Full Length Research Paper

# Anti-diabetic effects of walnut oil on alloxan-induced diabetic rats

Parivash Rahimi<sup>1</sup>, Najmeh Kabiri<sup>1</sup>\*, Sedigheh Asgary<sup>2</sup> and Mahbubeh Setorki<sup>3</sup>

<sup>1</sup>Department of Biology, Faculty of Sciences, Isfahan University, Isfahan, Iran.

<sup>2</sup>Isfahan Cardiovascular Research Center, Applied Physiology Research Center, Isfahan University of Medical Sciences, Isfahan, Iran,

<sup>3</sup>Department of Biology, Izeh Branch, Islamic Azad University, Izeh, Iran.

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Diabetes arises from a deficient production of insulin by the  $\beta$ -cells of the pancreatic islets. Many traditional treatments have been recommended in the alternative system of medicine for treatment of diabetes mellitus; however, the mechanism of most of the herbs used has not been defined. This study was carried out to clarify the effect of walnut (*Juglans regia*) oil on blood glucose and their possible effect on pancreatic tissue. Diabetes mellitus was induced in 20 adult male rats, using intraperitoneal injection of 75 mg/kg body weight (BW) alloxan. The diabetic rats were divided into two groups; the first group (positive control) received an ordinary diet, and the second group, diabetic rats treated with walnut oil. The non-diabetic rats (negative control group) received neither alloxan nor the oil. Following consumption of oil, blood glucose was measured and on the last day the pancreas were removed and stained with hematoxylin and eosin (H&E), and morphology of the pancreatic sections was studied. The results of this study indicate that walnut oil was able to reduce blood glucose significantly when compared with the control group (P < 0.05). In the control, positive group average area of islets in pancreatic sections was significantly reduced in comparison with the control negative (normal health) and diabetic treated with walnut oil (P < 0.05). These results suggest the validity of the clinical use of walnut oil in the treatment of diabetes mellitus type I.

Key words: Walnut, alloxan, diabetes mellitus type I, rat.

# INTRODUCTION

Diabetes and its complications are major causes of morbidity and mortality in the U.S. and contribute substantially to health care costs (Centers for Disease Control and Prevention). These data have shown that older age, family history and obesity, are strong and consistent determinants of type 2 diabetes (Knowler et al., 1981; King et al., 1984; Haffner et al., 1991). Physiological data show that peripheral insulin resistance and pancreatic  $\beta$ -cell dysfunction are precursors and key determinants of type 2 diabetes (Lillioja et al., 1993). The factors involved to the development of diabetes is changes in sex steroid hormones. However, the mechanisms responsible for these associations as well as the link to

sexual dysfunction are not well understood. Imbalances in sex steroid hormone levels are strongly associated with diabetes and this may negatively impact upon sexual function (Kim, 2009). Diabetes mellitus is a group of metabolic disorders characterized by elevation of blood glucose concentration and is associated with increased prevalence of microvascular complications. Type 1 diabetes mellitus results from cellular mediated autoimmune destruction of pancreatic  $\beta$ -cells of islets of Langerhans and results in loss of insulin production (Shivananda and Geetha, 2005). Therefore, damage to pancreatic b cells due to the release of tumor necrosis factor-a (TNF-a) and interleukin-1 (IL-1) produced by infiltrating macrophages, lymphocytes and monocytes leads to the development of type 1 diabetes mellitus (DM) (Nielsen, 1986; Dunger et al., 1996). The positive effects of camel milk on collagen could be explained by its effects on glucose levels and

<sup>\*</sup>Corresponding author. E-mail: kabiri\_s97@yahoo.com.

insulin action. Camel milk has been shown to normalize insulin action and improve insulin sensitivity (Agrawal et al., 2004) in diabetic rats. The hypoglycemic activity of camel milk may be because of high concentrations of insulin like protein in camel milk which contains about 45 to 128 units/L (Singh, 2001) and also it contains high amount of zinc (Mohamed et al., 1995). Zinc is playing a major role for insulin secretory activity in pancreatic beta cells. in streptozotocin-diabetic rats (Al-Numair et al., 2011). Experiments on rats with streptozotocin-induced diabetes, provided evidence that resveratrol (3,5,4-trihydroxystilbene)-a naturally occurring phytoalexin present in numerous plant species exerts beneficial effects in the organism and may be helpful in preventing and treating some metabolic diseases, including diabetes (Fr ojd o et al., 2008; Szkudelska and Szkudelski, 2010) and induced to decrease blood glucose (Szkudelski and Szkudelska, 2011). Kumar et al. (2011) reported after 60 days of feeding, fish oil diet group had a 17.7% decrease in fasting blood glucose, suggesting overall protection of fish oil from hyperglycemia (Kumar et al., 2011). Flachs et al. (2006) reported a 4% decrease in blood glucose in mice fed with fish oil diet. Walnuts, the seeds of Juglans regia L. (Juglandaceae), are a highly nutritious food (Martínez et al., 2010). They are also used as a traditional therapy for treating cough, stomach ache (Perry, 1980) and cancer in Asia and Europe (Duke, 1989). Walnut fat is mostly unsaturated fatty acids, such as linoleic and oleic acid, which may give walnuts additional antiatherogenic properties (Fukuda et al., 2003; Feldman, 2002). Walnuts are highly enriched in omega-6 and omega-3 polyunsaturated fatty acids (PUFA), which are essential dietary fatty acids. Epidemiological and clinical trials suggest that omega-3 PUFA might have a significant role in the prevention of coronary heart disease (CHD) (Harper and Jacobson, 2001; Bucher et al., 2002). Walnut consumption improves endothelial function in type 2 diabetic individuals and may therefore help reduce cardiovascular disease risk in this high-risk population (May et al., 2010). Glibenclamide is an hypoglycemic agent which is also used in researches as a standard medication in the treatment of diabetes (Dhandapani, 2002). At the end, treatment with glibenclamide and walnut oil will be compared.

This study assessed the effect of walnut oil, on blood glucose and some other biochemical parameters in alloxan-induced diabetic rats.

## MATERIALS AND METHODS

#### Preparation of walnut oil

Walnuts used in this experiment were collected in June, 2005 from gardens in Baghbadoran, Isfahan. They were identified and a voucher specimen was deposited at the Herbarium of the Department of Biology, Faculty of Science, Isfahan University (voucher no. 2041), and its oil fraction was extracted with hexane.

#### Animal treatments and induction of diabetes

Twenty four male Wistar rats, 4 to 5 weeks old, weighing about 150 to 200 g, obtained from Pasteur Institute, Tehran. They were kept under the care of experienced animal technicians. The animals were housed in cages at  $24 \,^\circ$ C, with a 12 h light-dark cycle. The animals were given a standard rat chow diet.

Experimental diabetes was induced by administration of alloxan monchydrate (75 mg/kg of body weight) intraperitoneally two weeks before starting the treatment. After a 2-week experimental period, the blood glucose level of the animals was tested for evidence of a diabetic state, following a 12 h fast. The animals that had a blood glucose level equal to or greater than 250 mg/dl were included in the study (El-demerdash et al., 2005; Kulkarni et al., 2002; Ragavan and Krishnakumari, 2006). The study was reviewed and approved by the Ethics Committee of Isfahan University of Medical Sciences.

The rats were randomly divided into four groups (n = 6): normal control (non-diabetic control rats given normal saline intraperitoneally, daily for six weeks), diabetic control (untreated diabetic control rats given normal saline intraperitoneally, daily for six weeks to equalize stress induced by injections in all groups), diabetic-walnut oil (diabetic rats given extract of walnut oil intraperitoneally, daily for six weeks) and diabetic-glibenclamide (diabetic rats given glibenclamide 0.6 mg/kg intraperitoneally, daily for six weeks) (Dhandapani et al., 2002).

### Blood sample collection and analysis

Blood samples after 16 h of fasting were collected three times: time 0 was before injection of alloxan, time 1 was two weeks after injection of alloxan and time 2 was six weeks after injection of alloxan. Plasma insulin levels were estimated by enzyme-linked immunosorbant (ELISA) method, using the commercially available kit obtained from Boehringer-Mannheim, Germany (Enzymun-test system) on ES 33 analyzer, that is, blood glucose, hemoglobin A1c (HbA1c), total cholesterol (cho), triglyceride (TG) and low density lipoprotein-cholesterol (LDL-C), high density lipoprotein-cholesterol (HDL-C) and very low density lipoprotein-cholesterol (VLDL-C) levels were measured using special kits (DiaSys, Germany) which utilized the colorimetric method, in an autoanalyzer (Hitachi autoanalyzer, Hitachi Co., Tokyo).

#### Histological studies

After a 6-week experimental period and the last blood sampling, the whole pancreas was removed after sacrificing the animal and was fixed in 10% formalin for histopathological examination. Sections were cut and stained by hematoxylin and eosin (H&E) for histological examination (Nagappa et al., 2003).

#### Statistical analysis

All values were expressed as mean  $\pm$  standard deviation (SD). Significant differences among the groups were determined by oneway analysis of variance (ANOVA) using the Statistical Package for Social Sciences (SPSS) 13.0 software package program. Values of P < 0.05 were taken as statistically significant.

## RESULTS

## At time 0

As shown in Table 1, the levels of different biochemical

Parameter	Time	Non diabetic control	Diabetic control with no treatment	Diabetic treated with walnut oil	Diabetic treated with glibenclamide
	0	95.5 ± 13.52	98.5 ± 12.43	96.0 ± 13.21	95.0 ± 3.0
Glucose (mg/dl)	1	101.25 ± 26.22	492.4 ± 95.56* <sup>#</sup>	228 ± 22.13 <sup>#</sup> *	240 ± 26.45 <sup>#</sup> *
	2	97.25 ± 12.78	599 ± 82.56#**	86.3 ± 5.07‡	122.33 ± 32.30‡
Insulin (μu/ml)	0	13.52 ± 1.12	13.04 ± 1.26	13.1 ± 1.16	14 ± 1.35
	1	13.52 ± 1.06	$5.27 \pm 0.91^{#*}$	6.1 ± 1.07 <sup>#</sup> *	$6.6 \pm 0.83^{#*}$
	2	$12.55 \pm 1.35$	$4.86 \pm 1.41^{#**}$	$14.35 \pm 2.73^{\ddagger}$	12.5 ± 1.27 <sup>‡</sup>
HbA1c (%)	0	4.22 ± 0.22	4.25 ± 0.25	4.5 ± 0.27	4.03 ± 0.23
	1	$4.3 \pm 0.16$	4.44 ± 0.63	4.1 ± 0.12	$4.46 \pm 0.66$
	2	4.5 ± 0.21	$6.9 \pm 1.75^{#**}$	4.20 ± 0.21	$4.16 \pm 0.15$
LDL-C (mg/dl)	0	14.75 ± 2.5	11.16 ± 2.22	14.26 ± 3.22	13.66 ± 1.52
	1	15.0 ± 1.82	20.2 ± 1.48 <sup>#</sup> *	26.7± 3.05	$22 \pm 8.88^{#*}$
	2	16.25 ± 1.70	33.0 ± 3.36 <sup>#**‡</sup>	21 ± 3.11#** <sup>‡</sup>	15.66 ± 3.21
HDL-C (mg/dl)	0	37.25 ± 1.70	37.5 ± 1.87	36.5 ± 1.87	37.0 ± 1.0
	1	40.25 ± 10.87	$35.1 \pm 6.08^{\#}$	43.2± 5.54	49.0 ± 21.93
	2	41.75 ± 6.39	$33.33 \pm 6.65^{\#}$	41.4 ± 5.12	42.33 ± 9.45
VLDL-C (mg/dl)	0	17.45 ± 3.54	16.8 ± 3.32	16.8 ± 3.32	19.26 ± 1.13
	1	15.15 ± 3.04	23.56 ± 11.97 <sup>#</sup> *	16.45 ± 1.25 <sup>#</sup> *	17.66 ± 7.85 <sup>#</sup>
	2	$16.55 \pm 2.34$	$30.5 \pm 6.6^{\ddagger}$	$14.14 \pm 3.64$	$14.0 \pm 3.12$
Cholesterol (mg/dl)	0	61.25 ± 6.29	61.25 ± 6.18	61.5 ± 5.24	62.0 ± 7.21
	1	62.57 ± 3.20	91 ± 14.71 <sup>#</sup> *	96.6 ± 5.50* <sup>#</sup>	$100.0 \pm 34.21^{\#}$
	2	64.5 ± 5.19	127.5 ± 25 <sup>#</sup> ** <sup>‡</sup>	77.0 ± 5.31	$60.33 \pm 5.50$
Triglyceride (mg/dl)	0	87.25 ± 17.72	83.9 ± 16.52	84.0 ± 16.60	96.33 ± 5.68
	1	85.75 ± 15.23	117.8 ± 59.87 <sup>#</sup> *	85.2 ± 6.25	88.33 ± 39.27
	2	82.75 ± 11.70	52.5±33.04 <sup>#</sup> ** <sup>‡</sup>	69.7 ± 16.21	70.0 ± 15.62

**Table 1.** Different biochemical parameters levels in non-diabetic control, diabetic control, diabetic treated rats with walnut oil and diabetic treated rats with glibenclamide (Means ± SE).

\*Means differ significantly between time 0 and time 1(P < 0.05). \*\*Means differ significantly between time 2 and time 0 (P < 0.05). #Means differ significantly in comparison with non-diabetic control group (P < 0.05). ‡Means differ significantly between time 2 and time 1 (P < 0.05).

parameters in serum of all four groups had no meaningful deference.

# At time 1

Blood glucose level was found to be significantly increased in diabetic control rats, diabetic rats treated with walnut oil and diabetic rats treated with glibenclamide, and this was associated with decrease in insulin level. Cholesterol, VLDL-C, LDL-C and TG were significantly increased and HDL-C decreased in diabetic control. Cholesterol and VLDL-C were significantly increased in diabetic rats treated with walnut oil, and diabetic rats treated with glibenclamide LDL-C was significantly increased as compared to non diabetic control rats (P < 0.05), and also in comparison with time 0 (P < 0.05). Insulin significantly decreased at time 1 in all diabetic groups as compared to non diabetic control rats (P < 0.05), and also in comparison with time 0 (P < 0.05).

# At time 2

Treatments with either walnut oil or glibenclamide produced a significant decrease in blood glucose levels along with significant increase in insulin levels in comparison with time Table 2. Size of islets of Langerhans in four experimental groups.

	Time	Non diabetic control	Diabetic control with no treatment	Diabetic treated with walnut oil	Diabetic treated with glibenclamide
Size of Langerhans islets (micron)	2	1.64 ± 0.3	$0.62 \pm 0.4^{*}$	$1.02 \pm 0.44$	$1.48 \pm 0.6$

\*Means differ significantly between diabetic control and non-diabetic control group (P < 0.05).

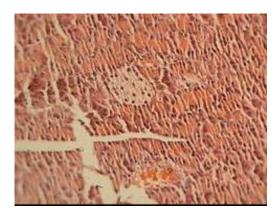


Figure 1. Cross-section of Langerhans islets in diabetic control group.

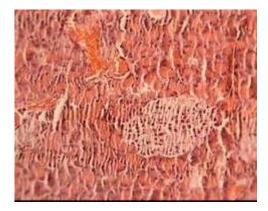


Figure 2. Cross-section of Langerhans islets in non-diabetic group.

1. LDL-C levels decreased significantly by walnut oil due to six weeks of treatment (Time 2). HbA1C was found to be significantly higher in diabetic control rats (P < 0.05). In contrast, diabetic rats treated with walnut oil and diabetic rats treated with glibenclamide showed lower mean HbA1C.

# Results of the histomorphometric study

Histopathological study of islets of Langerhans revealed that size of Langerhans islets in diabetic control group had a significant difference in comparison with non-

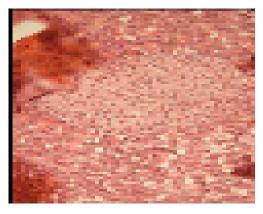
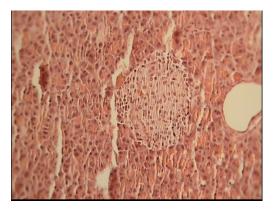


Figure 3. Cross-section of Langerhans islets in diabetic rats treated with walnut oil.



**Figure 4.** Cross-section of Langerhans islets in diabetic rats treated with glibenclamide.

diabetic control group (P < 0.05).

There was no significant difference in the size of Langerhans islets between the treated rats (with walnut oil or glibenclamide) and non-diabetic control rats (Table 2 and Figures 1 to 4).

# DISCUSSION

This study demonstrated that consumption of walnut oil for six weeks, significantly reduce biochemical parameters (blood glucose, insulin, HbA1C, total cholesterol, LDL-C, VLDL-C, HDL-C, and triglyceride) and size of Langerhans islets in alloxan-induced diabetic rats and when compared with the effects of glibenclamide. Alloxan and streptozotocin (STZ) are widely used to induce experimental diabetes in animals (Szkudelski, 2001). STZ action in B cells is accompanied by characteristic alterations in blood insulin and glucose concentrations. 2 h after injection, hyperglycemia is observed with a concomitant drop in blood insulin. About 6 h later, hypoglycemia occurs with high levels of blood insulin. Finally, hyperglycemia concentrations reflect abnormallities in B cell function. STZ impairs glucose oxidation (Bedoya et al., 1996) and decreases insulin biosynthesis and secretion (Bolaffi et al., 1987; Nukatsuka et al., 1990b). It was observed that STZ at first abolished the B cell response to glucose. Temporary return of responsiveness then appears, which is followed by its permanent loss and cells are damaged (West et al., 1996). Alloxan is rapidly taken up by the b cells, it is not toxic by itself (Wilson et al., 1984). The selective toxicity of alloxan to pancreatic b cells is due to its direct effect on islet cell permeability (Szkudelski, 2001; Watkins et al., 1979) and acts at the site of hexose transport as it inhibits glucose-stimulated insulin release (Robertson et al., 1988) and generates free radicals (Houee et al., 1981) that are toxic to various cellular constituents. It showed that significant decrease in blood glucose and elevated insulin level when STZ-diabetic rats were administered amaranth grain and its oil fraction (Kim et al., 2006). Walnut leaves constitute a good source of antioxidant compounds, namely phenolics, suggesting that it could be useful in prevention of diseases in which free radicals are implicated. Phenolic acid and flavonoid are two major groups of phenolic compounds in walnut leaves. The most important phenolic acid in walnut leaf is caffeoylquinic acid and the main flavonoid is guercetin (Fukuda, 2003; Pereira et al., 2007; Solar et al., 2006). Some studies have shown that flavonoids are able to decrease plasma glucose level (Li, et al., 2004). Quercetin inhibits glucose transporter (GLUT2), so diminishes glucose intestinal absorption (Kwon, 2007). Caffeoylquinic acid (chlorogenic acid) is a specific inhibitor of alucose-6-phosphate translocase and reduces hepatic glucose production (Hemmerle, 1997), thus decrease blood glucose level and HbA1C (Dhandapani, 2002). Plant antioxidants are able to restore and regenerate pancreatic B cells. The results from the studies on garlic, onion and fenugreek show that in diabetic rats treated with anti-oxidants the number of Langerhans islets has increased significantly (El-demerdash, 2005; Jelodar, 2005). Walnuts also contain several phytosterols, as it appears that they can inhibit intestinal absorption of cholesterol (Amaral et al., 2003). Fatty acid composition can influence various physiological and bio-chemical processes, including blood pressure regulation, glucose metabolism, lipid metabolism, platelet aggregation and erythrocyte deformability (Wright et al., 1988). Studies to evaluate that oils rich in v-3 and v-6 long-chain fatty acids can significantly attenuate alloxan-induced diabetes in

animals. These oils are able to inhibit or reduce the free radicals and thus prevent diabetes (Mohan and Das, 2001; Suresh and Das, 2006). Mohan and Das (2001) showed that PUFA-rich oils prevented alloxan-induced damage to the pancreatic b cells. Their ability to enhance antioxidant status, suppress cytokine production (Chew et al., 1997; Suresh and Das, 2001; Mohan and Das, 2001; Suresh and Das, 2003a; Suresh and Das, 2003b). Polyunsaturated fatty acids (PUFAs) and monounsaturated fatty acids (MUFAs) have received a lot of attention in the last decade and their health benefits are becoming increasingly evident (Jimenez-Gomez et al., 2008; Micallef and Garg, 2009; Ouellet et al., 2008). Oleic acid increase insulin secretion in the presence of the inflammatory cytokine TNF- $\alpha$ . The mechanisms of the anti-diabetogenic action of oleic acid are very likely multifaceted. The role of oleic acid in the presence of the proinflammatory cytokine TNF- $\alpha$  is clear. Dietary fatty acid intake, particularly unsaturated fatty acids, may also alter the composition of membrane phospholipids in addition to binding nuclear transcription factors. Changes phospholipid composition may in turn change in membrane fluidity; thereby, altering the binding of cytokines to their corresponding receptors. Arachidonic acidis is effective in preventing alloxan-induced toxicity to b cells in vivo, the ability of arachidonic acidis to prevent the development of diabetes induced by alloxan may be due to decreased production of free radicals and lipid peroxides (Laight et al., 1998; Das, 2001). Studies were also conducted specifically on walnuts, findings show that frequent consumption of moderate quantities of walnuts favorably modified the lipoprotein profile and decreased serum levels of total cholesterol (Houseknecht et al., 1998; Higa et al., 1999). Nuts are also rich in phytosterols, which due to their structural similarity with cholesterol, inhibit its intestinal absorption, thereby lowering total plasma cholesterol and LDL levels (Ha et al., 1990; Fukuzawa et al., 1999). The results of the present study suggest that walnut oil is effective in preventing alloxan-induced on biochemical parameters like blood glucose, insulin level, HbA1C, cholesterol, triglyceride, LDL-C, HDL-C and VLDL-C in diabetic rats.

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## REFERENCES

- Agrawal RP, Kochar DK, Sahani MS, Tuteja FC, Ghorui SK (2004). Hypoglycemic activity of camel milk in streptozotocin induced diabetic rats. Int. J. Diab. Dev. Countries, 24: 47-49.
- Al-Numair KS, Chandramohan G, Alsaif MA (2011). Effect of camel milk on collagen abnormalities in streptozotocin-diabetic rats. Afr. J.

Pharm. Pharmacol., 5(2): 238-243.

- Amaral JS, Casal S, Pereira JA, Seabra RM, Olivera BPP (2003). Determination of Sterol and Fatty Acid Compositions, Oxidative Stability, and Nutritional Value of Six Walnut (*Juglans regia* L.) Cultivars Grown in Portugal. J. Agric. Food Chem., 51: 7698–7702
- Bedoya FJ, Solano F, Lucas M (1996). N-monomethyl-arginine and nicotinamide prevent streptozotocin-induced double strand DNA break formation in pancreatic rat islets. Experientia, 52: 344-347.
- Bolaffi JL, Nagamatsu S, Harris J, Grodsky GM (1987). Protection by thymidine, an inhibitor of polyadenosine diphosphate ribosylation, of streptozotocin inhibition of insulin secretion. Endocrinology, 120: 2117-2122.
- Bucher HC, Hengstler P, Schindler C, Meier G (2002). N-3 polyunsaturated fatty acids in coronary heart disease: a metaanalysis of randomized controlled trials. Am. J. Med., 112: 298 304.
- Centers for Disease Control and Prevention. Preventing diabetes and its complications [article online], 2008. Available from http://www.cdc.gov/nccdphp/publications/factsheets/Prevention/diabe tes.htm. Accessed 22 July, 2009.
- Chew BP, Wong TS, Shultz TD, Magnuson NS (1997). Effects of conjugated dienoic derivatives of linoleic acid and beta-carotene in modulating lymphocyte and macrophage function. Anticancer Res., 17: 1099–1106.
- Das UN (2001). Nutritional factors in the pathobiology of human essential hypertension. 17: 337–346.
- Dhandapani S, Subramanian VR, Rajagopal S, Namasivayam N (2002). Hypolipidemic effect of *Cuminum cyminum* L. on alloxan– induced diabetic rats. Pharmacol. Res., 46: 251-255.
- Dunger A, Cunningham JM, Delaney CA, Lowe JE, Green MH, Bone AJ, Green IC.)1996). Tumor necrosis factor-alpha and interferongamma inhibit insulin secretion and cause DNA damage in unweaned rat islets: extent of nitric oxide involvement. Diabetes 45: 183–189.
- Duke JA (1989). Handbook of Nuts. CRC Press, London.
- Dhandapani S, Subramanian VR, Rajagopal S, Namasivayam N (2002). Hypolipidemic effect of *Cuminum cyminum* L. on alloxan– induced diabetic rats. Pharmacol. Res., 46: 251-255.
- El-demerdash FM, Yousef M I, Abou El Naga NI (2005). Biochemical study on the hypoglycemic effects of onion and garlic in alloxan induced diabetic rats. Food Chem. Toxicol., 43: 57-63.
- Feldman EB (2002). The scientific evidence for a beneficial health relationship between walnuts and coronary heart disease. J. Nutr., 132: 1062S–1101S.
- Flachs P, Mohamed-Ali V, Horakova O, Rossmeisl M, Hosseinzadeh-Attar MJ, Hensler M, Ruzickova J, Kopecky J.(2006). Polyunsaturated fatty acids of marine origin induce adiponectin in mice fed a high-fat diet. Diabetologia. 49:394-7.
- Fr ojd o S, Durand C, Pirola L (2008). Metabolic effects of resveratrol in mammals – a link between improved insulin action and aging. Curr. Aging Sci., 1: 145-151.
- Fukuda T, Ito H, Yoshida T (2003). Antioxidative polyphenols from walnuts (*Juglans regia* L.). Phytochemistry, 63: 795-801
- Ha YL, Storkson J, Pariza MW (1990). Inhibition of benzo(a)pyrene induced mouse forestomach neoplasia by conjugated dienoic derivatives of linoleic acid. Cancer Res., 50:1097
- Harper CR, Jacobson TA (2001). The fats of life: The role of Omega-3 Fatty Acids in the Prevention of Coronary Heart Disease. Arch. Intern. Med., 161: 2185 2192.
- Haffner SM, Hazuda HP, Mitchell BD, Patterson JK, Stern MP (1991). Increased incidence of type II diabetes in Mexican Americans. Diabetes Care, 14: 102-108.
- Hemmerle H, Burger HJ, Below P, Schubert G, Ripple R, Schindler PW, Paulus E, Herling AW (1997). Chlorogenic acid and synthetic chlorogenic acid derivatives: novel inhibitors of hepatic glucose-6phosphate translocase. J. Med. Chem., 40: 137-145.
- Higa M, Zhou YT, Ravazzola M, Baetens D, Orci L, Unger RH. (1999). Troglitazone prevents mitochondrial alterations, beta cell destruction, and diabetes in obese prediabetic rats. Proc. Natl. Acad. Sci. USA. 96: 11513.
- Houseknecht KL, Vanden Heuvel JP, Moya-Camarena SY, Portocarrero CP, Peck LW, Nickel KP, Belury MA (1998). Dietary conjugated linoleic acid normalizes impaired glucose tolerance in the Zucker diabetic fatty fa/fa rat. Biochem. Biophys. Res. Commun., 1998;

244:678.

- Houee C, Gardes M, Pucheault J, Ferradini C (1981). Radical chemistry of alloxan-dialuric acid: role of the superoxide radical, Bull. Eur. Physiopathol. Respir., 17: 43-48.
- Jelodar G, Maleki M, Motadayen MH, Sirus S (2005). Effect of fenugreek, onion and garlic on blood glucose and histopathology of pancreas of alloxan-induced diabetic rats. Indian J. Med. Sci., 59: 64-69.
- Jimenez-Gomez Y, Lopez-Miranda J, Blanco-Colio LM, Marin C, Perez-Martinez P, Ruano J, Paniagua JA, Rodriguez F, Egido J, Perez-Jimerez F (2008). Olive oil and walnut breakfasts reduce the postprandial inflammatory response in mononuclear cells compared with a butter breakfast in healthy men. Atherosclerosis, 204:e70–76.
- Kim HK, Kim MJ, Cho HY, Kim EK, Shin DH (2006). Antioxidative and anti-diabetic effects of amaranth (*Amaranthus esculantus*) in streptozotocin-induced diabetic rats. Cell Biochem. Funct., 24: 195-199.
- Knowler WC, Pettitt DJ, Savage PJ, Bennett PH (1981). Diabetes incidence in Pima Indians: contribution of obesity and parental diabetes. Am. J. Epidemiol., 113: 144-156.
- King H, Zimmet P, Raper LR, Balkau B (1984). The natural history of impaired glucose tolerance in the Micronesian population of Nauru: a six year follow-up study. Diabetologia, 26: 39-43.
- Kim NN (2009). Sex steroid hormones in diabetes-induced sexual dysfunction: focus on the female gender. J. Sex Med., 6(3): 239-246.
- Kulkarni JS, Metha AA, Santani DD, Goyal RK (2002). Effects of chronic treatment with cromakalim and glibenclamide in alloxan–induced diabetic rats. Pharmacol. Res., 46: 101-105.
- Kumar MV, Vanegas SM, Patel N, Aitken JD, Ziegler TR, Ganji V (2011). Fish oil rich diet in comparison to saturated fat rich diet offered protection against lipopolysaccharide-induced inflammation and insulin resistance in mice. Nutrition & Metabolism, 8: 16, 11-30.
- Kwon O, Esk P, Chen S, Corpe CP, Lee JH, Kruhlak M, Levine M (2007). Inhibition of the intestinal glucose transporter GLUT2 by flavonoids. FASEB J., 21: 366-377.
- Laight DW, Kaw AV, Carrier MJ, Anggard EE (1998). Interaction between superoxide anion and nitric oxide in the regulation of vascular endothelial function, Br. J. Pharmacol., 124: 238-244.
- Lillioja S, Mott DM, Spraul M, Ferraro R, Foley JE, Ravussin E, Knowler WC, Bennett PH, Bogardus C(1993). Insulin resistance and insulin secretory dysfunction as precursors of non-insulin-dependent diabetes mellitus: prospective studies of Pima Indians. N. Engl. J. Med., p. 329.
- Li WL, Zheng HC, Bukuru J, De Kimpe N (2004). Natural medicines used in the traditional Chinese medical system for therapy of diabetes mellitus. J. Ethnopharmacol., 92: 1-21.
- Mohamed A, Mehaia MA, Hablas KM, Abdel-Rahman SA (1995). El-Mougy Milk composition of Majaheim, Wadah and Hamra camels in Saudi Arabia. Food Chem., 52: 115-122.
- Mohan IK, Das UN (2001). Prevention of chemically induced diabetes mellitus in experimental animals by polyunsaturated fatty acids. Nutrition, 17: 126-151.
- May Y, Njike VY, Mille TJ, Dutta S, Doughty K, Treu J, Katz DL (2010). Effects of Walnut Consumption on Endothelial Function in Type 2 Diabetic Subjects Diabetes Care, 33: 227-232.
- Micallef MA, Garg ML (2009). Anti-inflammatory and cardioprotective effects of n-3 polyunsaturated fatty acids and plant sterols in hyperlipidemic individuals. Atherosclerosis, 204: 476-482.
- Martínez ML, Labuckas DO, Lamarque AL, Maestri DM (2010). Walnut (*Juglans regia* L.): genetic resources, chemistry, by-products. J. Sci. Food Agric., 90: 12, 1959-1967.
- Nielsen JN (1986). Affinity purified human interleukin-1 is cytotoxic to isolated islets of Langerhans. Diabetologia, 29: 63–67.
- Nagappa AN, Thakurdesai PA, Venkat Rao N, Singh J (2003). Antidiabetic activity of *Terminalia catappa* Linn fruits. J. Ethnopharmacol., 88: 45-50.
- Nukatsuka M, Yoshimura Y, Nishida M, Kawada J (1990b). Importance of the concentration of ATP in rat. pancreatic beta cells in the mechanism of streptozotocin-induced cytotoxicity. J. Endocrinol., 127: 161-165.
- Ouellet V, Weisnagel SJ, Marois J, Bergeron J, Julien P, Gougeon R, Tchernof A, Holub BJ, Jacques H (2008). Dietary cod protein reduces

plasma C-reactive protein in insulin-resistant men and women. J. Nutr., 138: 2386-2391.

- Perry LM (1980). Medicinal Plants of East and Southeast Asia: Attributed Properties and Uses. MIT Press, Cambridge, MA, USA. ISBN 9780262160766.
- Pereira JA, Oliveira I, Sousa A, Valentao P, Andrade P, Ferreira I, Ferreres F, Bento A, Seabra R, Estevinho L (2007). Walnut (*Juglans regia* L.) leaves: phenolic compound, antibacterial activity and antioxidant potential of different cultivars. Food Chem. Toxicol., 45(11): 2287-2295.
- Robertson DG, DiGirolamo M, Merrill AH, Lambethll D (1988). Insulinstimulated Hexose Transport and Glucose Oxidation in Rat Adipocytes Is Inhibited by Sphingosine at a Step after Insulin Binding. J Biol. L. Chem., 269(12): 6773-6779.
- Ragavan B, Krishnakumari S (2006). Antidiabetic effect of *T. Arjuna* bark extract in alloxan induced diabetic rats. Indian J. Clin. Biochem., 21: 123-128.
- Shivananda NB, Geetha B (2005). Relationship between Sialic acid and metabolic variables in Indian type 2 diabetic patients. Lipids in Health and Disease 4: 15 doi: 10.1186/1476-511X-4-15.
- Singh R (2001). Annual Report NRCC, Bikaner, p. 50.
- Solar A, Colaric M, Usenik V, Stampar F (2006). Seasonal variations of selected flavonoids, phenolic acids and quinones in annual shoots of common walnut (*Juglans regia* L.). Plant Sci., 170: 453-461.
- Suresh Y, Das UN (2001). Protective action of arachidonic acid against alloxan-induced cytotoxicity and diabetes mellitus, Prostaglandins Leukot. Essent. Fatty Acids, 64: 37-52.
- Suresh Y, Das UN (2003a). Long-chain polyunsaturated fatty acids and chemically induced diabetes mellitus: effect of o-6 fatty acids. Nutrition, 19: 93-114.
- Suresh Y, Das UN (2003b). Long-chain polyunsaturated fatty acids and chemically induced diabetes mellitus: effect of o-3 fatty acids. Nutrition, 19: 213-228.
- Szkudelski T (2001). The mechanism of alloxan and streptozotocin action in B cells of the rat pancreas. Physiol. Res., 50: 536-546.

- Szkudelski T, Szkudelska K (2011). Anti-diabetic effects of resveratrol. Ann. N.Y. Acad. Sci., 34-39.
- Szkudelska K, Szkudelski T (2010). Resveratrol, obesity and diabetes. Eur. J. Pharmacol., 635: 1-8.
- Suresh Y, Das UN (2006). Differential effect of saturated, monounsaturated, and polyunsaturated fatty acids on alloxaninduced diabetes mellitus Prostaglandins. Leukotrienes and Essential Fatty Acids, 74: 199-213
- Watkins D, Cooperstein SJ, Feil S (1979). Studies on the selectivity of alloxan for the beta cells of the islets of langerhans: effect of pH on the in vitro action of alloxan. J. Pharmacol. Exp. Ther., 208: 184-189.
- West E, Simon OR, Morrison EY (1996). Streptozotocin alters pancreatic beta-cell responsiveness to glucose within six hours of injection into rats. West Indian Med. J., 45: 60-62.
- Wright JR, Lefkowith JB, Schreiner G, Lacy PE (1988). Essential fatty acid deficiency prevents multiple low-dose streptozotocin-induced diabetes in CD-1 mice. Proc. Natl. Acad. Sci. USA, 85: 6137
- Wilson GL, Patton NJ, McCord JM, Mullins DW, Mossman BT (1984). Mechanisms of streptozotocin- and alloxan-induced damage in rat B cells. Diabetologia, 27(6): 587-591.