

Review

Wheat bran as a brown gold: Nutritious value and its biotechnological applications

Muhammad Mohsin Javed*, Sana Zahoor, Sarah Shafaat, Iffat Mehmooda, Ambreen Gul, Huma Rasheed, Syed Ali Imran Bukhari, Muhammad Nauman Aftab and Ikram-ul-Haq

Institute of Industrial Biotechnology, GC University, Lahore, Pakistan.

Accepted 29 December, 2011

Wheat bran, a by-product of wheat milling industry, is the outermost covering of wheat grain. It has relatively diverse application in food, feed, medicine and fermentation industries due to its richness in carbohydrates (mostly fibers), protein and fats which makes it an important dietary element. Its importance from medical view point is well documented especially in reducing blood plasma cholesterol and prevention of cardiovascular diseases and certain cancers, that is, colon cancer. In fermentation industry, wheat bran can be used as a substrate for production of a range of products such as biomass, enzyme, biofuel and production of other metabolites. In short wheat bran has immense applications and market value which makes it so important to be considered as brown gold.

Key words: Wheat bran, enzyme production, biofuel, food, health.

INTRODUCTION

Worldwide consumption of wheat, estimated by World agricultural supply and demand estimates (WASDE), has been found to be 652.18 million ton for year 2010 (WASDE, 2010). Bran is created as a by-product in milling industries (Hemery et al., 2007) and one million tons of wheat can produce upto 0.25 million tons of wheat bran (WB). WB is produced abundantly in all agricultural countries like Pakistan. WB production is coupled to the production of wheat. Wheat (*Triticum aestivum*) is an ancient known food crop, cultivated since the beginning of human civilization and ranks first among world cereal crops). Production of wheat is closely related to the supply of irrigation water and amount of rain fed water (Ahmad et al., 2010).

Although WB production is mainly dependent upon the nature of the soil (Safdar et al., 2009) and water supply but approximate yield of wheat bran is 14 to 19% of

wheat kernel (Safdar, 2005). Production of WB is also dependent upon the choice of milling procedure and the physical characteristics of wheat bran tissue. More nutritionally active wheat bran fractions after debranning and degerning can be obtained by considering physical characteristics like rheology and dielectric constant. Adoption of more advanced milling techniques give rise to a quality product with more retention of the nutrient compounds and removal of the unwanted substances from the environmentally exposed outer layer of wheat. Wheat and wheat based products are primarily manufactured for human consumption (Hemery et al., 2007).

WHEAT BRAN COMPOSITION

Wheat kernel is made up of three major parts; seed coat or pericarp (bran), endosperm and germ (Hoseney, 1994). Detailed study by Safdar (2005) about the structure of wheat kernel shows that it contains about 68 to 80% endosperm, 14 to 19% bran and 2 to 3% germ. Antoine et al. (2002) have found that WB is a composite material made up of three discrete layers that are formed from numerous histological tissues. These tissue layers

*Corresponding author. E-mail: mmj_bot@yahoo.com. Tel: +92 321 5794762. Fax: +92 321 5794762.

Abbreviations: WB, Wheat bran; SSF, solid state fermentation; SmF, submerged fermentation, LDL, low density lipoprotein.

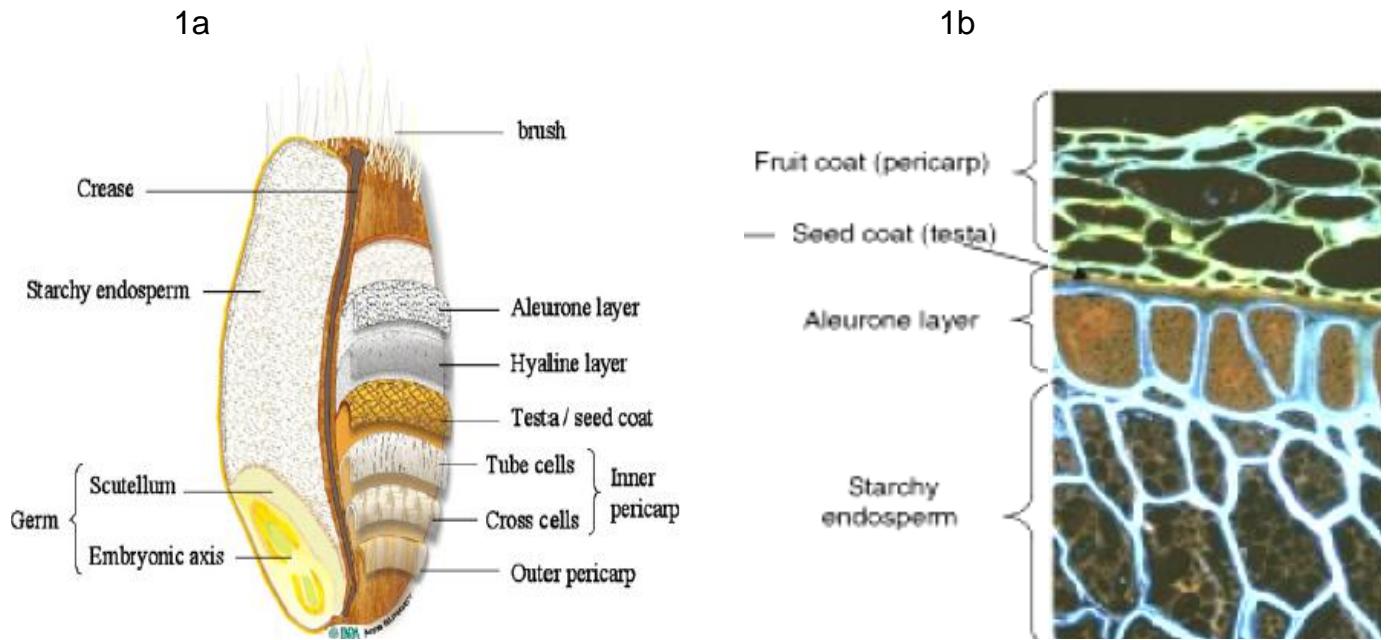


Figure 1 a). Histological composition of wheat grain. Adopted from Surget and Barron (2005), b) Microstructure of parts of intact grains of wheat. The sections have been stained with Acid Fuchsin and Calcofluor: protein appears red, cell walls rich in β -glucan appear light blue and lignified cell walls of the fruit coat appear yellowish-brown (adapted from Kamal-Eldin et al., 2009).

Table 1. Composition of wheat bran (adopted from Palmarola-Adrados et al., 2005).

Components	g/100g Wheat bran
a) Non-starch polysaccharide	
Glucan	10.5
Xylan	18.3
Arabinan	10.1
Galactan	1.1
b) Starch	34.0
c) Klason lignin	5.0
d) Crude protein	13.5
Total	92.5

are divided into outer and inner pericarp (tube cells and cross cells), testa or seed coat, hyaline layer and aleurone layer (Figure 1a and b).

WB is rich in carbohydrates (60%), protein (12%), fat (0.5%), minerals (2%), bioactive compounds and vitamins (Slavin, 2003) (Table 1). Along with these wheat bran also contains several important compounds such as phenolic acids, carotenoids, lignans, phytosterols, flavonoids, α -tocopherol and phytic acid which are distributed unequally in different WB tissues (Table 2). Precise composition of macro and micronutrients may vary from cultivar to cultivar and the extraction technique of these compounds from bran. Wheat ash content is usually measured for quantification of bran. Amount of ash (mineral content) present is a true reflective of the bran quantity in wheat (Safdar et al., 2009).

Pericarp

The outermost covering pericarp is rich in insoluble dietary fiber (cellulose, cuticle material and complex xyloans with high arabinose to xylose ratio), lignin, ferulic acid and other bioactive compounds (Hemery et al., 2007). Likes et al. (2007) has investigated the presence of two important bioactive compounds; betaine (an osmolyte and methyl donor) and choline in WB and found that betaine is concentrated more in the bran fraction (1293.3 mg/100g flour) than the germ fraction (1163.5 mg/100g of flour) of wheat (Waggle et al., 1967).

Aleurone layer

Aleurone layer is rich in minerals, most of B vitamins and

Table 2. Location of nutrients in wheat bran layers.

Wheat bran layer	Nutrients	Reference
The pericarp	Insoluble dietary fiber Ferulic acid Bioactive compounds Vitamins	Hemery et al. (2007).
Aleurone layer	Minerals B vitamins Protein Lignans Bioactive compounds Antioxidant Phytate Phenolic acids	Pomeranz, (1988); Antoine et al. (2002) Rhodes and Stone, (2002); Buri et al. (2004). Slavin et al. (1999) Zhou and Yu (2004); Esposito et al. (2005)
Testa tissue	Alkylresorcinols	Landberg et al. (2008)

covers about 7% of wheat grain dry mass (Antoine et al., 2002). This layer is also rich in protein and contains a good balance of amino acids than the endosperm (Rhodes and Stone, 2002; Buri et al., 2004). Lignans are also enriched in the aleurone layer of WB (Buri et al., 2004). Being an agro-economical residue it is a good source of bioactive compounds, essential vitamins and most importantly the antioxidant compounds. Other compounds that are enriched in the aleurone layer are phytate (Slavin et al., 1999) and phenolic acids (Zhou and Yu, 2004; Esposito et al., 2005). Although the presence of various phenolic acids (vanillic, syringic, *p*-coumaric, *o*-coumaric, caffeic, and *p*-hydroxybenzoic, gallic, and gentistic acids) has been reported (Zhou et al., 2004a; Zhou et al., 2004b; Zhou and Yu, 2004; Kim et al., 2006) but the major one is the ferulic acid with a range of 1550 to 2119 mg/g bran (Li et al., 2005).

Testa tissue

According to Landberg et al. (2008) almost all alkylresorcinols (a phenolic lipid) has been present in the testa tissue of WB. The detailed composition of the three wheat bran layers is described in Table 2.

ROLE OF WHEAT BRAN

Fermentation industry

Many agro-industrial by-products are replacing the synthetic and expensive substrates for the production of biotechnological products. Among the agro-industrial substrates, WB is one of the most attractive alternatives to synthetic medium in fermentation processes (Pandey, 1992). The coarse variety of WB is an efficient substrate due to its heat dissipation, better air circulation, loose

particle binding and efficient penetration by mycelia and it is cheaper than fine bran so it is a better prospect economically in fermentation industry (Malathi and Chakraborty, 1991). Almost every type of enzyme can be produced by fermentation of WB both by utilizing solid state fermentation (SSF) and submerged fermentation (SmF) systems (Table 3). Now a days WB is widely used in solid state fermentation for the production of secondary metabolites and other industrial products because it reduces pollution affects (Pandey et al., 1999). Wheat bran is a potential candidate in fermentation industry because of its unique properties listed below:

Water retaining ability

Apart from the presence of important nutritional components, physical characteristics of wheat bran also play vital role in fermentation process. WB has the ability to retain high moisture content in SSF. This ability of wheat bran promotes the fungal growth just as in the natural environmental conditions. Wheat bran proved a suitable substrate for the growth of *Trichoderma harzianum*, *Trichoderma viride*, *Trichoderma koningii*, and *Trichoderma polysporum* by SSF. There was no need of additional nutrients in WB medium for the production of *Trichoderma* spores (Cavalcante et al., (2008).

Complex substrate

Complex nature of WB lies in its unique nutrient composition. The higher starch content of WB i.e., 75.6% as compared to other agro-industrial wastes such as rice bran (coarse waste 71.1%> rice powder 55.8%> medium waste 48.6%> fine waste 34.2%) can be correlated for

Table 3. Enzyme production on wheat bran.

Enzymes	Producer microorganism	Enzyme activity	Reference
Proteases			
	<i>Aspergillus sp. and Mucor pusilus</i>	--	Sumantha et al. (2006)
Acid Protease	<i>Aspergillus oryzae</i>	--	Tsujita and Endo (1976); Sandhya et al. (2005)
Neutral metallo protease	<i>Aspergillus oryzae NRRL 2217</i>	--	Sumantha et al. (2005)
Acidic	<i>Aspergillus oryzae MTCC 5431</i>	8.26×10^5 U/g cell mass	Vishwanatha et al. (2009)
Alkaline	<i>Thermoactinomyces thalophilus PEE14</i>	1620 U/g cell mass	Divakar et al. (2004)
α- amylase			
	<i>Bacillus licheniformis</i>	--	Ikram-ul-Haq et al. (2003)
	<i>Bacillus cereus MTCC 1305</i>	122 U/g cell mass	Anto et al. (2006a)
	<i>Aspergillus niger</i>	--	Kaur et al. (2003)
	<i>Aspergillus awamori: Nakzawa MTCC 6652</i>	9420 U/g cell mass	Negi and Banerjee (2006)
Lipase			
	<i>Aspergillus niger</i>	4.8 IU/ml	Falony et al. (2006)
	<i>Bacillus megatarium AKG 1</i>	--	Sekhon et al. (2004)
	<i>Alkalophilic Yeast</i>		Bhushan et al. (1994)
Pectinase			
	<i>Streptomyces lydicus</i>	--	Jacob and Prema (2006)
	<i>Bacillus sp. DT7</i>	8050 IU/g cell mass	Kashyap et al. (2003)
	<i>Aspergillus niger</i>	36.3 IU/g cell mass	Debing et al. (2006)
	<i>Penicillium vermiculatum RFC3</i>	100 IU/ml	Silva et al. (2005)
	<i>Aspergillus foetidus NRRL 341</i>	1860 IU/g cell mass	Cavalitto et al. (1996)
Glucoamylase			
	<i>Aspergillus HA-2</i>	264 U/g cell mass	Anto et al. (2006b)
	<i>Aspergillus sp.</i>	454 U/g cell mass	Ellaiah et al. (2002)
	<i>Aspergillus oryzae</i>	1986 U/g cell mass	Zambare (2010)
Xylanase			
	<i>Aspergillus niger</i>	33%	Dobrev et al. (2007)
	<i>Trichoderma longibrachiatum</i>	592.7 U/g cell mass	Azin et al. (2007)
	<i>Bacillus sp. AR009</i>	720 U/g cell mass	Gessesse and Mamo (1999)
	<i>Bacillus licheniformis A99</i>	1.63 U/g cell mass	Archana and Satyanarayana (1997)
	<i>Aspergillus niger XY-1</i>	14637 U/g cell mass	Xu et al. (2008)

Table 3. Cont.

Cellulase	<i>Staphylococcus sp. SG-13</i>	4525 U/l	Gupta et al. (2001)
	<i>Trichoderma reesei RUT C30</i>	--	Sukumaran et al. (2009)
	<i>T. viride CMGB</i>	3.18 FPU/ml	Vintila et al. (2009)
Tannase	<i>Paecilomyces variotii</i>	--	Battestin and Macedo (2007)
	<i>Aspergillus aculeatus DBF9</i>	8.4 U/g cell mass	Banerjee et al. (2007)

higher amylase production (Ellaiah et al., 2002). WB can be used as an inducer for a multitude of enzymes such as CMCCase, xylosidase, glucosidase, α -L-arabinofuranosidase, amylase, protease, pectinolytic enzymes, rennet, alpha galactosidase, lipase, invertase and phytase (Maheswari and Chandra, 2000; Sindhu et al., 2009; Soarse et al., 2010; Javed et al., 2011). Xylanase production on commercial scale can also be achieved by using WB as a substrate as it is an agro-economical inducer due to its high xylan content (12.65% of dry material) (Kulkarni et al., 1999; Subramanyan and Prema, 2002). Battestin and Macedo (2007) studied tannase production by *Paecilomyces variotii* and reported that the presence of important mineral contents of WB was essential for the mold growth resulting in an 8.6 fold increase in tannase production.

Nitrogen source

Naturally, higher amount of the nitrogen also requires no or little addition of other nitrogen supplements in WB containing medium. Elevated nitrogen content of WB makes it suitable for the production of enzymes such as protease, amylase

and glucoamylase. Increase in the production of acid protease with an increase in the nitrogen content of WB has been reported by Vishwanatha et al., 2009. Supplementation of WB with additional protein sources such as soy flour, defatted sesame flour, casein and peptone facilitate acid protease production. Although WB alone can be used as an efficient nitrogen source but supplementation of WB with glucose, peptone, yeast extract, KH_2PO_4 and CaO resulted in the highest spore production 1.7×10^{11} spore/g dry substrate (Vishwanatha et al., 2009).

Metabolite production: WB has been used for the production of various interesting metabolites. Bacitracin was produced by *Bacillus licheniformis* using WB and soya bean in 1 : 3 (Farzana et al., 2005). Ten folds increase in the production of cyclosporine-A was observed in SSF utilizing WB than in SmF (Sekar et al., 1997). Bandelier et al. (1997) reported the production of an important plant hormone, gibberellic acid by *Gibberella fujikoroi* using WB in SSF.

Biofuel production: To deal with the problem of depletion of fossil fuel reserves with each passing

years, researchers are now focusing on bioethanol production from natural substrates to meet energy challenges of the millennium (Shafiee and Topal, 2009). Wheat milling by products are now being focused for fermentative production of bioethanol (Palmarola-Adrados et al., 2005; Hawkes et al., 2008; Manikandan and Viruthagiri, 2009). Manikandan and Viruthagiri (2009) investigated the simultaneous saccharification and fermentation of WB that resulted in highest ethanol concentration of 23.1 g/L after 48 h of fermentation. Palmarola-Adrados et al. (2005) converted the complex polysaccharides in WB to sugar rich feedstock for conversion to ethanol. The overall sugar yield by combined hydrolysis method (acid treatment and enzymatic hydrolysis) reached 80% of the theoretical and it consisted of 13.5 g arabinose, 22.8 g xylose and 16.7 g glucose per 100 g starch-free bran. In addition to using WB as sole source for ethanol production, it can also be used as a nutritive supplement for ethanol production by *Zymomonas mobilis* rather than the synthetic supplements (Shamala and Sreekanthiah, 1988). Moreover WB can also act as potential substrate for biobutanol production, which may be used as a replacement for gasoline (Liu et al., 2010).

Fermentative bio-hydrogen production from carbohydrate-rich substances can be achieved through anaerobic digestion by bacteria (Pam et al., 2006). Acid treatment of WB followed by anaerobic digestion with mixed anaerobic culture resulted in maximum hydrogen yield of 128.2 ml/g total volatile solid (TVS) and hydrogen production rate of 2.50 ml/ (g-TVS h). Maximum hydrogen content was 62% with negligible methane production (Pan et al., 2008). Treatment of WB with NaOH and H₂O₂ and then fermentation with mixed culture in sewage sludge produced 22 and 31 m³ H₂ per ton dry weight assuming that all the sugar is hexose. Fermentation of unhydrolysed wheat feed is also known to improve H₂ yield (Hawkes et al., 2008).

Bioremediation

The removal of heavy metal ions from abandoned industrial sites is a major challenge for decontamination and rehabilitation of industrial wastewaters. Presence of heavy metal ions such as Cu (II), Pb (II), and Cd (II) is a potential threat for human health. The use of lignocellulosic compounds from wheat bran for the removal of these heavy metal ions offers a cheap and flexible substrate. WB contains lignin, cellulose and fatty acid units whose functional group content (hydroxylic, carboxylic and phenolic) is ideal for ion fixation. It can be used as a natural filter for decontaminating industrial effluent containing heavy metals. This method can be a cheap alternative to conventional pollution control methods for wastewater by use of synthetic resins for heavy metal adsorption (Dupont et al., 2003).

Health aspects

Compounds (such as vitamins, lignans, phenolic acids and alkylresorcinols) present in bran are desirable ingredients to be added in daily diet to improve the nutritional value of food. So WB can be used as natural and cheaper source of value added products in preparation of functional food ingredients or for fortification of certain products. The quality, safety and stability of food products can be enhanced by adding antioxidants in the food. These antioxidants can terminate the free radical chain reactions and can reduce the incidence of diseases such as cancer, cardiovascular disease, parkinson's disease, cell injury, cell death, and slow down the aging process, when these are consumed in the human diet (Halliwell, 1996). Lignan metabolites act as antioxidants and play a role in antitumor activities resulting in the control of estrogen level (Qu et al., 2005).

Soluble and insoluble dietary fiber of WB is known to lower blood plasma cholesterol, serves as an effective laxative and plays a role in colon cancer prevention (Lupton and Turner, 1999; Topping, 2007). Several

epidemiological studies have revealed that the risk of cardiovascular disease can be reduced by consuming bran based products (Halliwell et al., 1992; Gorinstein et al., 1998; Sabovic et al., 2004; Willcox et al., 2004; Minhajuddin et al., 2005; Jensen et al., 2006). Antioxidant compounds present in WB (Table 4) play a vital role in reducing the risk factor for different diseases by a variety of preventive mechanisms (Flight and Clifton, 2006). The predominant phenolic acid in wheat grain is ferulic acid, which has the potential to restore endothelial function in aortas of spontaneously hypertensive rats and to prevent trimethylin-induced cognitive dysfunction in mice (Kim et al., 2007; Suzuki et al., 2007). Oxidation of low density lipoprotein (LDL) cholesterol induced by copper can lead to atherosclerosis and ultimately the coronary heart disease. WB phenolic's binding to apo-lipoprotein has been shown to hinder the copper binding to LDL (Satue-Gracia et al., 1997) and also in copper removal from LDL surface (Decker et al., 2001). Craig (2004) demonstrated the protective role of betaine and choline against osmotic stress in internal organs and improvement of vascular risk factors. Protective role of betain against coronary heart disease has also been evidenced (Craig, 2004).

WB is also a rich source of lipid-soluble compounds and phytochemicals such as phytate, phytosterols, tocopherols etc. Among which phytates play an important role in prevention of colon cancer (Reddy et al., 2000). Sang et al. (2006) studied the effect of WB oil and its subfractions in mouse model of human colon cancer cell lines and demonstrated the intestinal cancer preventive activity of wheat bran oil. Reddy et al., (2000) also demonstrated that oil fraction of WB contains bioactive compounds that inhibits colon carcinogenesis.

From the facts it can be deduced that addition of whole grain or bran containing diet exerts certain metabolic benefits and reduces the risk of diabetes (Jensen et al., 2004) cardiovascular diseases (Pereira et al., 2002; Jensen et al., 2004), colon cancer risk (Lupton and Turner, 1999; Reddy et al., 2000; Freudenberg et al., 1990), hypertension (Whelton et al., 1997) and coronary heart disease (Craig, 2004).

As food and feed additive

20% WB supplementation to flour and baked product offer an inexpensive and interesting alternative to the synthetic vitamins and nutrient supplements for poor people by adopting some pretreatment procedures (Gomez et al., 2003). Pretreatment (grinding to obtain smaller particle size, heat treatment, pre-fermentation with yeast and lactic acid bacteria and extrusion) of bran improves the nutritional as well as physical quality of bread and baked products (De Kock et al., 1999; Salmenkallio-Marttila et al., 2001).

WB is a very acceptable raw material for animal feed preparations. Addition of 30% wheat bran and 30% rice

Table 4. Concentration of antioxidants in wheat bran.

Antioxidant	Concentration ($\mu\text{g}/100 \text{ g}$)	Reference
Lignans		
lignan aglycones	2774	
Syringaresinol	1953	
Isolariciresinol	297	
Lariciresinol	257	Begum et al. (2004)
Secoisolariciresinol	142	
pinoresinol	106	
matairesinol	9.4	
Carotenoids	0.68	
Lutein	1.8	Adom et al. (2003)
Zeaxanthin	0.54–27	
Tocopherols		
α tocopherols,	1.28–21.29	
δ tocopherols,	0.23–7.0	Zhou and Yu (2004)
γ tocopherols,	0.92–6.90	

bran in daily feed of cows improved the daily milk yield up to 14.65 and 12.87 L respectively. Results proved that wheat bran is nutritionally better than rice bran for the lactation of Holstein Friesian cows (Tahir et al., 2002). Saima et al. (1999) reported the production of biomass protein by growing *Candida utilis* and *Brevibacterium fluvium* strains on wheat bran. Crude protein (35.97%) and true protein (30.18%) was measured in biomass. Furthermore, chicks fed on biomass protein gained comparable protein efficiency ratios and meat protein than the fish meal protein.

ECONOMIC ASPECT AND FUTURE RECOMMENDATIONS

Each year million tons of wheat-milling by-product that is, bran has been produced but only a little is consumed as a food supplement. 24.214 million tons of wheat was produced in the year 2010 to 2011 in Pakistan which can ideally yield 6.0 million tons of wheat bran. By considering the cost of wheat as \$200/ton, thousands of dollars can be earned by WB inclusion in fermentation industry and several value added fermented products can also be brought into market with remarkable low price.

There is a strong emphasis that wheat should be consumed as whole grain flour (including the bran) or the bran should be supplemented in various bakery products to enrich the daily consumables with an inexpensive nutrient reservoir. In addition being a rich source of fiber it also serves as a solid inert matrix for absorption of heavy metal ions in the filtration plant manufacturing. These properties of WB has made it importance upto a million dollar product. If an effective strategy is applied then this

bran can be exploited for fermentation business and food supply for health benefits. Economics of dairy and poultry industries can be improved by utilizing wheat bran pellets as cattle and broiler chicks feed. As a result it can not only support the economics sector but in addition it will prove to be a cheap raw material for production of various products of fermentation industry.

Comments

Wheat is the most important food component around the globe. It is commonly used in refined form excluding its outer husk 'The Bran'. Each year millions and tons of bran has been produced as a by-product of wheat milling industry. Wheat bran is rightly known to be the gold product and finds its applications not only in fermentation industry but also in pharmaceuticals and biomedical research. Pakistan, being an agricultural country, produces tons of wheat each year. Wheat bran from wheat-milling industries in our country can serve as a potential nutritious and cheap raw material for fermentation industry while its anti-oxidative and anti-inflammatory properties can bring about a revolution in the field of melanoma research. Hence, it is rightly known to be the brown gold for its amazing properties.

REFERENCES

- Adom KK, Sorrells ME, Liu RH (2003). Phytochemical profiles and antioxidant activity of wheat varieties. *J. Agric. Food Chem.*, 51: 7825–7834.
- Ahmad M, Ghafoor A, Asif M, Farid HU (2010). Effect of irrigation techniques on wheat production and water saving in soils. *Soil Environ.*, 29(1): 69–72.

- Anto H, Trivedi U, Patel K (2006a). Alpha Amylase Production by *Bacillus cereus* MTCC 1305 using solid-state fermentation. *Food Technol. Biotechnol.*, 44: 241–245.
- Anto H, Trivedi UB, Patel KC (2006b). Glucoamylase production by solid-state fermentation using rice flake manufacturing waste products as substrate. *Bioresource Technol.*, 97: 1161–1166.
- Antoine C, Lullien-Pellerin V, Abecassis J, Rouau X (2002). Nutritional interest of the wheat seed aleurone layer. *Sci. Aliment.*, 22: 545–556.
- Archana A, Satyanarayana T (1997). Xylanase production by thermophilic *Bacillus licheniformis* A99 in solid-state fermentation. *Enzyme Microb. Tech.*, 21: 12–17.
- Azin M, Moravej R, Zareh D (2007). Production of xylanase by *Trichoderma longibrachiatum* on a mixture of wheat bran and wheat straw: Optimization of culture condition by Taguchi method. *Enzyme Microb. Tech.*, 40: 801–805.
- Bandelier S, Renaud R, Durand A (1997). Production of gibberellic acid by fed-batch solid state fermentation in an aseptic pilot-scale reactor. *Process Biochem.*, 32: 141–145.
- Banerjee D, Mondal KC, Pati BR (2007). Tannase production by *Aspergillus aculeatus* DBF9 through solid-state fermentation. *Acta Microbiol. Hung.*, 54: 159–166.
- Battestini V, Macedo GA (2007). Tannase production by *Paecilomyces variotii*. *Bioresource Technol.*, 98: 1832–1837.
- Begum AN, Nicolle C, Mila I, Lapierre C, Nagano K, Fukushima K, Heinonen SM, Adlercreutz H, Remesy C, Scalbert A (2004). Dietary lignins are precursors of mammalian lignans in rats. *J. Nutr.*, 134: 120–127.
- Bhushan B, Dosanjh NS, Kumar K, Hoondal GS (1994). Lipase production from alkalophilic yeast by solid state fermentation. *Biotechnol. Lett.*, 16: 841–842.
- Buri RC, von Reding W, Gavin MH (2004). Description and characterization of wheat aleurone. *Cereal Food World*, 49: 274–282.
- Caalcante RS, Lima HLS, Pinto GAS, Gava CAT, Rodrigues S (2008). Effect of moisture on *Trichoderma* conidia production on corn and wheat bran by solid state fermentation. *Food Bioprocess Technol.*, 1: 100–104.
- Cavalalito SF, Jorge AA, Hours RA (1996). Pectinase production profile of *Aspergillus foetidus* in solid state cultures at different acidities. *Biotechnol. Lett.*, 18: 251–256.
- Craig SAS (2004). Betaine in human nutrition. *Am. J. Clin. Nutr.*, 80: 539–549.
- De Kock S, Taylor J, Taylor JRN (1999). Effect of heat treatment and particle size of different brans on loaf volume of brown bread. *Lebensmittel Wissenschaft Technol.*, 32: 349–356.
- Debing J, Peijun L, Stagnetti F, Xianzhe X, Li L (2006). Pectinase production by solid fermentation from *Aspergillus niger* by a new prescription experiment. *Ecotoxicol. Environ. Saf.*, 64: 244–250.
- Decker EA, Ivanov V, Zhu BZ, Frei B (2001). Inhibition of low-density lipoprotein oxidation by carnosine and histidine. *J. Agric. Food Chem.*, 49: 511–516.
- Divakar G, Sunitha M, Vasu P, Udaya shanker P, Ellaiah P (2004). Optimization of process parameters for alkaline protease production under solid state fermentation by *Thermoactinomyces thalpophilus* PEE14. *J. Biotechnol.*, 5: 80–83.
- Dobrev GT, Pishtiyski IG, Stanchev VS, Mircheva R (2007). Optimization of nutrient medium containing agricultural wastes for xylanase production by *Aspergillus niger* B03 using optimal composite experimental design. *Bioresource Technol.*, 98: 2671–2678.
- Dupont L, Bouanda J, Dumonceau J, Aplincourt M (2003). Metal ions binding onto a lignocellulosic substrate extracted from wheat bran: a NICA-Donnan approach. *J. Colloid Interface Sci.*, 263: 35–41.
- Ellaiah P, Adinarayana K, Bhavani Y, Padmaja P, Srinivasulu B (2002). Optimization of process parameters for glucoamylase production under solid state fermentation by a newly isolated *Aspergillus* species. *Process Biochem.*, 38: 615–620.
- Esposito F, Arlotti G, Bonifati AM, Napolitano A, Vitale D, Fogliano V (2005). Antioxidant activity and dietary fibre in durum wheat bran by-products. *Food Res. Int.*, 38: 1167–1173.
- Falony G, Armas CJ, Mendoza D, Hernandez MLJ (2006). Production of extracellular lipase from *Aspergillus niger* by solid state fermentation. *Food Technol. Biotechnol.*, 44: 235–240.
- Farzana K, Shah SNH, Butt FB, Awan SB (2005). Biosynthesis of bacitracin in solid-state fermentation by *Bacillus licheniformis* using defatted oil seed cakes as substrate. *Pak. J. Pharm. Sci.*, 18: 55–57.
- Flight I, Clifton P (2006). Cereal grains and legumes in the prevention of coronary heart disease and stroke: a review of the literature. *Eur. J. Clin. Nutr.*, 60 (10): 1145–1159.
- Freudenheim JL, Graham S, Horvath PJ, Marshall JR, Haughey BP, Wilkinson G (1990). Risks associated with source of fiber and fiber components in cancer of the colon and rectum. *Cancer Res.*, 50: 3295–3300.
- Gessesse A, Mamo G (1999). High-level xylanase production by an alkaliophilic *Bacillus* sp. by using solid-state fermentation. *Enzyme Microb. Tech.*, 25: 68–72.
- Gomez M, Ronda F, Blanco CA, Caballero PA, Apesteguia A (2003). Effect of dietary fibre on dough rheology and bread quality. *Eur. Food Res. Technol.*, 216: 51–56.
- GORinstein S, Bartnikowska E, Kulasek G, Zemser M, Trakhtenberg S (1998). Dietary persimmon improves lipid metabolism in rats fed diets containing cholesterol. *J. Nutr.*, 128: 2023–2027.
- Gupta S, Kuhad RC, Bhushan B, Hoondal GS (2001). Improved xylanase production from a haloalkalophilic *Staphylococcus* sp. SG-13 using inexpensive agricultural residues. *World J. Microbiol. Biotechnol.*, 17: 5–8.
- Halliwell B (1996). Oxidative stress, nutrition and health. Experimental strategies for optimization of nutritional antioxidant intake in humans. *Free Radic. Res.*, 25: 57–74.
- Halliwell B, Gutteridge JM, Cross CE (1992). Free radicals, antioxidants, and human disease: where are we now? *J. Lab. Clin. Med.*, 119: 598–620.
- Hawkes F, Forsey H, Premier GC, Dinsdale RM, Hawkes DL, Guwy AJ, Maddy J, Cherryman S, Shine J, Auty D (2008). Fermentative production of hydrogen from a wheat flour industry co-product. *Bioresource Technol.*, 99: 5020–5029.
- Hemery Y, Rouau X, Lullien-Pellerin V, Barron C, Abecassis J (2007). Dry processes to develop wheat fractions and products with enhanced nutritional quality. *J. Cereal Sci.*, 46: 327–347.
- Hoseney RC (1994). Principles of cereal science and technology. 2nd ed. Am. Assoc. Cereal Chem. St. Paul, MN.
- Ikram-ul-Haq, Ashraf H, Iqbal J, Qadeer MA (2003). Production of alpha amylase by *Bacillus licheniformis* using an economical medium. *Bioresource Technol.*, 87: 57–61.
- Jacob N, Prema P (2006). Pectinases from Actinomycetes. *Food Technol. Biotechnol.*, 44: 263–267.
- Javed MR, Rashid MH, Mukhtar Z, Riaz M, Nadeem H, Huma T, Asiq N (2011). Kinetics and thermodynamics of high level β -glucosidase production by mutant derivative of *Aspergillus niger* under submerged growth conditions. *Afr. J. Microbiol. Res.*, 5(17): 2528–2538.
- Jensen MK, Koh-Banerjee P, Franz M, Sampson L, Gronbaek M, Rimm EB (2006). Whole grains, bran, and germ in relation to homocysteine and markers of glycemic control, lipids, and inflammation. *Am. J. Clin. Nutr.*, 83: 275–283.
- Jensen MK, Koh-Banerjee P, Hu FB, Franz M, Sampson L, Gronbaek M, Rimm EB (2004). Intakes of whole grains, bran, and germ and the risk of coronary heart disease in men. *Am. J. Clin. Nutr.*, 80: 1492–1499.
- Kamal-Eldin A, Lærke HN, Knudsen KE, Lampi AM, Piironen V, Adlercreutz H., Katina K, Poutanen K, Aman P (2009). Physical, microscopic and chemical characterisation of industrial rye and wheat brans from the Nordic countries. *Food Nutr. Res.*, 53: 1–11.
- Kashyap DR, Soni SK, Tewari R (2003). Enhanced production of pectinase by *Bacillus* sp. DT7 using solid state fermentation. *Bioresource Technol.*, 88: 251–254.
- Kaur P, Grewal HS, Kocher GS (2003). Production of α -amylase by *Aspergillus niger* using wheat bran in submerged and solid state fermentations. *Indian J. Microbiol.*, 43: 143–145.
- Kim K, Tsao R, Yang R, Cui SW (2006). Phenolic acid profiles and antioxidant activities of wheat bran extracts and the effect of hydrolysis conditions. *Food Chem.*, 95: 466–473.
- Kim M, Choi J, Lim SJ, Kim ST, Heo HK, Kim HJ, Jun EK, Cho WJ, Shin HY (2007). Ferulic acid supplementation prevents trimethyltin-induced cognitive deficits in mice. *Biosci. Biotechnol. Biochem.*, 71: 1063–1068.

- Kulkarni N, Shendye A, Rao M (1999). Molecular and biotechnological aspects of xylanases. *FEMS Microbiol. Lett.*, 23: 411–456.
- Landberg R, Kamal-Eldin A, Salmenkallio-Marttila M, Rousset M, Aman P (2008). Localization of alkylresorcinols in wheat, rye and barley kernels. *J. Cereal Sci.*, 48: 401-406.
- Li W, Shan F, Sun S, Corke H, Beta T (2005). Free radical scavenging properties and phenolic content of Chinese black-grained wheat. *J. Agric. Food Chem.*, 53: 8533–8536.
- Likes R, Madl RL, Zeisel SH, Craig SAS (2007). The betaine and choline content of a whole wheat flour compared to other mill streams. *J. Cereal Sci.*, 46: 93–95.
- Liu Z, Ying Y, Li F, Ma C, Xu P (2010). Butanol production by *Clostridium beijerbeckii* ATCC 55025 from wheat bran. *J. Ind. Microbiol. Biotechnol.*, 37: 495-501.
- Lupton JR, Turner ND (1999). Potential protective mechanisms of wheat bran fiber. *Am. J. Med.*, 106: 24–27.
- Maheswari MU, Chandra TS (2000). Production and potential applications of a xylanase from a new strain of *Streptomyces cupidosporus*. *World J. Microbiol. Biotechnol.*, 16: 257-263.
- Malathi S, Chakraborti R (1991). Productions of alkaline protease by a new *Aspergillus flavus* isolate under solid substrate fermentation conditions for use as a depilation agent. *Appl. Environ. Microbiol.*, 57: 712-716.
- Manikandan K, Viruthagiri T (2009). Simultaneous saccharification and fermentation of wheat bran flour into ethanol using coculture of amylolytic *Aspergillus niger* and thermotolerant *Kluveromyces marxianus*. *Front. Chem. Eng. China*, 3: 240–249.
- Minhajuddin M, Beg ZH, Iqbal J (2005). Hypolipidemic and antioxidant properties of tocotrienol rich fraction isolated from rice bran oil in experimentally induced hyperlipidemic rats. *Food Chem. Toxicol.*, 43: 747–753.
- Negi S, Banerjee R (2006). Amylase and protease production from *A. awamori*. *Food Technol. Biotechnol.*, 44: 257–261.
- Palmarola-Adrados B, Borska PC, Galbe M, Zacch G (2005). Ethanol production from non-starch carbohydrates of wheat bran. *Biores. Technol.*, 96: 843–850.
- Pam C, de Vrije T, Grabarczyk R, Urbaniec K (2006). Development of fermentation based process for biomass conversion to hydrogen gas. In Proceedings of the 9th Conference on Process Integration, Modelling and Optimisation for Energy Saving Pollut. Reduction, 4: 1130.
- Pan C, Fan Y, Hou H (2008). Fermentative Production of hydrogen from wheat bran by mixed anaerobic cultures. *Ind. Eng. Chem. Res.*, 47: 5812–5818.
- Pandey A (1992). Recent process developments in solid-state fermentation. *Process Biochem.*, 27: 109–117.
- Pandey A, Selvakumar P, Soccol CR, Nigam P (1999). Solid-state fermentation for the production of industrial enzymes. *Curr. Sci.*, 77: 149–162.
- Pereira MA, Jacobs DRJ, Pins JJ, Raatz SK, Gros MD, Slavin JL, Seaquist ER (2002). Effect of whole grains on insulin sensitivity in overweight hyperinsulinemic adults. *Am. J. Clin. Nutr.* 75: 848–855.
- Pomeranz Y (1988). Wheat, Chemistry and Technology. Am. Assoc. Cereal Chem. St. Paul, MN.
- Qu H, Madl RL, Takemoto DJ, Baybutt RC, Wang W (2005). Lignans are involved in the antitumor activity of wheat bran in colon cancer SW480 cells. *J. Nutr.*, 135: 598–602.
- Reddy BS, Hirose Y, Cohen LA, Simi B, Cooma I Rao, C.V (2000). Preventive potential of wheat bran fractions against experimental colon carcinogenesis: Implications for human colon cancer prevention. *Cancer Res.*, 60: 4792–4797.
- Rhodes DL, Stone BA (2002). Proteins in walls of wheat aleurone cells. *J. Cereal Sci.*, 36: 83–101.
- Sabovic M, Lavre S, Keber I (2004). Supplementation of wheat fibre can improve risk profile in patients with dysmetabolic cardiovascular syndrome. *Eur. J. Cardiovasc. Prev. Rehabil.*, 11: 144–148.
- Safdar MN (2005). Composition of wheat grain. *Wheat Milling News*.
- Safdar MN, Naseem K, Amjad M, Mumtaz A and Raza S (2009). Physicochemical quality assessment of wheat grown in different regions of Punjab. *Pak. J. Agri. Res.*, 22: 1-2.
- Saima, Akhtar M, Hashmi AS (1999). Bioconversion of wheat bran to biomass protein through metabiosis and its biological evolution in broiler chicks. *Int. J. Agri. Biol.*, 1: 39-41.
- Salmenkallio-Marttila M, Katina K, Autio K (2001). Effects of bran fermentation on quality and microstructure of high-fiber wheat bread. *Cereal Chem.*, 78: 429–435.
- Sandhya C, Sumantha A, Szakacs G, Pandey A (2005). Comparative evaluation of neutral protease production by *Aspergillus oryzae* in submerged and solid-state fermentation. *Process Biochem.*, 40: 2689-2694.
- Sang S, Ju J, Lambert JD, Lin Y, Hong J, Bose M, Wang S, Bai N, He K, Reddy BS, Ho CT, Li F, Yang CS (2006). Wheat bran oil and its fractions inhibit human colon cancer cell growth and intestinal tumorigenesis in Apc(min/+) mice. *J. Agric. Food Chem.*, 54: 9792-9797.
- Satue-Gracia MT, Heinonen M, Frankel EN (1997). Anthocyanins as antioxidants on human low-density lipoprotein and lecithin-liposome systems. *J. Agric. Food Chem.*, 45: 3362–3367.
- Sekar C, Ragasekar VW, Balaraman K (1997). Production of Cyclosporin A by solid state fermentation. *Bioproc. Biosyst. Eng.*, 17: 257-259.
- Sekhon A, Dahiya N, Tewari RP, Hoondal GS (2004). Production of lipase from *Bacillus megaterium* AKG-1 using wheat bran in soild substrate fermentation. *Indian J. Microbiol.*, 44: 219-220.
- Shafee S, Topal E (2009). When will fossil fuel reserves be diminished? *Energ. Policy*. 37: 181-189.
- Shamala TR, Sreekanthia KR (1988). Use of wheat bran as a nutritive supplement for the production of ethanol by *Zymomonas mobilis*. *J. Appl. Bacteriol.*, 65: 433-436.
- Silva D, Tokuoshi K, da Silva Martins E, Da Silva R, Gomes E (2005). Production of pectinase by solid-state fermentation with *Penicillium viride* RFC3. *Process Biochem.*, 40: 2885-2889.
- Sindhu R, Suprabha GN, Shashidhar S (2009). Optimization of process parameters for the production of alpha amylase from *Penicillium janthinellum* (NCIM 4960) under solid state fermentation. *Afr. J. Microbiol. Res.*, 3(9):498-503.
- Slavin J (2003). Why whole grains are protective: biological mechanisms. *Proc. Nutr. Soc.*, 62: 129–134.
- Slavin JL, Martini MC, Jacobs DR. and Marquar L (1999). Plausible mechanisms for the protectiveness of whole grains. *Am. J. Clin. Nutr.*, 70: 459S–463S.
- Soarse FEF, Braga FR, Genier HLA, Araujo JV, Ferriera SR, Aaujo JM, Tavela A de O, Vilela VLR, Queiróz JH (2010). Optimization of medium composition for protease production by *Paecilomyces marquandii* in solid-state-fermentation using response surface methodology. *Afr. J. Microbiol. Res.*, 4(24): 2699-2703.
- Subramany S, Prema P (2002). Biotechnology of microbial xylanases: Enzymology, molecular biology and application. *Crit. Rev. Biotechnol.*, 22: 33–64.
- Sukumaran RK, Singhania RR, Mathew GM, Pandey A (2009). Cellulase production using biomass feed stock and its application in lignocellulose saccharification for bio-ethanol production. *Renew. Energ.*, 34: 421–424.
- Sumantha A, Larroche C, Pandey A (2006). Microbiology and industrial biotechnology of food-grade proteases. *Food Technol. Biotechnol.*, 44: 211–220.
- Sumantha A, Sandhya C, Szakacs G, Soccol CR, Pandey A (2005). Production and partial purification of a neutral metalloprotease by fungal mixed substrate fermentation. *Food Technol. Biotechnol.*, 43: 313–331.
- Surget A, Barron C (2005). Histologie du grain du ble. *Industrie des céréales*. 145: 3-7.
- Suzuki A, Yamamoto M, Jokura H, Fujii A, Tokimitsu I, Hase T, Saito I (2007). Ferulic acid restores endothelium-dependent vasodilation in aortas of spontaneously hypertensive rats. *Am. J. Hypertens.*, 20: 508–513.
- Tahir MI, Khalique A, Pasha TN, Bhatti JA (2002).Comparative evaluation of maize bran, wheat bran and rice bran on milk production of holstein friesian cattle. *Int. J. Agr. Biol.* 4: 559–560.
- Topping D (2007). Cereal complex carbohydrates and their contribution to human health. *J. Cereal Sci.*, 46: 220–229
- Tsujita Y, Endo A (1976). Purification and characterization of the two molecular forms of *Aspergillus oryzae* acid protease. *Biochim. Biophys. Acta.*, 445: 194–204.

- Vintila T, Dragomirescu M, Jurcoane S, Caprita R, Maniu M (2009). Production of cellulase by submerge and solid state cultures and yeasts selection for conversion of lignocellulose to ethanol. *Rom. Biotechnol. Lett.*, 14: 4275-4281.
- Vishwanatha KS, Appu Rao AG, Singh SA (2009). Acid protease production by solid-state fermentation using *Aspergillus oryzae* MTCC 5341: optimization of process parameters. *J. Ind. Microbiol. Biotechnol.*, 85: 1849-1859.
- Waggle DH, Lambert MA, Miller GD, Farrel EP, Deyoe CW (1967). Extensive analyses of flours and millfeeds made from nine different wheat mixes. II. Amino acids, minerals, vitamins, and gross energy. *Cereal Chem.*, 44: 48-60.
- WASDE (2010). World Agricultural Supply and Demand Estimates, World Agricultural outlook board: June, 2010.
- Whelton PK, He J, Cutler JA, Brancati FL, Appel L J, Follmann D and Klag MJ (1997). Effects of oral potassium on blood pressure: metaanalysis of randomized controlled clinical trials. *J. Am. Med. Assoc.*, 277: 1624-1632.
- Willcox J K, Ash SL, Catignani GL (2004). Antioxidants and prevention of chronic disease. *Crit. Rev. Food Sci. Nutr.*, 44: 275-295.
- Xu YX, Li Y L, Xu S C, Liu Y, Wang X, Tang JW (2008). Improvement of xylanase production by *Aspergillus niger* XY-1 using response surface methodology for optimizing the medium composition. *J. Zhejiang Univ. Sci. B.*, 9: 558-566.
- Zambare V (2010). Solid State Fermentation of *Aspergillus oryzae* for glucoamylase production on Agro residues. *Int. J. Life Sci.*, 4: 16-25.
- Zhou K, Lan S, Yu L (2004a.) Phytochemicals and antioxidant properties of wheat bran. *J. Agric. Food Chem.*, 52: 6108-6114.
- Zhou K, Laux JJ, Yu L (2004b). Comparison of Swiss red wheat grain and fractions for their antioxidant properties. *J. Agric. Food Chem.*, 52: 1118-1123.
- Zhou K, Yu L (2004). Antioxidant properties of bran extracts from Trego wheat grown at different locations. *J. Agric. Food Chem.*, 52: 1112-1117.