomyces</i> filtrate						
	Days post incubation						
	4	6	8	10	15	30	60
Bacteria	-	+	++	++	+++	+++	++++
Fungi	+	++	++	++	+++	+++	++++

-, No growth; +, weak growth; ++, moderate growth; +++, good growth; +++++, abundant growth.



Figure 1. Type of contaminants in control-tissue culture banana jars (untreated with *Streptomyces* filtrate).

Table 2. Type and time of appearance of contaminants in tissue culture banana jars treated with the filtrate of *S. noboritoensis* 14 as a bio-control agent for controlling the contaminants in banana tissue cultures.

Type of contaminant	Jars of banana tissue culture untreated with <i>Streptomyces</i> filtrate						
	Days post incubation						
	4	6	8	10	15	30	60
Bacteria	-	-	-	±	+	+	+
Fungi	-	-	-	-	-	+	+

-, No growth; ±, in doubt; +, weak growth.

Table 3. Level and time of appearance of contaminants in tissue culture banana jars treated with the filtrate of *S. noboritoensis* 14 as a bio-control agent for controlling the contaminants in banana tissue cultures.

		Week post inoculation																		
		1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19
Treatment A: Non-sterilized banana shoots cultivated on sterilized medium without <i>Streptomyces</i> filtrate		-	+	++	+++	+++	+++	+++	+++	+++	+++	+++	+++	+++	+++	+++	+++	+++	+++	+++
Treatment B: Non-sterilized banana shoots cultivated on sterilized with <i>Streptomyces</i> filtrate		-	-	-	-	-	+	+	+	++	+++	+++	+++	+++	+++	+++	+++	+++	+++	+++
Treatment C: Sterilized banana shoots cultivated on sterilized medium without <i>Streptomyces</i> filtrate		-	-	-	-	-	-	-	-	+	+	+	+	++	++	++	++	++	++	++
Treatment D: Sterilized banana shoots cultivated on sterilized medium with <i>Streptomyces</i> filtrate		-	-	-	-	-	-	-	-	-	-	-	-	+	+	+	+	++	++	++
Treatment E: Banana shoots sterilized and dipped in <i>Streptomyces</i> filtrate then cultivated on sterilized medium without <i>Streptomyces</i> filtrate		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	+	+
Treatment F: Banana shoots sterilized and dipped in <i>Streptomyces</i> filtrate then cultivated on sterilized medium with <i>Streptomyces</i> filtrate		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

-, No contamination; +, weak; ++, moderate; +++, high.



Figure 2. Level and time of appearance of contaminants in different tissue culture banana jar treatments (A, B and C) 15, 16, 17, 18 and 19 weeks post incubation (From top to bottom).

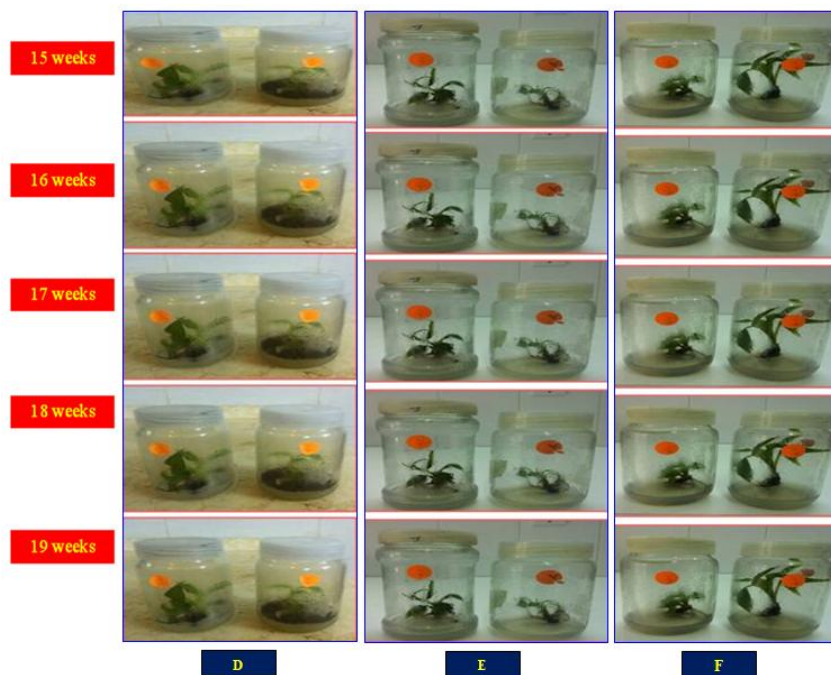


Figure 3. Level and time of appearance of contaminants in different tissue culture banana jar treatments (D, E and F) 15, 16, 17, 18 and 19 weeks post incubation (From top to bottom).

contaminants than the bacterial-tissue culture contaminants. These results are in harmony with that of Sabaratnam and Traquair (2002), who reported that formulations of a *Streptomyces* biological control agent for *Rhizoctonia* damping-off in tomato seedlings were developed for the first time from vegetative propagules obtained from actively growing, nonsporulating liquid cultures. Also, Strap and Crawford (2006) showed that rhizosphere-streptomycetes in soils could act as producers of bioactive metabolites and could be used commercially as inoculants bio-control agents primarily to control fungal root diseases. In Hungary, Dormanns-Simon (2007) reported that microbiological agents such as Mycostop based on *Streptomyces griseoviridis* K61 was used for the control of damping-off and *Fusarium* wilt. The MycostopReg was a commercial formulation of strain K61 of *S. griseoviridis*. Smith et al. (2012) showed that the Mycostop Reg could be used as a fungicidal biological control agent against blueberry blossom blight.

Results of *in vitro* application (Table 3, Figures 2 and 3) show that the filtrate of *S. noboritoensis* 14 as a bio-control agent could be used for controlling the contaminants in banana. Data showed that the treatment F (sterilized shoots treated with *Streptomyces* filtrate and cultivated on *Streptomyces*-inoculated medium) was the most effective one followed by treatment E (sterilized shoots untreated with *Streptomyces* filtrate and cultivated

on *Streptomyces*-inoculated medium). This was clearly concluded by the absence of the microbial contamination in jars of treatment F up to 19 weeks post incubation compared to the control jars.

REFERENCES

- Ara I, Rizwana H, Al-Othman MR, Bakir MA (2012). Studies of actinomycetes for biological control of *Colletotrichum musae* pathogen during post harvest anthracnose of banana. *Afr. J. Microbiol. Res.* 6(17):3879-3886.
- Becerra DC, Forero AP, Gongaro GA (2004). Age and Physiological Condition of Donor Plants Affecting *In Vitro* Morphogenesis in Leaf Explants of *Passiflora edulis* f. *flavicarpa*. *Plant Cell, Tiss. Organ Cult.* 79(1):87-90.
- Dhanasekaran D, Sivamani P, Panneerselvam A, Thajuddin N, Rakakumar G, Selvamani S (2005). Biological control of tomato seedling damping off with *Streptomyces* sp. *Plant Pathol. J.* 4(2):91-95.
- Dormanns-Simon E (2007). Biological agents for the control of soil-borne pests. Technical Workshop on non-chemical alternatives to replace methyl bromide as a soil fumigant, Budapest, Hungary.
- Dornelas MC, Vieira MLC (1994). Tissue Culture Studies on Species of *Passiflora*. *Plant Cell, Tiss. Organ Cult.* 36:211-217.
- EI-Tarabily KA, Nassar AH, Hardy GE, St J, Sivasithamparam K (2009). Plant growth promotion and biological control of *Pythium aphanidermatum*, a pathogen of cucumber, by endophytic actinomycetes. *J. Appl. Microbiol.* 106(1):13-26.
- Fischer HP, Nyfeler R, Pachlatko JP (1992). New agrochemicals based on microbial metabolites. In new biopesticides. Proceedings of the Agricultural Biotechnology Symposium, Suweon, Korea, September

- 25, 1992. Kim, S.U., Ed. The Research Center for new Biomaterials in Agriculture, suweon, korea. pp. 17-54.
- Hassan AA, El-Barawy AM, El Mokhtar NM (2011). Evaluation of biological compounds of *Streptomyces* species for control of some fungal diseases. J. Amer. (4):752-760.
- Kane M (2003). Bacterial and fungal indexing of tissue cultures. <http://plant-c.cfans.umn.edu/listserv/1996/log9612/indexing.htm>.
- Murashige T, Skoog F (1962). A revised medium for rapid growth and bioassays with tobacco tissue cultures. *Physiol. Plant.* 1:474-497.
- Pal KK, McSpadden GB. (2006). Biological control of plant pathogens. The Plant Health Instructor. DOI: 10.1094/PHI-A-2006-1117-02.
- Raudales RE, McSpadden Gardener B. (2008). Microbial biopesticides for the control of plant diseases in organic farming. The Ohio State University, Fact Sheet, Agriculture and Natural Resources, pp. 1-5.
- Sabaratnam S, Traquair JA (2002). Formulation of a *Streptomyces* biocontrol agent for the suppression of *Rhizoctonia* damping-off in tomato transplants. *Biol. Cont.* 23(3):245-253.
- Sadeghi A, Hesani AR, Askari H, Qomi DN, Farsi M, Hervan EM (2009). Biocontrol of *Rhizoctonia solani* damping-off of sugar beet with native *Streptomyces* strains under field conditions. *Biocont. Sci. Technol.* 19(9/10):985-991.
- Samac DA, Kinkel LL (2001). Suppression of the root lesion nematodes (*Pratylenchus penetrans*) in alfalfa (*Medicago sativa*) by *Streptomyces* sp. *Plant and Soil* 235:35-44.
- Shori GBO, Mohamed SH, Abdel-Salam SM and Sadik AS (2012). Characterization of streptomycetes having antibiosis activities isolated from soil in western region of KSA. *Pakist. J. Biotechnol.* 9(1): 1-12.
- Smith BJ, Sampson BJ, Walter M (2012). Efficacy of bumble bee disseminated biological control agents for control of *Botrytis blossom* blight of rabbit eye blueberry. *Int. J. Fruit Sci.* 12(1/3):156-168.
- Strap JL, Crawford DL (2006). Ecology of *Streptomyces* in soil and rhizosphere. In: *Molecular approaches to soil, rhizosphere and plant microorganism analysis*, (Cooper, J. E.; Rao, J. R., eds), Editors Book ISBN 1-84593-062-2, 1:166-182.
- Zarandi ME, Bonjar GHS, Dehkaei FP, Moosavi SAA, Farokhi PR, Aghighi S (2009). Biological control of rice blast (*Magnaporthe oryzae*) by use of *Streptomyces sindeneusis* isolate 263 in greenhouse. *Amer. J. A. Sci.* 6(1):194-199.