African Journal of Cellular Pathology 7:1-5 (2016) The Official Journal of the Society for Cellular Pathology Scientists of Nigeria

ISSN 2449 - 0776 www.ajcpath.com

EFFECT OF PRENATAL ETHANOL EXPOSURE AND EXTRACT OF TAMARINDUS INDICA PULP ON THE CEREBRAL CORTEX OF WISTAR RATS

Usman IM¹, Buraimoh AA², Ibegbu AO³

1. Department of Human Anatomy, Faculty of Medicine, Ahmadu Bello University, Zaria, Nigeria

2. Department of Human Anatomy, Faculty of Medicine, Kaduna State University, Kaduna, Nigeria

3. Department of Human Anatomy, Faculty of Medicine, Federal University Dutse, Jigawa, Nigeria

Corresponding Author: Usman IM Email: gopama13@gmail.com

ABSTRACT

Aim: The aim of the study was to evaluate the effect of ethanolic pulp extract of Tamarindus indica (EPTI) on the cerebral cortex in prenatal ethanol exposure in Wistar rats.

Method: Twenty four pregnant rats were divided into 7 groups. The Control received 1ml of water, Groups 2 and 3 received 200mg/kg and 300mg/kg body weight (bw) of EPTI and Vitamin E respectively, Group 4 received 0.1ml of olive oil, Group 5 received 30% v/v of ethanol, Groups 6 and 7 received 30% v/v of ethanol and 200mg/kg bw of EPTI; and 30% v/v of ethanol and 300mg/kg bw of Vitamin E respectively. All administrations were via oral route, from prenatal day 7 to 14. At littering the brain tissues of the pups were collected for histological studies while some pups were used for Neurobehavioral studies.

Results: The result of elevated plus maze test showed significant increase in the time spent in the closed arm, rearing and grooming in Group 5 compared to the Control (p<0.05). Histological studies showed normal architecture in Groups 1, 2, 3 and 4, Group 5, 6 and 7 showed degenerative changes compared to the Control. **Conclusion:** Treatments with EPTI and Vitamin E have shown some potential protective effects on the Cerebral cortex of Wistar rats during prenatal ethanol exposure.

Key words: Cerebrum, Ethanol, Prenatal, Tamarindus indica

INTRODUCTION

Alcoholism and its related complication is one of the most common medical disorders (Musa et al., 2012). Maternal alcohol consumption during pregnancy may lead to the delivery of offsprings who may be diagnosed with fetal alcohol syndrome (FAS) (Maier and West, 2001). FAS characterized by growth deficiency, are microcephally and central nervous system dysfunction (May and Gossage 2001; Onu et al., 2014). Changes in central nervous system function may be the most sensitive signs of prenatal alcohol exposure (Clarren, 1982). The decrease in mental capacity and delayed maturation following prenatal ethanol exposure are associated with alteration in number and structures of neurons throughout the cerebral

cortex and other parts of the brain (Musa et al., 2012; Ahveninen et al., 2000; Sampson et al., 2000). Ethanol is capable of generating free radicals, thereby impairing the antioxidant defensive mechanisms in humans and experimental animals (Guidot and Duncan 2002). Many ethanol-induced adverse effects can be prevented or attenuated by antioxidants (Shirpoor et al., 2014). Tamarindus indicia, belongs to the Dicotyledonous family Leguminosae and Sub family Caesalpiniaceae, which is the third largest family of flowering plants with a total of 727 genera and 19,327 species (Lewis et al., 2005). Tamarindus indica has been used in the treatment of many diseases such as fever, dysentery, jaundice, gonococci and gastrointestinal disorders (Kobayashi et al.,

1996; Ferrara, 2005). All extracts of Tarmarindus indica exhibited good antioxidant activity (Siddhuraju 2007). The aim of the study was to evaluate the effect of EPTI on the cerebral cortex during prenatal ethanol exposure in Wistar rats.

MATERIALS AND METHODS Vitamin E and Ethanol Preparation

Capsules of Vitamin E from GLPL; Gujarat Liquid Pharmacaps Pvt. were cut open and emptied into a clean container. Olive oil was added to prepare a suspension containing 67 mg of the Vitamin E in 0.1 ml of the suspension. Vitamin E was protected from direct contact with air and sunlight to avoid degradation by stocking in a dark and air-tight jar. Absolute alcohol from Anala7 R analytical reagent; BDH Chemical Ltd Poole England were obtained and a stock solution of 30% v/v was prepared by diluting 30ml of absolute alcohol with 70ml of distilled water.

Plant Material

Tamarindus indica pulp was obtained from Samaru Market, Zaria, Kaduna State, and authenticated, with a voucher number of 602 in the Herbarium of the Department of Biological Science, Faculty of Sciences, Ahmadu Bello University, Zaria, Kaduna State. The extraction of the plant was carried out by maceration as outlined by Jindal et al., (2011).

Experimental Design

Twenty four pregnant timed rats were grouped into seven on the seventh day of gestation. Group 1: Made of 4 pregnant rats served as the Control Group and received 1ml of distilled water only. Group 2: Made of 3 pregnant rats were given 200mg/kg bw of EPTI only. Group 3: Made of 3 pregnant rats were given 300mg/kg bw of Vitamin E only. Group 4: Made of 4 pregnant rats were given 0.1ml of olive oil only. Group 5: Made of 4 pregnant rats were given 2ml of 30% v/v of ethanol only. Group 6: Made of 3 pregnant rats were given 2ml 30% v/v of ethanol and 300mg/kg bw of vitamin E. Group 7: Made of 3 pregnant rats were given 2ml 30% v/v of ethanol and 200mg/kg bw of ethanolic extract of Tamarindus indica pulp. All administrations were done orally by gastric intubation for seven days from the 7^{th} day to the 14^{th} day of gestation. On post gestation day zero, pups were

sacrificed humanely from each group for histological studies of the cerebral cortex, while others were used for neuro-behavioural test using elevated plus maze test on post gestation week 8 according to the methods of Shrestha and Singh (2013).

Neurobehavioral Studies

The apparatus consisted of two open arms (50 $cm \times 10$ cm) and two closed arms (50 cm $\times 10$ $cm \times 40$ cm) which were connected through a central platform (10 cm \times 10 cm). The arms were arranged in a cross shape with the two open arms facing each other and two closed arms facing each other. The maze was kept 45 cm above the floor. The test rat was placed at the center of the maze with its face directed towards one of the closed arms and observed for 5 minutes. The number of entries into closed arm and open arm, time spent in the open and closed arm, as well as number of rearing and grooming were observed. The floor and the walls of the open and closed arms were cleaned with 70% alcohol before each trial.

Histological Studies

The tissues were routinely processed as outlined by Culling (1981) and stained with H&E. Microphotographs were taken at $\times 400$ using MD900 Amscope digital camera.

Statistical Analysis

Data were presented as mean \pm SEM. For establishing significant differences, data were analyzed by one-way analysis of variance (ANOVA), followed by LSD post hoc test. Values were considered statistically significant when P value was ≤ 0.05).

RESULTS

Elevated Plus Maze Result

The result of the elevated plus maze test performed by the test rats on 8th week of postnatal life (Table 1) showed that the least number of entry into the open arm (NEOA) (3.0 ± 2.1) and the highest number of entries into the closed arm (NECA), the time spent in the closed arm (TSCA), rearing and grooming are 11.5 ± 3.51 , 271.2 ± 10.8 , 23.5 ± 13.10 and 24.0 ± 15.55 respectively were observed in Group 5. TSCA, rearing and grooming observed in Group 5 were significantly high when compared to the values observed in the Control Group as shown in Table 1.

Histological Study

Groups 1, 2, 3 and 4 showed a normal histological section with intact pyramidal cell body and neuroglia cells. Group 5 treated with ethanol showed evidence of degeneration i.e.

vacoulation and chromatolysis. On the other hand Group 6 and 7 showed less degenerative changes when compared to the Control group.

Table 1: Elevated plus maze result of rats from dams in the various treatment groups on the 8th week of postnatal life.

| GROUP | Treatment | NEOA/5min | TSOA(sec) | NECA/5min | TSCA(sec) | Rearing/5min | Grooming/5min |
|-------|--------------------------|--------------|------------------|----------------|--------------------|----------------------|--------------------|
| | | mean±SEM | mean±SEM | mean±SEM | mean±SEM | mean±SEM | mean±SEM |
| 1 | Control group | 5.0 ± 2.58 | 38.0 ± 5.65 | 6.7 ± 1.50 | $154.0{\pm}158.39$ | 13.7 ± 4.85 | 7.2 ± 1.25 |
| 2 | Extract (200mg/kg) | 5.5 ± 3.53 | 29.0 ± 10.9 | 7.0 ± 1.41 | 216.5 ± 80.05 | 13.5 ± 2.12 | 10.0 ± 4.08 |
| 3 | Vitamin E (300mg/kg) | 3.7 ± 1.70 | 36.0 ± 20.89 | 6.7 ± 1.50 | 236.0 ± 68.07 | 16.2 ± 4.64 | 11.2 ± 5.73 |
| 4 | Olive oil (0.1ml) | 6.0 ± 2.44 | 39.0 ± 24.52 | 8.6 ± 4.45 | 257.2 ± 25.06 | 13.7 ± 4.85 | 17.5 ± 12.81 |
| 5 | Ethanol (30% v/v) | 3.0 ± 2.16 | 32.7 ± 14.40 | 11.5 ± 3.51 | $271.2\pm10.8*$ | $23.5 \pm 13.10^{*}$ | $24.0 \pm 15.55 *$ |
| 6 | Ethanol and vitamin E | 3.7 ± 2.60 | 36.0 ± 20.89 | 9.0 ± 6.00 | 264.0 ± 20.89 | 16.2 ± 2.98 | 14.5 ± 6.35 |
| 7 | Ethanol and extract | 5.6 ± 4.98 | 42.8 ± 25.06 | 11.5 ± 3.41 | 267.2 ± 14.40 | 14.4 ± 4.56 | 16.2 ± 13.0 |

NEOA: Number of entries into open arm, TSOA: Time spent in the open arm, NECA: Number of entries into closed arm, TSCA: Time spent in the closed arm. * represents significance when compared to the Control ($p \le 0.05$).

DISCUSSION

The elevated plus maze test which is a typical test widely employed to study potential anxiolytic and antidepressant compound in animal model (Acevado et al., 2014). Anxiety like behavior is measured as preference for the closed arm or less activity (entry or duration) in the open arm (Rodgers and Johnson, 1995), Grooming and Rearing are also associated with anxiety like behaviors in the elevated plus maze test (Espejo, 1997). The results of the elevated plus maze test showed a significant increase in TSCA, grooming and rearing in Group 5 when compared to the Control on the 8th week of post gestational life, indicating that prenatal ethanol exposure in this present study was associated with anxiety-like behavior. This observation is in line with the findings of Dursun et al., (2006), who found increased anxiety-like behavior in the EPM and open field tests, respectively, in young adult (PN80-85) offspring after prenatal exposure to high doses of ethanol (6 g/kg, GD7-20). Brocardo et al., (2012) found that exposure to high doses of ethanol during gestation (4.3 g/kg, GD1-22) and the early postnatal period (4 g/kg, PN4-10) resulted in significantly decreased open arm exploration in the EPM in both male and female young adult (PN60) offspring. Cullen et al., (2013) also observed that exposure of pregnant Dawley rat to ethanol (6% v/v) throughout gestation was associated

with anxiety like behavior at 8 month and 15 month of post gestational life. Treatment with vitamin E and ethanol pulp extract of Tarmarindus indica did not show significant difference when compared to the Control Group. This observation could be associated with the protective effect accrued to antioxidants. The result of the histological studies showed that prenatal ethanol exposure was associated with neuro-degenerative changes such as pyknosis and vacoulation. These observations are in line with the findings of Iqbal et al., (2004) and Allam and Abdul-hamid (2013), who reported similar changes as a result of prenatal ethanol exposure. On the other hand treatments with ethanol pulp extract of Tarmarindus indica and Vitamin E showed some protection. This is in line with the finding of Zhu et al., (2007) who that Vitamin E administration reported prevented oxidative stress and tissue damage caused by ethanol consumption in the brain, with Tarmarindus indica also appearing on the scene. The observed effect may be due to their antioxidant properties as well as non-antioxidant dependent activities (Shirpoor et al., 2014).

CONCLUSION

Treatments with EPTI and Vitamin E have been shown to have potential protective effect on the cortex of Wistar rats during prenatal ethanol exposure.



Plate 1: Sections of cerebral cortex of pups in Group 1, Group 2, Group 3 and Group 4, showing pyramidal cell body and glia cell. Groups 5, 6 and 7, showed evidence of degeneration; vacuolation and pyknosis (H and E; $\times 400$).

REFERENCES

Acevedo MB, Nizhnikov ME, Molina JC, Pautassi RM (2014). Relationship between ethanol-induced activity and anxiolysis in the open field, elevated plus maze, light-dark box, and ethanol intake in adolescent rats.Behavioral Brain Research, 265: 203–215

Ahveninen J, Escera C, Polo MD, Grau C, Jääskeläinen IP (2000). Acute and chronic effects of alcohol on Preattentive auditory processing as reflected by mismatch negativity. Audiology Neurotology, 5: 303-311.

Allam A, Abdul-Hamid M (2013). Effect of ethanol ngestion in the pregnant albino rats on the development of pyramidal neurons.Life Science Journal, 10(4).241-247.

Brocardo PS, Boehme F, Patten A, Cox A, Gil-Mohapel J, Christie BR (2012). Anxiety and depression-like behaviors are accompanied by an increase in oxidative stress in a rat model of fetal alcohol spectrum disorders: Protective effects of voluntary physical exercise. Neuropharmacology 62: 1607–1618.

Clarren, S.K. (1982). The diagnosis and treatment of fetal alcohol syndrome. Comprehensive Therapy, 8:41–46.

Cullen CL, Burne TH, Lavidis NA, Moritz KM (2013). Low dose prenatal ethanol exposure induces anxiety-like behaviour and alters dendritic morphology in the basolateral amygdala of rat offspring. PLoS ONE, 8(1), e54924.

http://doi.org/10.1371/journal.pone.0054924

Culling CF (1981). Handbook of Hispathology and histochemical technique including museum techniques, Butterworths, London.

Dursun I, Jakubowska-Dogru E, Uzbay T (2006). Effects of prenatal exposure to alcohol on activity, anxiety, motor coordination, and memory in young adult Wistar rats. Pharmacology Biochemistry and Behavior 85: 345–355.

Espejo EF (1997). Effects of weekly or daily exposure to the elevated plus-maze in male mice.Behavioural Brain Research, 87, 233-238.

Ferrara L (2005). Antioxidant activity of Tamarindusindica L., Ingredient alimentary, 4(6): 13-15.

Guidot DM, Duncan J (2002). Chronic ethanol ingestion increases susceptibility to acute lung injury of oxidative stress and tissue remodeling. Chest Journal, 122: 309-314.

Iqbal, U, Dringenberg HC, Brien JF, Reynolds JN (2004). Chronic prenatal ethanol exposure alters hippocampal GABAA receptors and impairs spatial learning in the guinea pig. Behavioural Brain Research. 150:117-125.

Jindal V, Dhingra D, Sharma S, Parle M, Harna RK (2011). Hypolipidemic and weight reducing activity of the ethanolic extract of Tamarindus indica fruit pulp in cafeteria diet- and sulpiride-induced obese rats. J. of Pharmacology and Pharmacotherapeutics, 2(2), 80–84.

Kobayashi A, Adenan ML, Kajiyama SI, Kanzaki H, Kawazu K (1996). A cytotoxic principle of Tamarindusindica, di-n-butyl malate and the structure-activity relashionship of its analogues. Journal of Biosciences, 51(3-4): 233-242.

Lewis G, Schrire B, Mackinder B, Lock M (2005). Legumes of the World.Royal Botnaic Gardens, Kew.

Maier SE, West JR (2001). Regional differences in cell loss associated with binge-like alcohol exposure during the first two trimester's equivalent in the rat. Alcohol, 23: 49–57.

Marcondes FK, Bianchi FJ, Tanno AP (2002). Determination of the estrous cycle phases of rats: some helpful considerations. Brazilian Journal of Biology, 62:609-614.

May PA, Gossage JP (2001)."Estimating the prevalence of fetal alcohol syndrome.A summary."Alcohol Research Health Journal, **25**(3): 159–67.

Musa AS, Ibrahim S, Umana UE, Adebisi SS, Hamman WO (2012). Pathological Lesions in the Lungs of Neonatal Wistar Rats from Dams Administered Ethanol during Gestation. Asian Journal of Medical Sciences, 4(1): 4-7. Onu JE, Oke BO, Ojegbe PC, Oyewale JO (2014). Morphological alteration of seminiferous tubule of the testes of wistarrats offspring exposed to alcohol during pregnancy and/ or lactation. International Journal of Biology and chemical Science.54-356.

Rodgers RJ, Johnson NJ (1995). Factor analysis of spatiotemporal and ethological measures in the urine elevated plus-maze test of anxiety. Pharmacology Biochemistry and Behaviour 52: 297-303.

Sampson PD, Streissguth AP, Bookstein FL, Barr H. (2000). On categorizations in analyses of alcohol teratogenesis. Environmental Health Perspectives, 108: 421-428.

Shirpoor A, Norouzi L, Khadem-Ansari MH, Ilkhanizadeh B, Karimipour M (2014). The Protective Effect of Vitamin E on Morphological and Biochemical Alteration Induced by Pre and Postnatal Ethanol Administration in the Testis of Male Rat Offspring: A Three Months Follow-up Study. Journal of Reproduction & Infertility, 15(3), 134-141.

Shrestha U, Singh M (2013). Effect of folic acid in prenatal alcohol induced behavioral impairment in Swiss albino mice. Annals of Neurosciences, 20(4), 134–138.

Siddhuraju P (2007). Antioxidant activity of polyphenolic compounds extracted from defatted raw and dry heated Tamarindusindica seed coat. LWT Food Science and Tech, 40: 982 - 990.

Zhu Q, Emanuele MA, LaPaglia N, Kovacs EJ, Emanuele, N.V. (2007). Vitamin E prevents ethanol-induced inflammatory, hormonal, and cytotoxic changes in reproductive tissues. Endocrine, 32(1):59-68.