

Short Communication

First report on the nematode-trapping fungus, *Arthrobotrys javanica*, from the Soil of Ulleung Island, Korea

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***Arthrobotrys javanica* captures nematodes in adhesive networks. This fungus was isolated from the soil around the root of *Dystaenia takeshimana* in Ulleung Island, South Korea. It produces 1-septate conidia with a size of 31.1 × 12.0 μm on a candelabrum-like apical branch. This feature distinguishes *A. javanica* from other species of *Arthrobotrys*. Chlamydospores were observed (d = 17 μm), which were not previously mentioned in the original description. The present study is the first report on *A. javanica* in Korea.**

Key words: *Arthrobotrys javanica*, nematode-trapping fungus, adhesive network.

INTRODUCTION

Nematode-trapping fungi are ubiquitous in soil environments. They have been studied worldwide because of their potential applications as bio-control agents for plant and animal parasitic nematodes (Mankau, 1980; Bordallo et al., 2002; Soder et al., 2005). These fungi capture nematodes by three-dimensional networks, adhesive columnar branches, adhesive knobs, constricting rings, and non-constricting rings. They digest the internal organs of nematodes and form reproductive cells. Few studies on nematode-trapping fungi are available in South Korea.

The main objective of this study was to describe morphological characters of a potential fungus, which was collected during survey of bio-control fungi for plant-parasitic nematodes in Korea in 2009.

MATERIALS AND METHODS

Fresh soil around the root of *D. takeshimana* was collected in Ulleung Island, Korea on August 2009. The nematode-trapping fungi were isolated through a modified sprinkling-baiting technique (Barron, 1977). Subsamples of 1.0 g soil were sprinkled onto plates of both 1.7% corn meal agar (CMA, Difco) and 2% water agar (WA). About 200 nematodes (*Rhabditis* spp.) were added to the surface of Petri dish plates (D = 10 cm) as bait for the nematode-trapping fungi. Two CMA and two WA plates were used for each soil sample. The plates were incubated for two to four weeks at 25 °C and examined every other day under a dissecting microscope (Olympus SZ×12) to detect the appearance of nematode-trapping fungi. All detected nematode-trapping fungi were photographed using an attached digital camera (Nikon DXM1200F), and transferred to CMA for pure culture. A new recorded species of nematode-trapping fungus was isolated. The description of this species is presented in the current paper.

RESULTS AND DISCUSSION

The taxonomic revisions of this species are given below:

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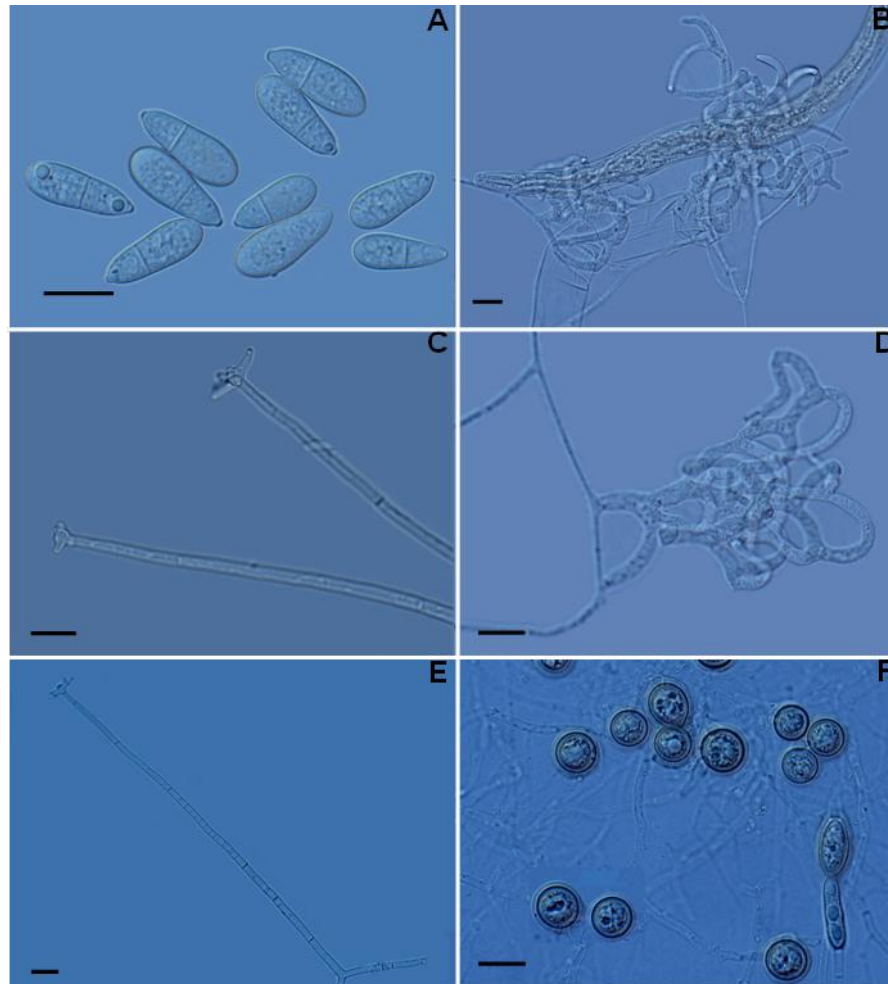


Figure 1. *Arthrobotrys javanica*. **A**, conidia; **B**, partially digested nematode captured by adhesive network; **C** and **E**, conidiophores; **D**, adhesive network; **F**, chlamydospores. Bars = 20 μm .

Arthrobotrys javanica

1. *Candelabrella javanica* (Rifai and Cooke, 1966).
2. *Arthrobotrys javanica* (Jarowaja, 1970).

The mycelium was hyaline, septate, branched, smooth walled, and 3 to 7 μm in diameter. The conidiophores were erect, straight, subulate, 275 to 440 μm high, approximately 9 μm wide at the base, and gently attenuating to 2 to 3 μm near the apex. They were hyaline, smooth and slightly enlarged as ramification occurred, and bearing a lax, short, candelabrum-like apical branching system. The conidia arose singly as blown-out ends of the conidiophore and the subsequently developed growing points. They were narrowly obovoid, 1-septate, 25.0 to 42.5 μm long, 10.0 to 15.6 μm wide, slightly constricted at the septum, rounded distally with the obconical basal cell tapering to a truncate base, smooth walled and hyaline. In pure culture on CMA, the conidia were smaller, up to 35 μm long and 11.3 to 13.8 μm wide. The detached

conidia left behind subcylindrical conidial pegs up to 15 μm long and approximately 2 μm in diameter. The fungus trapped nematodes by means of adhesive networks.

This species was originally described by Rifai and Cooke (1966) from the soil in Bogor Botanic Garden, Java, on sterile rabbit dung agar. The fungus in this study identified as *A. javanica* was first isolated from the soil around *Dystaenia takeshimana* in Ulleung Island, Korea. The fungus produces narrowly obovoid large conidia on the surface of CMA. The conidiophores characteristics (Figures 1A and B), conidia shape (Figure 1C), and three-dimensional adhesive networks (Figures 1D and E) well fitted with Rifai and Cooke's description of *A. javanica*. Slight differences were observed on the conidia and conidiophores but within the range of *A. javanica* Jarowaja originally described by Rifai and Cooke (1966).

Chlamydospores were not mentioned in the original description. In the present study, the chlamydospores were observed at 50 days after incubation at 25°C on

Table 1. Comparison of *Arthrobotrys javanica* isolates from Korea with the original description and its closely related species *Arthrobotrys musiformis* (Rifai and Cooke, 1966).

Character	Korea isolate		Original description	
		(Range)	<i>A. javanica</i> (Range)	<i>A. musiformis</i> (Range)
Conidiophores	Length (µm)	359.6 (207.4–528.2)	275.0–440.0	Up to 600
Conidia	Length (µm)	31.1 (24.9–38.3)	25.0–42.5	29.0–45.0
	Width (µm)	12.0 (10.5–15.5)	10.0–15.6	8.0–13.5
	Shape	Narrowly obovoid	Narrowly obovoid	Ellipsoid or narrowly obovoid, generally curved
	Septa	1, slightly constricted	1, slightly constricted	1, non-constructed
Chlamydospores		Thick walled, smooth, globose, or elongate ovoid	None reported	Thick-walled, tuberculated, and globose

CMA (Figures 1F). Generally, thick-walled, smooth, yellow, and 17.0 µm in diameter globose chlamydospores are produced terminally on short lateral hypha, along with less frequent elongate ovoid chlamydospores (27.6 × 19.0 µm). Among the species in the genus *Arthrobotrys*, only *A. javanica* and *Arthrobotrys musiformis* produced conidia on candelabrum-like apical branches (Figures 1A and B), with adhesive networks. However, *A. javanica* is different from *A. musiformis*. The conidium of *A. javanica* is shorter, wider, not curved, and slightly constricted at the septum (Table 1) (Rifai and Cooke, 1966).

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