

## Full Length Research Paper

# Screening of *Lactobacillus* spp. from raw goat milk showing probiotic activities against pathogenic bacteria

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This research work was executed to isolate potential *Lactobacillus* spp. from local raw goat milk samples for their antimicrobial activities against different human intestinal pathogens. Isolation of such bacteria was carried out by Man Rogosa Sharpe (MRS) agar media. Identification of the isolated strain was conducted according to the morphological and biochemical tests. Furthermore, growth optimization and their antimicrobial activity also studied against the pathogenic bacteria (*Escherichia coli* ATCC 25921, *Pseudomonas aeruginosa* ATCC 9027, *Vibrio cholera* ATCC 14035 and *Salmonella enteric* ATCC 14028). A total of 23 isolates were obtained from the raw goat milk samples, among them four isolates were selected on the basis of their antimicrobial activities against pathogenic bacteria. The *Lactobacillus* spp. showed better growth in 6.5 to 7.5 pH range and temperature range was 35 to 40°C. Thus it can be said that the goat milk provides a natural dwelling place for *Lactobacillus* spp., which are beneficial to human health.

**Key words:** Antimicrobial activity, growth optimization, intestinal pathogens, *Lactobacillus* spp., Man Rogosa Sharpe (MRS) agar media, pH, raw goat milk.

## INTRODUCTION

*Lactobacillus* is one of the most important genera of lactic acid bacteria (LAB) (Coeuret et al., 2003). They are considered as generally recognized as safe (GRAS) organisms and can be safely used as probiotics for medical and veterinary purposes (Fuller, 1989). Probiotics, as defined in the FAO/WHO (2002) report, are

“live microorganisms which when administered in adequate amounts confer a health benefit on the host”. Probiotics are beneficial bacteria in that they favorably alter the intestinal microflora balance, inhibit the growth of harmful bacteria, promote good digestion, boost immune function and increase resistance to infection (Helland et

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al., 2004). Other physiological benefits of probiotics include removal of carcinogens, lowering of cholesterol, immunostimulating and allergy lowering effect, synthesis and enhancing the bioavailability of nutrients, alleviation of lactose intolerance (Parvez et al., 2006).

Study on lactic acid bacteria specially *Lactobacillus* spp. are drawing a worldwide interests significantly in worldwide due to its applied beneficiary effects in the prevention, control and treatment of diseases and health maintenance (Osuntoki et al., 2008). Now, *Lactobacillus* spp. are widely used as probiotics which play a key role in enhancing resistance to colonization by exogenous, potentially, pathogenic organisms in the intestinal tract. They produce a variety of substances e.g., bacteriocin, nisin, lactacin etc. that are effective against different types of enteric pathogens like *Escherichia coli*, *Salmonella* spp., *Shigella* spp., *Vibrio* spp., *Bacillus* spp., *Klebsiella* spp., *Staphylococcus* spp., *Pseudomonas* spp., *Proteus* spp. etc. These effects can be described as the improvement of lactose digestion and the treatment of diarrheal disorders (Abdullah and Osman, 2010).

*Lactobacillus* spp. is widely distributed in nature and found as indigenous microflora in raw milk (Rodriguez et al., 2000) and fermented milk with spontaneous fermentation. LAB mostly found in milk of human (Martin et al., 2003) and other animals (Fujisawa and Mitsuoka, 1996). The present study was designed to characterize some *Lactobacillus* spp. from various goat milk samples collected from different regions of Chittagong district to investigate their optimum growth conditions, and some probiotic properties like sensitivity to antibiotics and antagonistic activity to pathogenic bacteria.

## MATERIALS AND METHODS

### Sample collection

Five different raw goat milk samples were collected from different local area of Noakhali zone under the Chittagong division in Bangladesh in sterile bottles and packed in sterile polythene bags. After packing the bags, the milk samples were carried to microbiology laboratory of globe pharmaceuticals LTD, Bangladesh in an ice box contained ice which maintained the temperature at 4°C within four hours, and used for further studies (isolation, identification and antimicrobial activity). The remaining milk samples were stored at 0°C for further use.

### Isolation of *Lactobacillus* spp. from goat milk

Samples were cultured by pour plat method on the Man Rogosa Sharpe (MRS) agar media after serial dilution and incubated at 37°C for 24 to 48 h. The selected strains on the bases of their activity in MRS media were then subcultured onto MRS agar slants which were incubated at 37°C for 24 h and preserved in 20% glycerol (Oxoid, Canada) at -20°C until further used.

### Identification of isolated *Lactobacillus* spp.

The selected isolates were examined for their morphological

properties, such as size, shape, cell arrangement and gram-staining properties. Cultural properties including form, colour, elevation, margin, surface of colonies on MRS agar plate and slant were also recorded. Physiological and biochemical characteristics of the isolates were evaluated by Voges-proskauer, methyl red, indole, and citrate utilization (IMViC) catalase and oxidase tests. The ability of the isolates to ferment a number of sugars including glucose, xylose, arabinose, lactose, glycerol, starch, and manitol were also tested. The isolates were identified up to species based on comparative analysis of the observed characteristics with the standard description of bacterial strains in Bergey's Manual of Determinative Bacteriology.

### Optimization of growth parameters

Optimum temperature and pH were determined. Optimum growth temperature was determined by growing the selected isolates in MRS broth and incubating at different temperatures (20 to 45°C) for 24 h. MRS broth of different pH (pH 4.5 to 9) were inoculated and incubated for 24 h to determine the optimum pH. The optimum parameters for highest growth of the identified *Lactobacillus* spp. were determined by measuring and comparing the optical density (OD) at 600 nm (OD600) (Barua et al., 2015).

### Anti-microbial activity of isolated *Lactobacillus* spp.

Cross-streak method was used to detect the anti-microbial activity of *Lactobacillus* spp. (Lertcanawanichakul and Sawangnop, 2008). against selected gram negative pathogenic bacteria such as *Salmonella* spp., *Vibrio cholerae*, *Pseudomonas aeruginosa* and Enterotoxigenic *E. coli* (ETEC). Each pure isolated *Lactobacillus* spp. culture from previous MRS agar slant was individually streaked in half portion of different MRS agar plates with a smear line. Then, all plates were incubated at 37°C for 3 days. After 2 days incubation period, the isolates secreted anti-metabolites into the medium and then, test pathogens were cross-streaked along the line of fully grown isolates following Cross-streak method. Each streaking was started from near the edge of the plates and was streaked toward the growth line of the isolated *Lactobacillus* spp. After streaking into the medium, the plates were incubated at 37°C for 24 h. After overnight incubation, a clear zone of inhibition along the growth line of the isolates were observed which indicated growth inhibition of pathogenic test bacteria due to anti-microbial activity of isolated *Lactobacillus* spp. The microbial interactions were analyzed by the observation of the size of the inhibition zone (Madigan et al., 1997).

### Assay of antibiotic sensitivity pattern

To assess the antibiotic sensitivity pattern, disk diffusion method was followed (Ivanova et al., 2000). Culture inoculums of the isolates grown in MRS broth was taken as amount of 0.5 ml, and was mixed in 5 ml of the same medium containing 0.5% agar, and aseptically poured into glass Petri dishes containing MRS agar medium. The antibiotic disks (Ampicillin, Amoxicilline, Tetracycline, Erythromycin and Gentamicin) were placed on the surface of the plate at equidistance. The plates were then kept at 4°C for 1 h for proper diffusion of antibiotics. The plates were incubated at 37°C for 24 h. The antibiotic sensitivity or resistance was determined by observing the presence of zone of inhibition. The zone of inhibition was measured by a millimeter scale.

## RESULTS

### Isolation

After primary screening of five goat milk samples by MRS

**Table 1.** Microscopic and biochemical tests of isolates with carbohydrate utilization ability.

Tests	LB <sub>a</sub>	LB <sub>b</sub>	LB <sub>c</sub>	LB <sub>d</sub>
Gram Staining	+	+	+	+
Form	Rod	Rod	Rod	Rod
Catalase	-	-	-	-
Oxidase	-	-	-	-
<b>IMViC</b>				
Indole	-	-	-	-
MR	-	-	-	-
VP	-	-	-	-
Citrate	-	-	-	-
<b>Carbohydrate fermentation</b>				
D (+) Xylose	+	+	+	+
Lactose	+	+	+	+
Glycerol	+	+	+	+
Starch	+	+	+	+
Manitol	+	+	+	+
Glucose	+	+	+	+

+, Positive result; -, Negative result.

agar media, 23 isolates were found, all the isolates produced small, irregular and round shape with shiny whitish cream or brownish colored which were morphologically similar to *Lactobacillus* spp. and among these isolets only 4 isolates were selected for further study according to their growth inhibition properties against four references pathogenic microorganisms *Salmonella* spp., *Vibrio cholerae*, *Pseudomonas aeruginosa* and Enterotoxigenic *E. coli* (ETEC).

### Identification

All isolates were examined under microscope to observe their microscopic properties. These isolates were found gram positive, short and medium rod shaped non-spore forming bacteria. Furthermore, some biochemical tests such as catalase test, oxidase test, IMViC tests (Indole test, Methyl Red (MR) test, Voges Proskauer (VP) test, citrate utilization test) and carbohydrate fermentation patterns were performed as depicted by Bergey's manual systematic bacteriology (Hensyl, 1994). The isolates were found catalase and oxidase negative and in indole, methyl-red, voges proskauar, citrate utilization (IMViC) tests all isolates were also found negative. In this study, all the four isolates were able to ferment 6 different carbohydrates, that is, Xylose, Lactose, Glycerol, Starch, Mannitol and Glucose indicating that they are able to grow in variety of habitats utilizing different type of carbohydrates. The summarized results of all bacteriological and biochemical tests are presented in Table 1.

### Optimum growth parameters for the selected *Lactobacillus* spp.

Growth of selected isolates (four isolates) studied on different growth parameters like pH and temperature to reveal their optimum growth parameter. Most of the isolates showed the best growth at neutral pH and the less growth observed when pH increased and decreased. Furthermore, it also found that the best bacterial growing temperature was between 35 to 40°C. Cellular growth was indicated by turbidity after incubated for 24 h on broth media. The results of morphological and physiological tests over four isolates were shown in the Table 2.

### Assay of anti-bacterial activity by cross-streak method

Zone of inhibition produced by the isolates against target pathogens were indicated their antimicrobial properties and identified isolates of *Lactobacillus* spp. showed that inhibitory properties against *Salmonella* spp., *Vibrio cholerae*, *Pseudomonas aeruginosa* and Enterotoxigenic *E. coli* (ETEC) (Table 3).

### Susceptibility and resistance to antibiotics

Four potential isolates (LB<sub>a</sub>, LB<sub>b</sub>, LB<sub>c</sub> and LB<sub>d</sub>) were subjected to the antibiotic resistance study, where we use five antibiotics (Ampicillin, Amoxicilline, Tetracycline, Erythromycin and Gentamicin). The result showed that all

**Table 2.** Growth ability of selected isolates in different pH and temperature.

Growth in different pH range					
4.5	+++	+++	+++	+++	+++
5.0	+++	+++	+++	+++	+++
5.5	+++	+++	+++	+++	+++
6.0	+++	+++	+++	+++	+++
6.5	+++	+++	+++	+++	+++
7.0	+++	+++	+++	+++	+++
7.5	+++	+++	+++	+++	+++
8.0	+++	+++	+++	+++	+++
8.5	+++	+++	+++	+++	+++
9.0	+++	+++	+++	+++	+++
Growth in different temperature (in °C) range					
20	+++	+++	+++	+++	+++
25	+++	+++	+++	+++	+++
30	+++	+++	+++	+++	+++
35	+++	+++	+++	+++	+++
40	+++	+++	+++	+++	+++
45	+++	+++	+++	+++	+++

+++ , Optimum growth; +---, poor growth.

**Table 3.** Antimicrobial activity test of *Lactobacillus* spp. against pathogenic bacteria.

Isolate	Zone of inhibition (in mm)			
	<i>Salmonella</i> spp.	<i>V. cholerae</i>	<i>P. aeruginosa</i>	<i>E. coli</i>
LB <sub>a</sub>	6.5	4.5	5.5	7.0
LB <sub>b</sub>	7.0	6.0	6.0	7.6
LB <sub>c</sub>	8.0	5.5	6.4	5.4
LB <sub>d</sub>	7.7	6.8	8.0	6.0

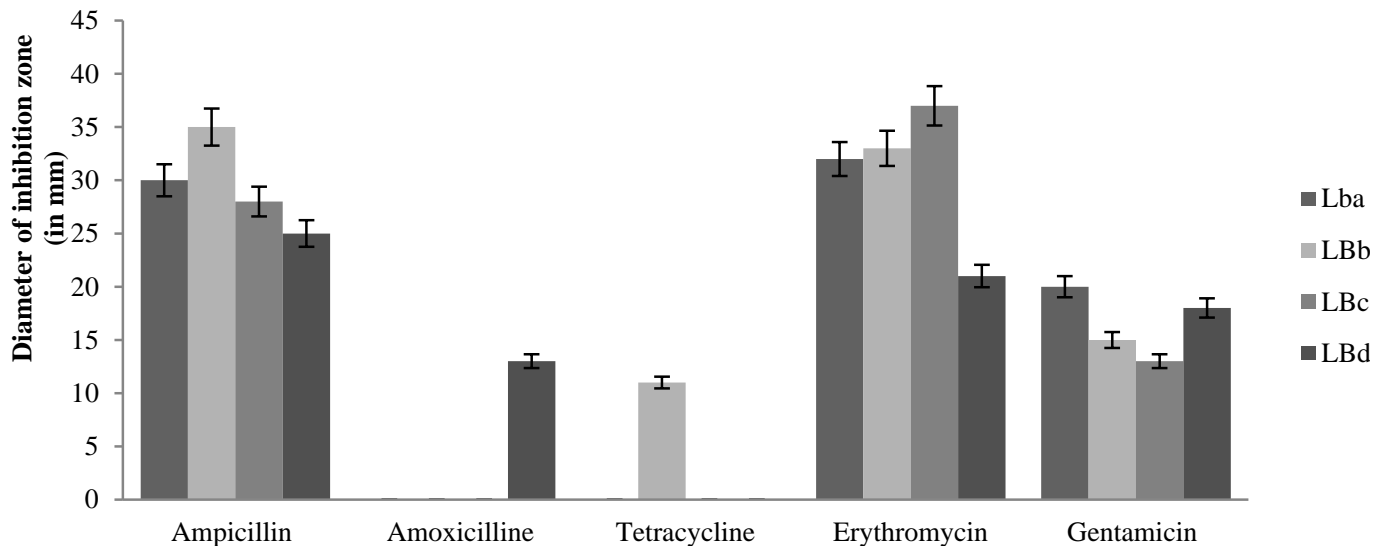
isolates were susceptible to Erythromycin, Gentamicin and Ampicillin. Whereas most of these isolates were resistant against remaining antibiotics (Figure 1).

## DISCUSSION

Milk and milk products are usually associated with probiotics bacteria, which provide supplements in maintaining beneficial intestinal balance (Tambekar and Bhutada, 2010). This study was designed to isolate and identify *Lactobacillus* spp. using MRS media and from bacterial morphological and biochemical characteristics. Moreover to determine probiotic activities of the isolates against various pathogens like *Salmonella* spp., *Vibrio cholerae*, *Pseudomonas aeruginosa* and Enterotoxigenic *E. coli* (ETEC). Man Rogosa Sharpe (MRS) agar media is most commonly used for cultivation of *Lactobacillus* spp. (El-Moez et al., 2001). This is why, after isolation of 23 *Lactobacillus* spp. from five raw goat milk samples using

MRS media, only four isolates showed better activities against pathogenic bacteria mentioned above. The *Lactobacillus* spp. can produce some metabolites such as Bacteriocins like Acidophilin, Acidolin, Lactocidin, Bulgarican, Lactolin, Lactobacillin and Lactobrevin which are antagonistic to various degrees against diarrheagenic intestinal pathogens (Tambekar and Bhutada, 2010). Four potential isolates (out of 23) were characterized during the isolation steps, according to their colony morphology (color, size, shape, appearance, gram staining) on MRS media, which showed better antibacterial activity against four reference strains. All the potential isolates showed moderate or strong inhibitory activity and zone of inhibition was measured. But the entire growth inhibition (in ml) was not same. Some isolates showed positive result against all reference strains but some isolates could not.

Selected isolates (LB<sub>a</sub>, LB<sub>b</sub>, LB<sub>c</sub> and LB<sub>d</sub>) found to be gram positive, short and medium rod shaped non-spore forming bacteria which indicated they are members of



**Figure 1.** Susceptibility and resistance of four isolates (LB<sub>a</sub>, LB<sub>b</sub>, LB<sub>c</sub> and LB<sub>d</sub>) of *Lactobacillus* spp. against antibiotics (Ampicillin, Amoxicillin, Tetracycline, Erythromycin and Gentamicin).

*Lactobacillus* spp. (Thamaraj and Shah, 2003). The isolates were showed catalase and oxidase negative and in IMViC tests all isolates were also found negative, thereby these might confirm the isolates were *Lactobacillus* spp. (Dhanasekaran et al., 2010). All these results are found relevant to the findings of Chowdhury et al. (2012).

These four isolates were also subjected to examine their optimum growth condition by allowing them to grow in different pH and temperature for 24 h in broth medium. They showed better result in 6.5 to 7.5 pH range and temperature range was 35 to 40°C. Findings of present study showed more or less similarities with the previous study where optimum pH and temperature for the growth of *Lactobacillus* spp. was neutral (7) and 37°C respectively (Barua et al., 2015). Antibiotic susceptibility and resistance test also observed of these four bacterial isolates, the result showed that all isolates are susceptible to Erythromycin, Gentamicin and Ampicillin. But most of these strains were resistant against remaining antibiotics. Study of Barua et al. (2015) and Anas et al., (2014) also support the present research.

It is clear that these isolates may produce some metabolites that play role on growth inhibition of pathogens. So, isolated *Lactobacillus* spp. might have a further research value.

## CONFLICT OF INTERESTS

The authors have not declared any conflict of interests.

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