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Review

Phytochemistry and mode of action of some tropical spices in the management of type-2 diabetes and hypertension

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Spices are important food supplements and/or food products, which have been used as flavouring agents and preservatives for thousands of years in tropical Africa, Asia and other parts of the world. They are well known for their medicinal properties, and their use in traditional systems of medicine has been on record for a long time. Although, epidemiological and clinical studies have indicated that spices are important source of natural antioxidant having the digestive stimulant action, bioavailability enhancement nature, carminative attribute, antimicrobial activity, hypolipidemic property, antidiabetic influence, antioxidant capacity, anti-inflammatory ability, anticarcinogenic potential and neuroprotective effect. The present review reports the phytochemical constituents and mode of action of some tropical spices as antidiabetic and antihypertensive agents. The conclusion of this review may help in undertaking research for the development of functional foods and nutraceuticals.

Key words: Spices, type-2 diabetes, hypertension, enzymes, antioxidant, phenolics, phytochemicals.

INTRODUCTION

Spices are food supplements or food products, which have been used not only as flavoring and coloring agents, but also as food preservatives and herbs in folk medicines for thousands of years in Africa, Asia and other parts of the world (Srinivasan, 2005a). They are consumed as whole spices or ground into powder and mixed with diets containing cereals, legumes, nuts, fruits, vegetables, milk and milk products. They are also used in soup preparation in various homes and serve as ingredients in the preparation of several traditional delicacies. Spices are utilized as herbs, mainly in the form of isolates from their extracts. Spices are considered to be good contributors to the total nutrient intake of protein, carbohydrates, fats, vitamins and minerals, thereby enhancing the nutritional quality of diets (Pradeep et al., 1993). Apart from the nutrients supplied by spices, they possess many phytochemicals which are

potential sources of natural antioxidant such as phenolic diterpenes, volatile oils, flavonoids, terpenoids, carotenoids, phytoestrogens, and phenolic acids (Cai et al., 2004; Suhaj, 2006; Kennedy et al., 2011).

Spice phytochemicals such as curcumin (turmeric), capsaicin (red chillies), eugenol (cloves), linalool (coriander), piperine (black pepper), zingerone (zinger) and cuminaldehyde (cumin) have been reported to inhibit lipid peroxidation (Shobana and Naidu, 2000; Oboh and Rocha, 2007). In recent times, spice antioxidants have raised considerable interest among food scientists, manufactures, and consumers because of their natural antioxidants (Lu et al., 2011). Consumers are increasingly aware of the risk posed by synthetic antioxidants due to their high volatility and instability at elevated temperatures. Therefore, focus has been shifted to the use of natural antioxidants in food preservation (Odukoya et al., 2005; Oboh and Rocha, 2007; Adefegha and Oboh, 2011a).

Food oxidation is considered a major cause of food deterioration and spoilage, causing rancidity in food (Sherwin, 1990). The resultant effect is noticed in the

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decreased nutritional quality, color, flavor, texture and safety of foods. Many spices have also been recognized to possess digestive stimulant action, carminative effect, antimicrobial activity, antioxidant capacity, antiinflammatory property, antimutagenic ability and anticarcinogenic potential (Srinivasan, 2005a). Spices contribute greatly to the daily antioxidant intake in most diets, especially in dietary cultures where spices are used as whole meal (Carlsen et al., 2010). Many spices have been shown to confer health benefits and have been proven to counteract oxidative stress in vitro and in vivo (Oboh et al., 2005, 2010a, 2012a; Shan et al., 2005; Wojdylo et al., 2007; Adefegha and Oboh, 2012a). They are common sources of phenolic compounds which have been reported to show superior antioxidant capacity to fruits, cereals, and nuts (Pellegrini et al., 2006; Carlsen et al., 2010). The main active components in spices are phenolic acids, flavonoids and volatile or essential oils (Shan et al., 2005; Wojdyło et al., 2007; Viuda-Martos et al., 2011; Lu et al., 2011).

In Nigeria, over 100 indigenous spices are used as important components of the "African/Nigerian dishes", bringing original favors and desirable sensory properties to food. Essentially, "Pepper soup" is famous for its sensory, aromatic, attractive, pungency and spicy flavor resulting from the use of bastered melegueta, clove, alligator pepper, ginger, black pepper, garlic, Ethiopian pepper, chili peppers and other spices. Common spices, such as sweet basil, clove, black pepper, turmeric, chili pepper, and ginger are usually part of daily African household meals and also used as traditional African medicine. Numerous studies have reported that spices are important source of natural antioxidant, possessing digestive stimulant action, bioavailability enhancement nature, carminative attribute, antimicrobial activity, hypolipidemic property, antidiabetic influence, anti-inflammatory ability, anticarcinogenic potential and neuroprotective effect (Srinivasan et al., 2004; Shan et al., 2005; Srinivasan, 2005a; Adefegha and Oboh, 2011b).

Diabetes mellitus (DM) is one of the leading causes of global morbidity and mortality, and a major risk for cardiovascular diseases (Alderman et al., 1999). Diabetes mellitus is a metabolic disease characterized by hyperglycaemia resulting from defects in insulin secretion, insulin action, or both (World Health Organization (WHO), 1999). The control of postprandial hyperglycemia is an important strategy in the management of diabetes mellitus, especially type 2 diabetis mellitus (T2DM), and reducing chronic complications associated with the disease (Kim et al., 2000; Ali et al., 2006; Ortiz-Andrade et al., 2007). Hence, the inhibition of enzymes (α-Glucosidase and α -amylase) involved in the digestion of carbohydrates can significantly decrease the postprandial increase of blood glucose after a mixed carbohydrate diet, by delaying the process of carbohydrate hydrolysis and absorption and phenolic phytochemicals from spices and have shown promising potentials (Oboh et al., 2010b;

Adefegha and Oboh, 2012a).

Hypertension or persistent high blood pressure is a common cardiovascular disease which has become a worldwide problem of epidemic proportions, affecting 15 to 20% of all adults with ailments such as arteriosclerosis, stroke, myocardial infarction and end-stage renal disease (Je et al., 2009). It is regarded as one of the long-term complications of T2DM. These two diseases (hypertension and T2DM) are interrelated metabolic disorders with persistent hypertension being the risk factors for strokes, heart attacks, heart failure and is a leading cause of chronic renal failure (Sowers and Epstein, 1995; Bakris et al., 2000). One of the therapeutic strategies towards the management of hypertension is the inhibition of angiotensin-I converting enzymes (ACE); an enzyme which play a pivotal role in rennin-angiotensin system by angiotensin-I to angiotensin-II converting vasoconstrictor). However, phenolic-rich spices have reported to act as good ACE inhibitors (Ranilla et al., 2010). Therefore, the present review highlights the phytochemical constituents and mechanism of action of some tropical spices as antidiabetic and antihypertensive agents. The pictures of some spices are as shown in Figure 1

Phytochemical composition

Spices in the diet are not considered vital from the nutritional point of view, though they are widely consumed throughout the world. They are not normally included in diet surveys, nor are they suggested or recommended in what are known as balanced diets. probably because it was thought that the intake of these spices was so small that their contribution of nutrients may not be significant (Pradeep et al., 1993). As part of normal diet, plant foods are thus not only a source of nutrients and energy provider, but may confer additional role of providing health benefits beyond their basic nutritional functions (Shahidi and Naczk, 2004). Attention is being focused on identifying dietary phytochemicals which are plant secondary metabolites (array of bioactive constituents) that are capable of eliciting health enhancing effects and disease preventing abilities (Visioli and Galli, 1998).

Phytochemicals describe the chemicals present in different parts of plant organs (leaves, stems, roots, flowers, fruits and seeds). Consumption of food rich in several phytochemicals such as saponins, alkaloids, terpenes, phenylpropanoids, isoprenoids, steroids, coumarins, flavonoids, phenolic acids, lignans, contain chemicals such as flavonoids, terpenoids, lignans, sulfides, polyphenolics, carotenoids, coumarins, saponins and plant sterols with biological activities that may provide therapeutic effects (Dorman et al., 2003, 2004; Cuvelier et al., 2004; Shan et al., 2005; Ninfali et al., 2005; Adefegha and Oboh, 2012b). The presence of



Figure 1. Pictures of spices.

phytochemicals has been shown to contribute immensely to the protective potential against degenerative diseases, therapeutic effects essential to preventing diseases, and nutritional quality of food and food products (Chu et al., 2002; Oboh et al., 2010b).

Our recent study revealed the presence of flavonoid and cardiac glycoside in Ethiopian pepper [Xylopia aethiopica [Dun.] A. Rich (Annonaceae)], nutmeg [Monodora myristica (Gaertn.) Dunal (Annonaceae)], clove [Syzygium aromaticum [L.] Merr. et Perry guineense (Myrtaceae)], ashanti pepper [Piper Schumach. et Thonn (Piperaceae)], bastered melegueta [Aframomum danielli K. Schum (Zingiberaceae)] and alligator pepper [Aframomum melegueta (Rosc.) K. Schum (Zingiberaceae)], tannin in Ethiopian pepper and clove, phlobatanin and anthraquinone in clove and saponin in Ethiopian pepper, clove, ashanti pepper, bastered melegueta and alligator pepper (Adefegha and Oboh, 2012a). Other active components of spices (Figure 2) such as curcumin (turmeric), capsaicin (bird's pepper), eugenol (cloves), linalool (coriander and sweet basil), piperine (black pepper), gingerol (ginger) and allicin (garlic) have been reported to inhibit lipid peroxidation in various tissues (Lawson, 1998; Ursell, 2000; Srinivasan, 2005a; Hiyasat et al., 2009; Adefegha and Oboh, 2011b).

Polyphenols

Food contains several chemicals, many of which have specific biological activity. The chemicals also interact with each other, confounding any effort to identify bioactives. Among these bioactive components are the phenolics. Dietary phenolics are secondary metabolites which are widely present in diets rich in vegetables, fruits, legumes, cereals, nuts, and have been linked to various beneficial effects on human health, such as minimizing the risk of developing coronary heart disease, cancer, hypertension, diabetes, and inflammatory processes (Scalbert et al., 2005; Zafra-Stone et al., 2007). They are present in plants as derivatives and/or isomers of flavones, isoflavones, flavonols, catechins, and phenolic acids.

Phenolic compounds in plants can be divided into two major categories: phenolic acids and flavonoids. Phenolic acids account for about 33% of the total phenolic intake; mainly the derivatives of benzoic or cinnamic acid. Flavonoids, on the other hand account for the remaining 67% of the total phenolic intake (Scalbert and Williamson, 2000). They are regarded as the most abundant polyphenols in human diets, and are mainly divided into: anthocyanins (colourful compounds) and anthoxanthins (colorless compounds), which can be subdivided into flavones, flavans, flavonols, flavanols, isoflavones, and their glycosides (Bravo, 1998; Liu, 2004). Phenolic compounds in plants are usually found either as free or bound forms (Chu et al., 2002; Sun et al., 2002). Free

soluble phenolics may be available as aglycones while the bound phenolics may be present as conjugates (glycosides and esters) or attached to the plant cell wall (Rice-Evan et al., 1996).

Hydrolysis by chemical and enzymatic means may be necessary to release the bonds (Stalikas, 2007). Most of the biological actions such as antimicrobial, hypolipidemic, antidiabetic, antilithogenic, antioxidant, antiinflammatory, antimutagenic, anticarcinogenic and neuroprotective properties observed in some spices have been attributed to the presence of phenolic compounds (Cai et al., 2004; Liu et al., 2008; Shan et al., 2005; Wojdylo et al., 2007; Muchuweti et al., 2007; Konczak et al., 2010). Shan et al. (2005) reported that spices from Labiatae, Myrtaceae and Compositae families are rich in rosmarinic acid, caffeic acid and volatile oil. In the same vein, chlorogenic acid, rutin, quercetin, and naringin were also identified as the dominant phenolic compounds in spices from the family Rutaceae (Lu et al., 2011).

In an experiment carried out on phenolic composition of three commercial herbal drugs and spices from lamiaceous species: Thymi herba (thyme), Serpylli herba (wild thyme) and Majoranae herba (sweet marjoram) using high performance liquid chromatography (HPLC) and high performance thin layer chromatography (HPTLC) methods, luteolin-7-O-β-glucuronide, lithospermic acid, rosmarinic acid and methyl rosmarinate, together with other known compounds, were detected quantified. Luteolin-7-O-β-glucuronide and lithospermic acid were identified as novel wild thyme luteolin-7-O-β-glucuronide and constituents. methyl rosmarinate as novel compounds in sweet marjoram.

Methyl rosmarinate was found to be present in thyme. The amount of polyphenol investigated in herbal drugs and spices has reached 84.3 mg/g dried spices (Fecka and Turek, 2008). Previous report on phenolic composition of thyme indicated the presence of caffeic acid, rosmarinic acid, apigenin, luteolin, luteolin-7-O-βglucuronide, luteolin-7-O-β-glucoside, 6-hydroxyluteolin glycosides, polymethoxyflavones, narirutin, eriodictyol, eriocitrin, hesperidin and taxifolin (Dapkevicius et al., 2002; Haraguchi et al., 1996; Kobayashi et al., 2003; Kosar et al., 2005; Miura et al., 2002; Watanabe et al., 2005). Wojdyło et al. (2007) also identified and quantified major phenolics by reverse-phase high-performance liquid chromatography (RP-HPLC) in thirty two (32) selected herbs and spices. The prominent phenolic acids reported were caffeic, p-coumaric, ferulic and neochlorogenic acids while the flavonoids detected were quercetin, luteolin, apigenin, kaempferol and isorhamnetin. Caffeic acid was reported as the predominant phenolic compound in sage (Salvia officinalis) (296 mg/100 g dry weight), thyme (T. vulgaris) (517 mg/100 g dry weight) and oregano (Origanum vulgare) (649 mg/100 g dry weight).

Luteolin (616 mg/100 g dry weight) and caffeic acid (406 mg/100 g dry weight) were also present in

Figure 2. Active ingredients of biological relevance present in abundance in some spices.

abundance in rosemary (Rosmarinus officinalis). Turmeric (Curcuma longa) was reported to contain pcoumaric acid (5.96 mg/100 g dry weight) and ferulic acid (17.6 mg/100 g dry weight) in small quantities. A minute amount of ferulic acid was reported to be present in nutmeg (M. fragrans). Quercetin was the only phenolic compound detected in clove (S. aromaticum) (Wojdyło et al., 2007). In a similar study, the phenolic compounds of some everyday-use spice plants, such as onion, dill, parsley and celery were identified and quantitatively assessed by HPLC. Chlorogenic acid was reported as the dominant phenolic compound in celery leaves and parsley leaves, and o-hydroxycinnamic acid dominant in dill (Stankevičius et al., 2010). Shan et al. (2005) also reported the presence of phenolic acids caffeoyl derivatives), (rosmarinic acid. phenolic diterpenes, volatile compounds (carvacrol), flavonoids (catechin) in sweet basil (O. basilicum L.), phenolic acids (rosmarinic acid), phenolic diterpenes (carnosic acid), volatile compounds and flavonoids in sage (S. officinalis L.), phenolic acids (caffeic acid, rosmarinic acid, caffeoyl derivatives), phenolic diterpenes (carnosic acid, carnosol, epirosmanol), volatile compounds (carvacrol), flavonoids in rosemary (R. officinalis L.), phenolic acids (gallic acid, caffeic acid, rosmarinic acid), volatile compounds (thymol), phenolic diterpenes, flavonoids in thyme (T. vulgaris L.), phenolic acids (gallic acid), flavonol glucosides, phenolic volatile oils (eugenol, acetyl eugenol), tannins in clove (*Eugenia caryophylata* Thunb.), phenolic volatile oils, phenolic acid (caffeic acid), flavanols (catechin) in nutmeg (M. fragrans Houtt) and phenolic acids (caffeic acid, p-coumaric acid, rosmarinic acid, caffeoyl derivatives), volatile compounds, (carvacrol) and flavonoids in oregano (*O. vulgare* L.).

In a similar manner, Hossain et al. (2010) reported the presence of thirty eight (38) phenolic compounds in five Lamiaceae spices: rosemary, oregano, sage, basil and thyme using Liquid chromatography coupled with electron span ionization detector and mass spectrometer (LC-ESI-MS/MS). Twenty (20), twenty six (26), twenty three (23), twenty four (24) and twenty (20) different phenolic compounds were found in rosemary, oregano, sage, basil and thyme, respectively. The structures of some of these phenolic compounds found in spices are as shown in Figure 3.

Antioxidant properties

Several oxygen-free radicals and other reactive oxygen species (ROS), which include free radicals such as superoxide anion radicals (O_2) , hydroxyl radicals (OH) and non free-radical species such as hydrogen peroxide (H_2O_2) and singlet oxygen (O_2) , may be formed in the human body during the normal cellular metabolism and in the food system in the course of food production and processing (Halliwell and Gutteridge, 1999; Halliwell, 2006; Oboh and Rocha, 2007). These radicals induce lipid peroxidation, thereby causing oxidative damage by oxidizing biomolecules such as proteins, lipids and DNA, leading to cell death, tissue damage and diseases such as atherosclerosis, cancer, emphysema, cirrhosis and arthritis (Kehrer, 1993). They could also result in food deterioration. On the other hand, antioxidant refers to a

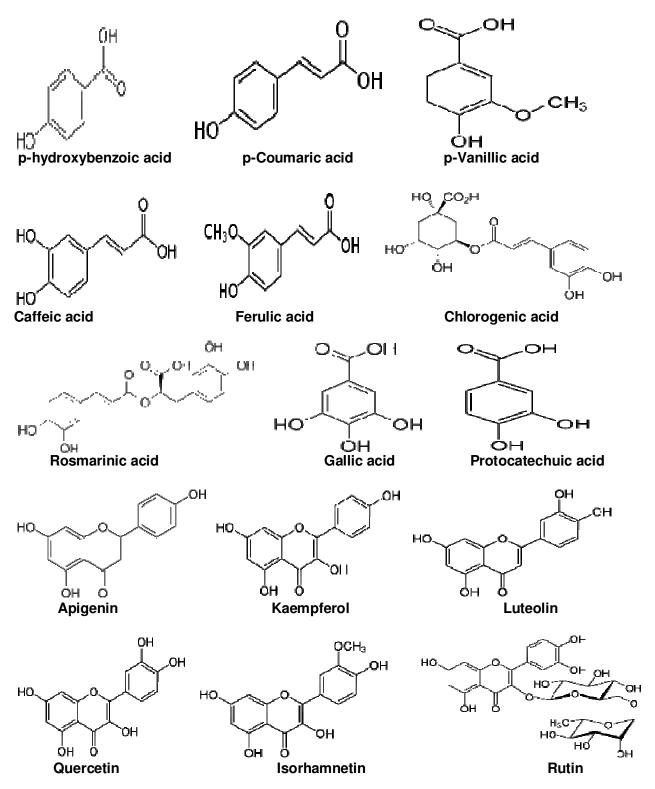


Figure 3. Structure of some phenolic constituents found in some spices.

compound that can delay or inhibit the oxidation of lipids or other molecules by inhibiting the initiation or propagation of oxidative chain reactions, and can thus prevent or repair the damage done to the body's cells by oxygen (Halliwell et al., 1995).

Antioxidants protect by contributing an electron of their own. In so doing, they neutralize free radicals and help prevent cumulative damage to body cells and tissues (Alia et al., 2003). Much of the total antioxidant activity of plant foods is related to their phenolic content, and not only to their vitamin contents (Chu et al., 2002; Sun et al., 2002; Oboh and Rocha, 2007; Oboh et al., 2008). They exert their antioxidant activity by removing free radicals, chelating metal catalysts, activating antioxidant enzymes, reducing α -tocopherol radicals, and inhibiting oxidases (Amic et al., 2003). Antioxidant may be endogenous or exogenous (can scavenge/deactivate this reactive free radicals, turning them to harmless particles) (Chu et al., 2002). Consumption of antioxidant rich food such as spices, fruits, and vegetables could be a practical approach towards improving antioxidant status, thereby enhancing good health and preventing disease (Chu et al., 2002; Oboh and Rocha, 2007; Carlsen et al., 2010).

Since the antioxidant capacity of plant extracts from different plant foods have been attributed to their high phenolic contents, in the last years, we have assessed the total phenols and flavonoids of different plant foods (Oboh and Rocha, 2007; Adefegha and Oboh, 2011a, 2011b; Oboh et al., 2008, 2010a; 2010b; 2011; 2012a, b, The antioxidant properties have also been investigated using several methods: 1,1-diphenyl-2picrylhydrazine (DPPH) radical scavenging (Gyamfi et al., 1999), β-carotene linoleic acid bleaching assay (Pratt, 1980), inhibition of linoleic acid peroxidation (Osawa and Namiki, 1981), ferric reducing antioxidant power (FRAP), total radical trapping antioxidant potential (TRAP) assay (Lissi et al., 1992, 1995), oxygen radical absorbance capacity (ORAC) assay (Cao et al., 1993). 15-lipoxygenase inhibition (Lyckander and Malterud, 1992), lipid peroxidation (LPO) method (Ohkawa et al., 1979), nitro blue tetrazolium (NBT) reduction assay or superoxide anion scavenging activity (Beauchamp and Fridovich, 1971), hydroxyl radical scavenging activity (Halliwell et al., 1987), hydrogen peroxide scavenging activity (Ruch et al., 1989), 2,2-azinobis-(3ethylbenzthiazoline-6-sulphonic acid) (ABTS) radical scavenging method (Re et al., 1999), reducing power assay (Oyaizu, 1986), ammonium thiocyanate (ATC) assay method (Masude et al., 1992) and ferric thiocyanate (FTC) method (Mitsuda et al., 1996).

In a number of studies, spices from different botanical families, such as Myristicaceae (for example nutmeg), Zingiberaceae (for example, ginger, galangal alligator pepper and bastered melegueta), Lamiaceae (for example, sweet basil, thyme, rosemary, oregano, and sage), Lauraceae (for example, cinnamon), Piperaceae (for example, black pepper, white pepper), Myrtaceae (for example, clove), Solanaceae (for example, chili pepper) and Umbelliferae (for example, Fennel, cumin, and Angelica dahurica), have been assessed for their antioxidant properties using some of the aforementioned assay methods (Zheng and Wang, 2001; Shan et al., 2005; Oboh et al., 2010b; Lu et al., 2011; Adefegha and Oboh, 2011b). Reports revealed that these spices possess moderate and high antioxidant activity (Shan et

al., 2005; Lu et al., 2011; Adefegha and Oboh, 2011b).

Previous studies have shown that these spices are rich in phenols and flavonoids, hence their disease preventing and health promoting abilities have been attributed to the presence of these phytochemicals (Fasoyiro et al., 2006; Olonisakin et al., 2006; Wojdyło et al., 2007; Uwakwe and Nwaoguikpe, 2008; Ezekwesili et al., 2010; Doherty et al., 2010). Several studies have also correlated antioxidant capacity with total phenolic content of legumes, vegetables, pepper, spices, medicinal herbs and other plant foods (Oboh, 2006; Oboh and Rocha, 2007; Oboh et al., 2008, 2011; Oboh and Ogunruku, 2010; Adefegha and Oboh, 2011b; Oboh and Ademosun, 2011). These reports may validate the claims that phenolic compounds are responsible for most of the antioxidant effects in plants (Pietta, 2000; Chu et al., 2002; Sun et al., 2002; Cai et al., 2004; Liu et al., 2008; Shan et al., 2005; Odukoya et al., 2005; Oboh and Rocha, 2007).

Our report on the phenolic content and antioxidant properties of aqueous extract of some Nigerian spices: M. myristica (Africa nutmeg), X. aethiopica (Ethiopian pepper), S. aromaticum (tropical cloves), P. guineense (Black pepper), A. danielli (bastered melegueta), A. melegueta (alligator pepper/grains of paradise) and Clerodendrum volubile (Locally known as "Obenetete")] indicated that the total phenol content of the spices range from 0.6 (M. myristica) to 2.28 mg gallic acid equivalents per g (mg GAE/g) (A. melegueta). In the same vein, A. melegueta (0.55 mg GAE/g) was reported to have the highest flavonoid content, followed Clerodendrum volubible (0.52 mg GAE/g), P. guineense (0.41 mg GAE/g), and Aframomum danielli (0.29 mg GAE/g), Syzygium aromaticum (0.26 mg GAE/g), Xylopia aethiopica (0.24 mg GAE/g) and Monodora myristica (0.21 mg GAE/g) (Adefegha and Oboh, 2011a).

Furthermore, the spice extracts also showed interesting antioxidant properties as typified by their ferric reducing antioxidant property, Fe2+-chelating ability, inhibition of Fe²⁺/H₂O₂-induced decomposition of deoxyribose and inhibition of Fe²⁺-induced lipid peroxidation in rat's brain (Adefegha and Oboh, 2011a). Shan et al. (2005) reported the phenolic contents and total antioxidant capacity of 26 spices by assessing the ability of their extracts to scavenge free radicals using the ABTS model. The ABTS radical scavenging activity of the spice extracts ranged from 0.55 (poppy) to 168.7 mmol trolox equivalents antioxidant capacity per 100 g (TEAC/100 g) dry weight (clove). Also, the total phenol content of the spices was reported to range from 0.04 to 14.38 g of gallic equivalents per 100 g (GAE/100 g) of dry weight (DW). Hence, positive correlation between the total phenol contents and the total antioxidant capacity was observed in the spices (Shan et al., 2005; Adefegha and Oboh, 2011a).

In another study where the antioxidant properties of 30 spices was assessed using FRAP, ABTS radical scavenging ability and microsomal lipid peroxidation

(MLP) assays (Hossain et al., 2008). It was reported that clove exhibited the highest ABTS radical scavenging ability, FRAP and anti-radical powers (ARP) on microsomal lipid peroxidation. Rosmarinic acid and eugenol, commonly found in clove was reported to possess higher antioxidant capacities than that of the synthetic antioxidants tested (Hossain et al., 2008). This could be an indication that natural antioxidants from spices might have more beneficial roles than the synthetic ones aside the advantage of being a safe alternative. Oxidative damage by free radicals has been implicated in the pathogenesis of vascular disease in diabetic complications, and several studies have revealed that antioxidants can attenuate these oxidative stressinduced changes in diabetes and hypertension (Ceriello, 2003; Vasdev et al., 2006). In an animal study carried out by Drobiova and his colleagues, garlic was reported to elevate serum antioxidant levels, decrease serum alucose in the garlic-treated diabetic rats and reduce systolic blood pressure in the garlic-treated hypertensive rats (Drobiova et al., 2010).

Inhibition of key enzymes linked to type-2 diabetes

Diabetes is one of the leading threats to worldwide public health and a major cause of global death (WHO, 1999). Reports mentioned that the number of people suffering from diabetes is about 171 million and this was projected to increase in geometric proportion to 366 million by 2030 (Wild et al., 2004). In all cases of diabetes, development of one or more complicated chronic diseases such as neuropathy, retinopathy, nephropathy and cardiomyopathy is common. There are two types of diabetes: type 1 and 2. Type 2 is more prevalent than type 1, and more than 90% of diabetes cases are that of the T2D.

T2D may be regarded as the second most common non communicable disorder, after hypertension in terms of public health significance (WHO, 1999). T2D is a metabolic disorder characterized by hyperglycemia, insulin resistance, insulin secretion and beta-cell dysfunction (WHO, 1999). There is growing scientific evidences that excess generation of highly reactive free radicals, largely due to hyperglycemia, cause oxidative stress, which further elevates the development and progression of diabetic complications (Johansen et al., 2005). Consequences of oxidative stress are damage to DNA, lipids, proteins, disruption in cellular homeostasis and accumulation of damaged molecules (Jakus, 2000). Oxidative stress is increased in diabetes because of multiple factors. These factors include glucose autooxidation, protein glycation, binding of advanced glycation end products (AGEs) to their receptors, oxidation/reduction imbalances. and reduction antioxidant defenses can lead to increased free radical production (Penckofer et al., 2002; Rahimi et al., 2005).

The use of natural antioxidants as a complementary

therapeutic approach in the management of diabetes is on the increase (Srinivasan, 2005b; Golbidi et al., 2011). Curcumin and turmeric were reported to be effective against the development of diabetic complication in rat's eyes (Suryanarayana et al., 2005), and turmeric was reported to reduce blood sugar level and modulate polyol pathway in diabetic albino rats (Arun and Nalini, 2002). In another study, curcumin, an active principle of turmeric, was reported to ameliorate diabetic nephropathy in streptozotocin-induced diabetic rats (Sharma et al., 2006). Supplementation of turmeric was also shown to attenuate proteinuria, TGF- β and IL-8 in patients with overt type 2 diabetic nephropathy and can be administered as a safe adjuvant therapy for these patients (Khajehdehi et al., 2011).

Administration of turmeric or curcumin was also reported to attenuate alloxan-induced diabetes in experimental rats (Arun and Nalini, 2002). Cinnamon is another spice that is known for its multiple health benefits. Available *in vitro*, *in vivo* and clinical evidences have indicated hypoglycaemic activity of cinnamon (Khan et al., 2003; Pham et al., 2007; Bandara et al., 2012). Although, several synthetic drugs have been developed to manage T2D but they come with their attendant side effects and are expensive. In recent times, investigations are being carried out to source natural and cheap plant foods for managing T2D and its complication through the consumption of food rich in spices, vegetables, legumes and fruits (Shim et al., 2003; Kwon et al., 2007; Ranilla et al., 2010; Oboh et al., 2010b).

Many studies have shown that inhibition of key enzymes (α- amylase and α-glucosidase) relevant to T2D could serve as therapeutic approach to the management of this disease, and some vital bioactive compounds such as polyphenols that possess interesting structure-function benefits have shown promising potentials (McCue et al., 2005; McDougall et al., 2005). α-Glucosidase and αamylase are the key enzymes involved in the digestion of carbohydrates (McCue et al., 2005; Ali et al., 2006). α-Amylase degrades complex dietary carbohydrates to oligosaccharides and disaccharides that are ultimately converted into monosaccharides by α-glucosidase (Figure 4). Liberated glucose is then absorbed by the gut and results in postprandial hyperglycemia (Kim et al., 2000; Shim et al., 2003). The inhibition of enzymes involved in the digestion of carbohydrates can significantly decrease the postprandial increase of blood glucose after a mixed carbohydrate diet by delaying the process of carbohydrate hydrolysis and absorption (Kwon et al., 2006; Oboh et al., 2010b).

The control of postprandial hyperglycemia is an important strategy in the management of diabetes mellitus, especially T2D, and reducing chronic complications associated with the disease (Kim et al., 2000; Ali et al., 2006). This is done by retarding the absorption of glucose through the inhibition of the carbohydrate-hydrolysing enzymes α -glucosidase and α -amylase in the digestive

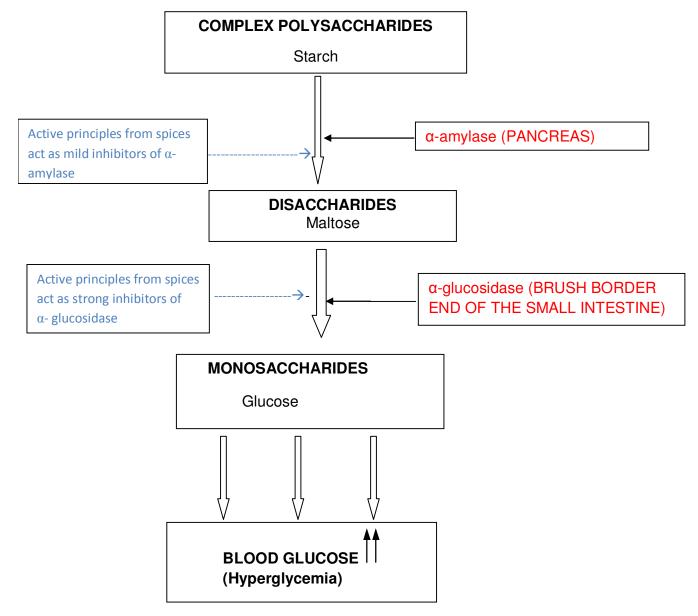


Figure 4. Schematic diagram showing the actions of α - amylase and α - glucosidase in starch digestion.

tract (Kim et al., 2000; Shim et al., 2003; Oboh et al., 2010b). Inhibitors of these enzymes could cause delay in carbohydrate digestion, prolong overall carbohydrate digestion time, causing a reduction in the rate of glucose absorption and consequently blunting the postprandial plasma glucose rise (Bhadari et al., 2008).

Several drugs such as acarbose, miglitol, voglibose, nojirimycin and 1-deoxynojirimycin have been developed and are currently in use. These drugs could act by either blocking or inhibiting these enzymes, however, they come with financial constraints and their attendant side effects, hence, alternative treatments need to be evaluated. Therefore, effective, nontoxic and cheap natural dietary inhibitors of α -amylase and α -glucosidase

with little or no side effects could be potentially promising and highly desirable. Moreover, there are strong evidences that dietary factors could be involved in the regulation and prevention of T2D (Kwon et al., 2007). Several medicinal plant and plant foods have been shown to exert their antihyperglycemic activity via inhibition of carbohydrate hydrolyzing enzymes (Ortiz-Andrade et al., 2005; McDougall et al., 2005; Cheplick et al., 2007; Oboh et al., 2010b; Pinto et al., 2010; Ranilla et al., 2010).

 α -Glucosidase and α -amylase inhibitors, which interfere with enzyme activity in the brush-border of the small intestine, could slow the liberation of D-glucose from oligosaccharide and disaccharides, resulting in delaying

glucose absorption and decreasing postprandial glucose levels. The toxicity of $\alpha\text{-glucosidase}$ and $\alpha\text{-amylase}$ inhibitors of natural source is much lower than that of the synthetic inhibitors. Kwon et al. (2007) reported that pepper varieties possess high antioxidant and good inhibitory profile on carbohydrate-degrading enzyme such as $\alpha\text{-glucosidase}$ related to glucose absorption. Green pepper and long hot pepper had little or no inhibitory effect on the $\alpha\text{-amylase}$ activity, which revealed their potential use with reduced side effects.

In our laboratory, two varieties of ginger were reported to show mild inhibition of α-amylase and strong inhibition of α-glucosidase, this suggest their potential use in nutritional intervention in the management or control of postprandial hyperglycemia associated with T2D. White ginger showed more promising attributes than red ginger (Oboh et al., 2010b). Nickavar and Yousefian (2009) investigated the inhibitory effects of six Allium spp. on αamylase activity, and four of the selected Allium spp. were reported to show appreciable α-amylase inhibition. In another study, some Cameroonian spices namely, A. daniellii, Hypodapnis zenkeri, Echinops giganteus, A. citratum, X. aethiopica and Scorodophloeus zenkeri were demonstrated to have anti-amylase and anti-lipase activities, as well as good antioxidant potentials (Etoundi et al., 2010). Cazzola et al. (2011) also reported that spices such as sage, rosemary, basil, parsley, chili, garlic and onion have interesting inhibitory activities on lipid peroxidation, α -glucosidase and α -amylase.

Furthermore, our recent report also revealed the inhibitory effects of some tropical spices: X. aethiopica [Dun.] A. (Ethiopian pepper), M. myristica (Gaertn.) Dunal (nutmeg), S. aromaticum [L.] Merr. et Perry (clove), P. guineense Schumach. et Thonn (ashanti pepper), A. danielli K. Schum (bastered melegueta) and A. melegueta (Rosc.) K. Schum (alligator pepper) on αamylase, α-glucosidase and sodium-nitroprusside (SNP)induced lipid peroxidation in pancreas (Adefegha and Oboh, 2012a). The anti-diabetic properties of the spices were attributed to the presence of biologically active phytochemicals such as phenolic constituents of the spices. Enzyme inhibition, free radical scavenging ability and prevention of lipid peroxidation may be part of the possible mechanism of action of the spices, and this might have accounted for their usage in folklore medicine as antidiabetic gents (Oboh et al., 2010b; Adefegha and Oboh, 2012a).

In a similar manner, Ranilla et al. (2010) also reported that high phenolic and antioxidant activity-linked spices (Huacatay, *Tagetes minuta* and Guascas, *Galinsoga parviflora*), and medicinal plants (Chancapiedra, *Phyllantus niruri* L. and Zarzaparrilla, *Smilax officinalis*), and herbal teas (Yerba Mate, *Ilex paraguayensis* St-Hil) in Latin America, have strong α-glucosidase inhibitory potential with no inhibition against porcine pancreatic α-amylase *in vitro*. Furthermore, Cat's claw (*Uncaria tomentosa*), cinnamon (*Cinnamomum zeylanicum* B.),

Linden tea Tilo (*Tilia platyphyllos*) and Boldo (*Peumus boldus*) were reported to strongly inhibit both α -glucosidase and α -amylase enzymes. In a related study, phenolic-rich (free and bound) extracts from clove buds also showed interesting inhibitory properties against α -glucosidase (Adefegha and Oboh, 2012b).

Ye et al. (2010) reported that some plant constituents commonly used in traditional Chinese medicine for the treatment of diabetes mellitus possess interesting inhibitory activities on α -amylase and α -glucosidase. These plant constituents are ginsenoside, puerarin, genistein, quercetin, chlorogenic dioscin. taraxasterol, kaemferol, betulinic acid and paeonol from some traditional Chinese medicinal plants, and spices for diabetes mellitus (Panax ginseng, treating notoginseng, Puerariae Iobata, Dioscorea opposite, Astragalus membranareus, Phaseolus calcaratus Gynostemma pentaphyllum, Lonicera japonica, Paeonia lactiflora. Ophiopogon iaponicas. Taraxacum mongolicum, Bupleurum chinense, Ziziphus jujuba var. spinosa) showed strong inhibitory activities on both αglucosidase and α -amylase (Ye et al., 2010).

Inhibition of angiotensin-I converting enzyme

Cardiovascular complications, characterized endothelial dysfunction and accelerated atherosclerosis, are the leading cause of morbidity and mortality associated with diabetes (Johansen et al., 2005). Hypertension is one of the commonest cardiovascular diseases which have become a global epidemic affecting 15 to 20% of all adult population (Miguel et al., 2007). Hypertension means persistent increase in blood pressure (BP). According to WHO, the normal BP for an individual should be 120/80 mmHg and if it exceeds 140/90 mmHg, it is classified as 'high BP', otherwise known as hypertension. Hyperglycemia resulting from T2D may lead to hypertension; a common cardiovascular disease.

The rennin-angiotensin system (RAS) plays a key role in the regulation of blood pressure regulation in humans (Coates, 2003). Renin produces angiotensin-I, an inactive decapeptide from angiotensinogen, after which it is cleaved by angiotensin-I converting enzyme (ACE) to release a potent vasoconstrictor angiotensin-II, an octapeptide (Je et al., 2009). ACE degrades bradykinin, a vasodilator in blood vessels, and stimulates the release of aldosterone in the adrenal cortex. The ACE activity is directly linked to hypertension, as angiotensin-II is the blood pressure regulating hormone. Increased ACE activity has been linked to narrowing of lumen of blood vessels, which results in increased blood pressure (Figure 5). Therefore, inhibition of ACE activity may provide a major anti-hypertension benefits by effectively lowering hypertension (Je et al., 2009).

Synthetic ACE inhibitors such as captopril, lisinopril,

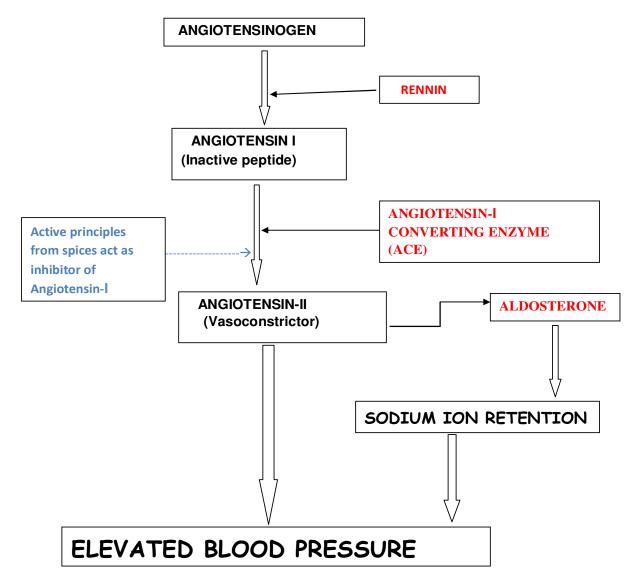


Figure 5. Schematic diagram showing the action of angiotensin-1 converting enzyme in rennin-angiotensin system.

enalapril, fosinopril and ramepril are currently in use and have shown to be very successful in controlling high blood pressure (Campos et al., 2010). They exert their antihypertensive effect by competiting for the same active site of ACE. Moreover, these drugs come with their financial constraints and their side effects such as cough. taste alterations and skin rashes. This has prompted the search for naturally-occurring ACE inhibitors, especially in flavonoids and peptides rich foods. There are indications that they are safer and lower-cost alternatives when compared to synthetic drugs (Je et al., 2009; Campos et al., 2010). Reports have shown the antihypertensive potentials and cardiovascular benefits of some medicinal plants and plant foods (Schmeda-Hirschmann et al., 1992; Hansen et al., 1995; Je et al., 2009; Pinto et al., 2009; Oboh and Ademosun, 2011; Ademiluyi and Oboh, 2012; Oboh et al., 2012c).

Ginger (Z. officinale) and red pepper (Capsicum annum) have been shown to possess high ACE inhibitory properties and could serve as dietary means of hypertension management (Ranilla et al., 2010). Clinical evidence has shown that garlic can reduce the diastolic blood pressure in hypertensive patients (McMahon and Vargas, 1993). In related study carried out to compare the cardioprotective properties of freshly crushed and processed garlic (Mukherjee et al., 2009), the authors discovered that both freshly crushed garlic and processed garlic provide cardioprotection, although the freshly crushed garlic showed a better potentials. In another clinical trial, cardamom was shown to effectively reduce blood pressure, enhance fibrinolysis and improve antioxidant status, without significantly altering blood lipids and fibrinogen levels in hypertensive patients (Verma et al., 2009).

Hypocholesteroleamic effect

Hypercholesterolemia, otherwise known as high blood cholesterol, is a major risk factor for the development of atherosclerosis and occlusive vascular disorders (Levy and Brink, 2005). WHO reported that hypercholesterolemia accounts for 18 and 56% of the world's population suffering from cerebrovascular disease and ischemic heart disease, respectively (WHO, 2002). Therapeutic life styles such as low saturated fat and cholesterol diet, weight management, and increased physical activity are vital for blood cholesterol regulation. Scientific evidences from several animal models revealed that curcumin from turmeric and capsaicin from red pepper are potent hypocholesterolaemic and hypolipidemic agents (Kempaiah and Srinivasan, 2002, 2004; Srinivasan et al.,

Commercially available drugs such as statins are presently used for blood cholesterol reduction in people with or at cardiovascular risk nowadays (Endo, 2004; Kapur et al., 2008). The drug, statin, acts by the inhibition of 3-hydroxy-3-methyl-glutaryl-CoA reductase, (HMG-CoA reductase). HMG-CoA reductase is the rate limiting enzyme that catalyzes the reduction of HMG-CoA to mevalonate and provides feedback regulation of cholesterol synthesis in cells (Brown and Goldstein, 1980). Several in vitro and in vivo studies have shown HMG-CoA reductase inhibitory effects of natural plant products. In an animal model, garlic supplemented diets was reported to reduce hepatic cholesterol synthesis by the inhibition of HMG-CoA reductase (Qureshi et al... 1987). Allicin, an organosulfur compound found in spice garlic (A. sativum) was reported to inhibit HMG-CoA reductase in both rat hepatocytes and HepG2 cells (Gebhardt et al., 1994; Gebhardt and Beck, 1996). β-Sitosterol from black cumin seeds was found to suppress hepatic HMG-CoA reductase activity in rats (Gylling and Miettinen, 2005). Curcumin from turmeric was also reported to decrease liver enzyme in cholesterol fed rats (Murugan and Pari, 2006). Quercetin, a bioflavonoid found in the skins of red onions, significantly lowers this liver enzyme in high cholesterol fed rats (Bok et al., 2002).

CONCLUSION AND RECOMMENDATION

Consumption of spice-rich foods and their ingredients could be a more effective strategy towards the management of DM and hypertension. The advantages of spice-rich food and spice antioxidant could be associated with high compliance and absence of side effects. Spice rich foods and spice antioxidant may exert their actions by possible inhibition of key enzymes linked to TY2DM (α - amylase and α -glucosidase), hypertension (ACE) and hypercholesterolaemia (HMG-CoA reductase). Spice phenolics may also serve as potential hurdles to counter the complications of diabetes arising from

oxidative dysfunction. Spices have been shown to possess good nutrient benefits with low calories, possess good inhibitory profiles on carbohydrate-modulating enzymes, ACE and HMG-CoA reductase, which correlates to their total phenolic contents, phenolic profile and antioxidant properties. This review points out the potential of spices, especially from the tropics for both T2D-linked hyperglycemia, hypertension and hypercholesterolaemia management. It also projects spice based diets as an effective dietary strategies for controlling early stages of postprandial hyperglycemia and associated hypertension. Overall, this review provides the biochemical rationale for further animal and clinical studies.

Due to the increased incidence and prevalence of several degenerative diseases such as diabetes, cardiovascular diseases including hypertension, cancer and neurodegenerative diseases including Alzheimer's diseases, concomitant drug resistance actions to these diseases and their attendant side effects, we therefore recommend an alternative dietary therapy via increased consumption of whole spice meal and spice-rich food. In a nutshell, we say "Spice up your life". This assertion supports what the great philosopher Hippocrates said about food "you are what you eat".

REFERENCES

Adefegha SA, Oboh G (2011a). Enhancement of Total phenolics and Antioxidant Properties of Some Tropical Green Leafy Vegetables by Steam Cooking. J. Food Process. Preserv. 35(5):615-622.

Adefegha SA, Oboh G (2011b). Water extractable phytochemicals from some Nigerian spices inhibit Fe2+- induced lipid peroxidation in rat's brain – in vitro. J. Food Process Technol. 2(1):1-6.

Adefegha SA, Oboh G (2012a). Inhibition of key enzymes linked to type-2 diabetes and sodium nitroprusside-induced lipid peroxidation in rat pancreas by water extractable phytochemicals from some tropical spices. Pharm. Biol. 50(7):857-865.

Adefegha SA, Oboh G (2012b). In vitro inhibition activity of polyphenolrich extracts from Syzygium aromaticum (L.) Merr. & Perry (Clove) buds against carbohydrate hydrolyzing enzymes linked to type 2 diabetes and Fe 2+- induced lipid peroxidation in rat pancreas. Asian Pac. J. Trop. Biomed. 2(10):774-781.

Adefegha SA, Oboh G (2012c). Effect of diets supplemented with Ethiopian pepper [Xylopia aethiopica (Dun.) A. Rich (Annonaceae)] and Ashanti pepper [Piper guineense Schumach. et Thonn (Piperaceae)] on some biochemical parameters in normal rats. Asian Pac. J. Trop. Biomed. S559-S567.

Ademiluyi AO, Oboh G (2012). In vitro anti-diabetes and anti-hypertension potential of phenolic extracts of selected underutilized Tropical Legumes. J. Basic Clin. Physiol. Pharmacol. 23(1):17-25. Alderman MH, Cohen H, Madhavan S (1999). Diabetes and cardiovascular events in hypertensive patients. Hypertens. 33:1130-1134.

Ali H, Houghton PJ, Soumyanath A (2006). Alpha amylase inhibitory activity of some Malaysian plants used to treat diabetes; with particular reference to *Phyllanthus amarus*. J. Ethnopharmacol. 107:449-455.

Alia M, Horcajo C, Bravo L, Goya L (2003). Effect of grape antioxidant dietary fiber on the total antioxidant capacity and the activity of liver antioxidant enzymes in rats. Nutr. Res. 23:1251-1267.

Amic D, Davidovic-Amic D, Beslo D, Trinajstic N (2003). Structureradical scavenging activity relationship of flavonoids. Croatia Chem. Acta 76:55-61.

Arun N, Nalini N (2002). Efficacy of turmeric on blood sugar and polyol pathway in diabetic albino rats. Plant Foods Hum. Nutr. 57(1):41-52.

- Bakris GL, Williams M., Dworkin L, Elliott WJ, Epstein M, Toto R., Tuttle K., Douglas J, Hsueh W, Sowers J (2000). Preserving renal function in adults with hypertension and diabetes: A consensus approach. National Kidney Foundation Hypertension and Diabetes Executive Committees Working Group. Am, J. Kidney Dis. 36(3):646-661.
- Bandara T, Uluwaduge I, Jansz ER (2012). Bioactivity of cinnamon with special emphasis on diabetes mellitus: a review. Int. J. Food Sci. Nutr. 63(3): 380 -386.
- Beauchamp C, Fridovich I (1971). Superoxide dismutase: improved assays and an assay applicable to acrylamide gels. Anal. Biochem. 44(1):276-287.
- Bhadari MR, Jong-Anarakkan N, Hong G, Kawabata J (2008). α-Glucosidase and α-amylase inhibitory activities of Nepalese medicinal herb Pakhanbhed (Bergenia ciliata, Haw). Food Chem. 106:247-252
- Bok SH, Park SY, Park YB, Lee MK, Jeon SM., Jeong TS, Choi MS (2002). Quercetin dehydrate and gallete supplements lower plasma and hepatic lipids and change activities of hepatic antioxidant enzyme in high cholesterol-fed rats. Int. J. Vitamin Nutr. Res. 72(3):161-169.
- Bravo L (1998). Polyphenols: Chemistry, dietary sources, metabolism, and nutritional significance. Nutr Rev. 56:317-333.
- Brown MS, Goldstein JL (1980). Multivalent feedback regulation of HMG CoA reductase, a control mechanism coordinating isoprenoid synthesis and cell growth. J. Lip. Res. 21:505-517.
- Cai YZ, Luo Q, Sun M, Corke H (2004). Antioxidant activity and phenolic compounds of 112 traditional Chinese medicinal plants associated with anticancer. Life Sci. 74(17):2157-2184.
- Cao G, Alessio HM, Cutler RG (1993). Oxygen-radical absorbance capacity assay for antioxidants. Free Rad. Biol. Med. 14: 303-311.
- Carlsen MH, Halvorsen BL, Holte K, Bohn SK, Dragland S, Sampson L, Willey C, Senoo H, Umezono Y, Sanada C, Barikmo I, Berhe N, Willett WC, Phillips KM, Jacobs DR, Blomhoff R (2010). The total antioxidant content of more than 3100 foods, beverages, spices, herbs and supplements used worldwide, Nutr. J. 9(3):1-11.
- Campos MRS, Guerrero LAC, Ancona DAB (2010). Angiotensin-l converting enzyme inhibitory and antioxidant activities of peptide fractions extracted by ultrafiltration of cowpea *Vigna unguiculata* hydrolysates. J. Sci. Food Agric. 90:2512-2518.
- Cazzola R, Camerotto C, Cestaro B (2011). Anti-oxidant, anti-glycant, and inhibitory activity against α-amylase and α-glucosidase of selected spices and culinary herbs. Int. J. Food Sci. Nutr. 62(2):175-184.
- Ceriello A (2003). New insights on oxidative stress and diabetic complications may lead to a causal antioxidant therapy. Diabetes Care 26(5):1589-1596.
- Cheplick S, Kwon Y, Bhowmik P, Shetty K (2007). Clonal variation in raspberry fruit phenolics and relevance for diabetes and hypertension management. J. Food Biochem. 3:656-679.
- Chu Y, Sun J, Wu X, Liu RH (2002). Antioxidant and antiproliferative activity of common vegetables. J. Agric. Food Chem. 50:6910-6916.
- Coates D (2003). The angiotensin converting enzyme (ACE). Int. J. Biochem. Cell Biol. 35:769-773.
- Cuvelier ME, Berset C, Richard H (1994). Antioxidant constituents in sage (Salvia officinalis). J. Agric. Food Chem. 1994, 42, 665-669.
- Dapkevicius A, Van Beek TA, Lelyveld GP, Van Veldhuizen, A, De Groot A, Linssen JPH (2002). Isolation and structure elucidation of radical scavengers from *Thymus vulgaris* leaves. J. Nat. Prod. 65:892–896.
- Doherty VF, Olaniran OO, Kanife UC (2010). Antimicrobial Activities of Aframomum melegueta (Alligator Pepper). Int. J. Biol. 2(2):126-131.
- Dorman HJD, Bachmayer O, Kosar M, Hiltunen R (2004). Antioxidant properties of aqueous extracts from selected Lamiaceae species grown in Turkey. J. Agric. Food Chem. 52:762-770.
- Dorman HJD, Kosar M, Kahlos K, Holm Y, Hiltunen R (2003). Antioxidant properties and composition of aqueous extracts from Mentha species, hybrids, varieties, and cultivars. J. Agric. Food Chem. 2003 51:4563–4569.
- Drobiova H, Thomson M, Al-Qattan K, Peltonen-Shalaby R, Al-Amin Z, Ali M (2011). Garlic Increases Antioxidant Levels in Diabetic and Hypertensive Rats Determined by a Modified Peroxidase Method. Evid Based Complement Alternat Med. 8 doi:10.1093.

- Endo A (2004). The discovery and development of HMG-CoA reductase inhibitors. Atherosc. Suppl. 5:67-80.
- Etoundi CB, Kuaté D, Ngondi JL, Oben J (2010). Anti-amylase, antilipase and antioxidant effects of aqueous extracts of some Cameroonian spices. J. Nat. Prod. 3:165-171
- Ezekwesili CN, Nwodo OFC, Eneh FU, Ogbunugafor HA (2010). Investigation of the chemical composition and biological activity of *Xylopia aethiopica* Dunal (Annonacae). Afr. J. Biotechnol. 9(43):7352-7356.
- Fasoyiro SB, Adegoke GO, Idowu OO (2006). Characterisation and Partial Purification of Antioxidant Component of Ethereal Fractions of *Aframomum danielli*. World J. Chem. 1(1):1-5.
- Fecka I, Turek S (2008). Determination of polyphenolic compounds in commercial herbal drugs and spices from Lamiaceae: thyme, wild thyme and sweet marjoram by chromatographic techniques. Food Chem. 108:1039-1053.
- Gebhardt R, Beck H, Wagner KG (1994). Inhibition of cholesterol biosynthesis by allicin and ajoene in rat hepatocytes and HepG2 cells. Biochim. Biophys. Acta 1213: 57-62.
- Gebhardt R, Beck H (1996). Differential inhibitory effects of garlicderived organosulfur compounds on cholesterol biosynthesis in primary rat hepatocyte culture. Lipids 31:1269-1276.
- Golbidi S, Ebadi SA, Laher I (2011). Antioxidants in the treatment of diabetes. Curr. Diabetes Rev. 7(2):106-125.
- Gyamfi MA, Yonamine M and Aniya Y (1999). Free-radical scavenging action of medicinal herbs from Ghana: Thonningia sanguinea on experimentally-induced liver injuries. Gen. Pharmacol. 32:661-667.
- Gylling H, Miettinen TA (2005). The effect of plant stanol- and sterolenriched foods on lipid metabolism, serum lipids and coronary heart disease. Ann. Clin. Biochem. 42:254-263.
- Hansen K, Nyman U, Smitt UW, Adsersen A, Gudiksen L, Rajasekharan S, Pushpangadan P (1995). In vitro screening of traditional medicines for anti-hypertensive effect based on inhibition of the angiotensin converting enzyme (ACE). J. Ethnopharmacol. 48(1995):43-51.
- Halliwell B, Murcia MA, Chirico S, Aruoma OI (1995). Free radicals and antioxidants in food and in vivo: what they do and how they work. Crit. Rev. Food Sci. Nutr. 35(1-2):7-20.
- Halliwell B (1997). Antioxidants and human diseases: a general introduction. Nutr. Rev. 55:S44–S52.
- Halliwel B, Gutteridge JMC (1999). Free radicals in biology and medicine (3rd Ed.). Oxford: Oxford University Press.
- Halliwell B (2006). Reactive species and antioxidants. Redox biology is a fundamental theme of aerobic life. Plant Physiol. 141:312- 322.
- Haraguchi H, Saito, T, Ishikawa H, Date H, Kataoka S, Tamura Y (1996). Antiperoxidative components in *Thymus vulgaris*. Plant Med. 62: 217–221.
- Hiyasat B, Sabha D, Grotzinger K, Kempfert J, Rauwald JW, Mohr FW, Dhein S (2009). Antiplatelet activity of Allium ursinum and Allium sativum. Pharmacol. 83:197-204.
- Hossain MB, Brunton NP, Barry-Ryan C, Martin-Diana AB, Wilkinson M (2008). Antioxidant activity of spice extracts and phenolics in comparison to synthetic antioxidants. Rasayan J. Chem. 1 (4): 751-756
- Hossain M, Dilip K, Brunton N, Martin-Diana A, Barry-Ryan C (2010). Characterization of Phenolic Composition in Lamiaceae Spices by LC-ESI-MS/MS. J. Agric. Food Chem. 58(19):10576-10581.
- Jakus V (2000). The role of free radicals, oxidative stress and antioxidant systems in diabetic vascular disease. Bratisl Lek Listy.101(10): 541-551.
- Je JY, Park PJ, Kim EK, Ahn CB (2009). Antioxidant and angiotensin I converting enzyme inhibitory activityof Bambusae caulis in Liquamen. Food Chem.13: 932-935.
- Johansen JS, Harris AK, Rychly DJ, Ergul A (2005). Oxidative stress and the use of antioxidants in diabetes: linking basic science to clinical practice. Cardiovasc. Diabetol. 4(1):5-15.
- Kapur NK, Ashen D, Blumenthal RS (2008). High density lipoprotein cholesterol: an evolving target of therapy in the management of cardiovascular disease. J. Vasc. Health Risk Man. 4:39-57.
- Kehrer JP (1993). Free radicals as mediators of tissue injury and disease. Crit. Rev. Toxicol. 23:21-48.
- Kempaiah RK, Srinivasan K (2002). Integrity of erythrocytes of

- hypercholesterolemic rats during spices treatment. Mol. Cell. Biochem. 236:155-161.
- Kempaiah RK, Srinivasan K (2004). Influence of dietary spices on the fluidity of erythrocytes in hypercholesterolemic rats. Br. J. Nutr. 93:81-92.
- Kennedy DO, Wightman EL (2011). Herbal Extracts and Phytochemicals: Plant Secondary Metabolites and the Enhancement of Human Brain Function. Adv. Nutr. 2:32-50.
- Khajehdehi P, Pakfetrat M, Javidnia K, Azad F, Malekmakan L, Nasab MH, Dehghanzadeh G. (2011). Oral supplementation of turmeric attenuates proteinuria, transforming growth factor-β and interleukin-8 levels in patients with overt type 2 diabetic nephropathy: a randomized, double-blind and placebo-controlled study. Scand J. Urol. Nephrol. 45(5): 365 370.
- Khan A, Safdar M, Ali KMM, Khattak KN, Anderson RA (2003). Cinnamon improves glucose and lipids of people with type 2 diabetes. Diabetes Care 26(12):3215-3218.
- Kim JS, Kwon CS, Son KH (2000). Inhibition of alpha-glucosidase and amylase by luteolin, a flavonoid. Biosci. Biotechnol. Biochem. 64: 2458-2461.
- Konczak I, Zabaras D, Dunstan M, Aguas P (2010). Antioxidant capacity and phenolic compounds in commercially grown native Australian herbs and spices, Food Chem. 122(1):260-266.
- Kwon YI, Jang HD, Shetty K (2006). Evaluation of Rhodiola crenulata and Rhodiola rosea for management of type II diabetes and hypertension. Asian Pac. J. Clin. Nutr. 15:425-432.
- Kwon YI, Apostolidis E, Kim YC, Shetty K (2007). Health benefits of traditional corn, beans and pumpkin: In vitro studies for hyperglycemia and hypertension management. J. Med. Food 10:266-275.
- Lawson LD (1998). Garlic, a review of its medicinal effects and indicated active compounds. In: Chemistry and Biological Activity, Series 691: Phytomedicines of Europe (Lawson LD, Bauer R, eds.). American Chemical Society, Washington, DC, 176–209.
- Lissi E, Pascual C, Castillo MD (1992). Luminol luminescence induced by 2,2'-azo-bis (2-amidinopropane) thermolysis Free Rod. Res. Commun.17:299-311.
- Lissi E, Sahm-Hanna M, Pascual C, Del Castillo MD (1995). Evaluation of total antioxidant potential (TRAP) and total antioxidant reactivity from luminol-enhanced chemiluminescence measurements. Free Rad. Bwl. Med. 2:153-158.
- Liu RH (2004). Potential synergy of phytochemicals in cancer prevention: mechanism of action. J. Nutr. 134:3479S–3485S
- Liu H, Qiu N, Ding H, Yao R (2008). Polyphenols contents and antioxidant capacity of 68 Chinese herbals suitable for medical or food uses, Food Res. Int. 41(4):363-370.
- Lu M, Yuan B, Zeng M, Chen J (2011). Antioxidant capacity and major phenolic compounds of spices commonly consumed in China. Food Res. Int. 44:530–536.
- Lyckander IM, Malterud KE (1992). Lipophilic flavonoids from Orthosiphon spicatus as inhibitors of 15-lipoxygenase. Acta Pharm. Nordica 4:159-166.
- Masude T, Isibe J, Jitoe A, Naramati N (1992). Antioxidant curcuminoids from rhizomes of Curcuma zanthorrhiza. Phytochem. 33:3645-3647.
- McCue P, Kwon YI, Shetty K (2005). Anti-diabetic and anti-hypertensive potential of sprouted and solid-state bioprocessed soybean. Asian Pac. J. Clin. Nutr. 14:145-152
- McDougall GJ, Shpiro F, Dobson P, Smith P, Blake A, Stewart D (2005): Different polyphenolic components of soft fruits inhibit α-amylase and α-glucosidase. J. Agric. Food Chem. 53:2760-2766.
- McMahon FG, Vargas R (1993). Can garlic lower blood pressure? A pilot study. Pharmacother. 13(4):406 407.
- Miguel M, Alonso M, Salaices M, Aleixandre A, L'opez-Fandi R (2007). Antihypertensive, ACE inhibitory and vasodilator properties of an egg white hydrolysate: effect of a simulated intestinal digestion. Food Chem. 104:163-168.
- Mitsuda H, Yuasumoto K, Iwami K (1996). Antioxidation action of indole compounds during the autoxidation of linoleic acid. Eiyo to Shokuryo: 19:210.
- Miura K, Kikuzaki H, Nakatani N (2002). Antioxidant activity of chemical components from sage (*Salvia officinalis* L.) and thyme (*Thymus*

- vulgaris L.) measured by the oil stability index method. J. Agric. Food Chem. 50(7):1845-1851.
- Muchuweti M, Kativu E, Mupure CH, Chidewe C, Ndhlala AR, Benhura MAN (2007). Phenolic composition and antioxidant properties of some spices. Am. J. Food Technol. 2 (5):414-420.
- Mukherjee S, Lekli I, Goswami S, Das DK (2009). Freshly crushed garlic is a superior cardioprotective agent than processed garlic. J. Agric. Food Chem. 57(15):7137-7144.
- Nickavar B, Yousefian N (2009). Inhibitory effects of six Allium species on α-amylase enzyme activity. Iranian J. Pharm. Res. 8:53-57.
- Ninfali P, Mea G, Giorgini S, Rocchi M, Bacchiocca M (2005). Antioxidant capacity of vegetables, spices, and dressings relevant to nutrition. Br. J. Nutr. 93:257-266.
- Oboh G, Ekperigin MM, Akindahunsi AA (2005). Responses of Rat Liver Enzymes and Metabolites to Diet Supplemented with Some Nigerian Spices. Nig. J. Biochem. Mol. Biol. 20(2):81-87.
- Oboh G (2006): Antioxidant properties of some commonly consumed & underutilized tropical legumes. Eur. Food Res. Technol. 224:61-65.
- Oboh G, Rocha JBT (2007). Antioxidant in Foods: A New Challenge for Food processors. Leading Edge Antioxidants Research, Nova Science Publishers Inc. New York US, 35- 64
- Oboh G, Raddatz H, Henle T (2008). Antioxidant properties of polar and non-polar extracts of some tropical green leafy vegetables. J. Sci. Food Agric. 88:2486-2492.
- Oboh G, Adefegha SA, Rocha JBT (2010a). Fe2+ and sodium nitroprusside-induced oxidative stress in rat's brain (in vitro): Protective Effects of Aqueous Extract of *Mentha viridis* and *Majarona hortensis* Leaves. Adv. Food Sci. 32(1):11-19.
- Oboh G, Akinyemi JA, Ademiluyi AO, Adefegha SA (2010b). Inhibitory effect of Aqueous extract of two varieties of Ginger on Key Enzymes linked with Type-2 Diabetes. J. Food Nutr. Res. 49(1):14-20.
- Oboh G, Ogunruku OO (2010). Cyclophosphamide induced oxidative stress in brain: Protective effect of Hot short pepper (*Capsicum frutescens* L Var. abbreviatum). Exp. Toxicol. Pathol. 62:227-233.
- Oboh G, Akomolafe TL, Adefegha SA, Adetuyi AO (2011). Inhibition of cyclophosphamide induced oxidative stress in brain by polar and non-polar extracts of Annatto (*Bixa orellana*) seeds. Exp. Toxicol. Pathol. 63(3):257-262.
- Oboh G, Ademosun AO (2011). Shaddock Peels (Citrus maxima) Phenolic Extracts Inhibit α-amylase, α--glucosidase and Angiotensin-I Converting Enzyme Activities: A Nutraceutical Approach to Diabetes Management. Diab. Met. Synd. Clin. Res. Rev. 5(3):148-152.
- Oboh G, Akinyemi AJ, Ademiluyi AO (2012a). Antioxidant and inhibitory effect of red ginger (Zingiber officinale var. Rubra) and white ginger (Zingiber officinale Roscoe) on Fe2+ induced lipid peroxidation in rat brain in vitro: Exp. Toxicol. Pathol. 64:31-36.
- Oboh G, Akomolafe TL, Adefegha SA, Adetuyi AO (2012b). Attenuation of cyclophosphamide-induced neurotoxicity in rat by yellow dye extract from root of Brimstone tree (*Morinda lucida*). Exp. Toxicol. Pathol. 64 (6):591-596.
- Oboh G, Ademiluyi AO, Akinyemi AJ, Saliu JA, Henle T, Schwarzenbolz U (2012c). Inhibitory effect of polyphenol-rich extracts of Jute leaf (Corchorus olitorius) on key enzyme linked to type-2 diabetes (α-amylase and α-glucosidase) and hypertension (Angiotensin I converting) *in vitro*. J. Funct. Foods 4(2):450-458.
- Odukoya AO, Ilori OO, Sofidiya MO, Aniunoh OA, Lawal BM, Tade IO (2005). Antioxidant activity of Nigerian dietary spices. Electr. J. Environ. Agric. Food Chem. 4(6):1086-1093.
- Ohkawa H, Ohishi N, Yagi K (1979). Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. Anal. Biochem. 95:351-358.
- Olonisakin Á, Oladimeji MO, Lajide L (2006). Chemical composition and antibacterial activity of steam distilled oil of Ashanti Pepper (Piper guineense) fruits (berries). Electr. J. Environ. Agric. Food Chem. 5(5):1531-1535.
- Ortiz-Andrade RR, Garcia-Jimenez S, Castillo-Espana P, Rameriz-Vila G, Villalobos-Molina R, Estrada-Soto S (2007). α-glucodidase inhibitory activity of the methanolic extract from Tourneforia hartwegiana: an anti-hyperglycemic agent. J. Ethnopharmacol. 109:48-53.
- Osawa T, Namiki N (1981). A novel type of antioxidant isolated from leaf wax of Eucalpyptus leaves. Agric. Biol. Chem. 45:735-739.
- Oyaizu M (1986). Studies on product of browning reaction prepared

- from glucosamine, Jpn. J. Nutr. 44:307-315.
- Pellegrini N, Serafini M, Salvatore S, Del Rio D, Bianchi M, Brighenti F (2006). Total antioxidant capacity of spices, dried fruits, nuts, pulses, cereals and sweets consumed in Italy assessed by three different in vitro assays. Mol. Nutr. Food Res. 50(11):1030-1038.
- Penckofer S, Schwertz D, Florczak K (2002). Oxidative stress and cardiovascular disease in type 2 diabetes: the role of antioxidants and prooxidants. J. Cardiovasc. Nurs. 16(2):68-85.
- Pham AQ, Kourlas H, Pham DQ (2007). Cinnamon supplementation in patients with type 2 diabetes mellitus. Pharmacother. 27(4):595-599.
- Pietta PG (2000). Flavonoids as antioxidants. J. Nat. Prod. 63:1035-1042.
- Pinto MD, Ranilla LG, Apostolidis E, Lajolo FM, Genovese MI, Shetty K (2009). Evaluation of antihyperglycemia and antihypertension potential of native peruvian fruits using *in vitro* models. J. Med. Food 12:278-291.
- Pradeep KU, Geervani P, Eggum BO (1993). Common Indian spices: Nutrient composition, consumption and contribution to dietary value. Pl. Foods for Human Nutr. 44:137-148.
- Pratt DE (1980). Natural antioxidants of soybean and other oil-seeds. In Simic MG, Karel M (Eds.), Autoxidation in food and biological systems (pp. 283–292). New York: Plenum Press.
- Qureshi AA, Crenshaw TD, Abuirmeileh N, Peterson DM, Elson CE (1987). Influence of minor plant constituents on porcine hepatic lipid metabolism: impact on serum lipid. Atheroscl. 64:109-115.
- Rahimi R, Nikfar S, Larijani B, Abdollahi M (2005). A review on the role of antioxidants in the management of diabetes and its complications. Biomed. Pharmacother. 59:365-373.
- Ranilla LG, Kwon YI, Apostolidis E, Shetty K (2010). Phenolic compounds, antioxidant activity and in vitro inhibitory potential against key enzymes relevant for hyperglycemia and hypertension of commonly used medicinal plants, herbs and spices in Latin America. Bioresour. Technol. 101(12):4676- 4689.
- Re R, Pellegrini N, Proteggente A, Pannala A, Yang M, Rice-Evans C (1999). Antioxidant activity applying an improved ABTS radical cation decolorization assay, Free Rad. Biol. Med. 26(9–10):1231-1237.
- Rice-Evans C, Miller NJ, Paganga G (1996). Structure—antioxidant activity relationships of flavonoids and phenolic acids. Free Rad. Biol. Med. 20:933-956.
- Ruch RJ, Cheng SJ, Klaunig JE (1989). Prevention of cytotoxicity and inhibition of intracellular communication by antioxidant catechins isolated from Chinese green tea. Carcinog. 10(6):1003-1008
- Scalbert A, Williamson G (2000). Dietary intake and bioavailability of polyphenols. J. Nutr. 130(8S):2073S-2085S.
- Scalbert A, Manach C, Morand C, Remesy C (2005). Dietary polyphenols and the prevention of diseases. Crit. Rev. Food Sci. Nutr. 45:287-306.
- Schmeda-Hirschmann G, Loyola JI, Sierra J, Retamal R, Rodriguez J (1992). Hypotensive Effect and Enzyme Inhibition Activity of Mapuche Medicinal Plant Extracts. Phytother. Res. 6:184-188.
- Shahidi F, Ho CT (2000). Phytochemicals and phytopharmaceuticals. Champaign, IL: ACS Press.
- Shahidi F, Naczk M (2004). Phenolics in food and nutraceuticals. Boca Raton, FL: CRC Press.
- Shan B, Cai YZ, Sun M and Corke H (2005). Antioxidant capacity of 26 spice extracts and characterization of their phenolic constituents. J. Agric. Food Chem. 53(20):7749-7759.
 - Sharma S, Kulkarni SK, Chopra K (2006). Curcumin, the active principle of turmeric (Curcuma longa), ameliorates diabetic nephropathy in rats. Clin. Exp. Pharmacol. Physiol. 33(10):940-945.
- Sherwin ER (1990). Antioxidants. In: Food additives (Branen AL, Davidson PM, Salminen S ed) pp.139-193.Marcek Dekker Inc, New York, USA.
- Shim YJ, Doo HK, Ahn SY, Kim YS, Seong JK, Park IS, Min BH (2003). Inhibitory effect of aqueous extract from the gall of *Rhus chinensis* on alpha-glucosidase activity and postprandial blood glucose. J. Ethnopharmacol. 85:283-287.
- Shobana S, Naidu KA (2000). Antioxidant activity of selected Indian spices. Prostaglandins, Leukotrienes and Essential Fatty Acids. Sol Genomics Network 62(2):107-110
- Sofowora LA (1993). Medicinal plants and traditional medicine in Africa. Spectrum Books Ltd, Ibaban. pp. 55-71.

- Sowers JR, Epstein M (1995). Diabetes mellitus and associated hypertension, vascular disease, and nephropathy. An update. Hypertens. 26:869–879.
- Srinivasan K, Sambaiah K, Chandrasekhara N (2004). Spices as beneficial hypolipidemic food adjuncts: A Review. Food Rev. Int. 20:187-220.
- Srinivasan K (2005a). Role of Spices beyond food flavoring: nutraceuticals with multiple health effects. Food Rev. Int. 21:167-188.
- Srinivasan K (2005b). Plant foods in the management of diabetes mellitus: Spices as beneficial antidiabetic food adjuncts. Int. J. Food Sci. Nutr.56 (6):399-414.
- Suryanarayana P, Saraswat M, Mrudula T, Krishna TP, Krishnaswamy K, Reddy GB (2005). Curcumin and turmeric delay streptozotocin-induced diabetic cataract in rats. Invest. Ophthalmol. Vis. Sci. 46(6):2092-2099.
- Stalikas CD (2007). Extraction, separation, and detection methods for phenolic acids and flavonoids. J. Sep. Sci. 30(18):3268-3295.
- Stankevičius M, Akuņeca I, Jākobsone I, Maruška A (2010). Analysis of phenolic compounds and radical scavenging activities of spice plants extracts. maisto chemija ir technologija. 44(2):85-91.
- Suhaj M (2006). Spice antioxidants isolation and their antiradical activity. J. Food Comp. Anal. 19:531-537
- Sun J, Chu Y, Wu X, Liu R (2002). Antioxidant and antiproliferative activities of common fruits. J. Agric. Food Chem. 50:7449-7454.
- Ursell A (2000). The Complete Guide to Healing Foods, pp. 112–114. London: Dorling Kindersley Ltd.
- Uwakwe AA, Nwaoguikpe RN (2008). In vitro antisickling effects of Xylopia aethiopica and Monodora myristica. J. Med. Plants Res. 2(6):119-124.
- Vasdev S, Gill VD, Singal PK (2006). Modulation of oxidative stressinduced changes in hypertension and atherosclerosis by antioxidants. Exp. Clin. Cardiol. 11(3):206 - 216.
- Verma SK, Jain V, Katewa SS (2009). Blood pressure lowering, fibrinolysis enhancing and antioxidant activities of cardamom (Elettaria cardamomum). Indian J. Biochem. Biophys. 46(6):503-506
- Visioli F, Galli C (1998). Olive oil phenols and their potential effects on human health. J. Agric. Food. Chem. 46:4292–4296.
- Viuda-Martos M, Rúiz-Navajas Y, Fernández-López J, Pérez-Álvarez JA (2011). Spices as functional foods. Crit. Rev. Food Sci. Food Saf. 51:13-28.
- Watanabe J, Shinmoto H, Tsushida, T (2005). Coumarin and favone derivatives from estragon and thyme as inhibitors of chemical mediator release from RBL-2H3 cells. Biosci. Biotechnol. Biochem. 69(1):1–6.
- Wojdylo A, Oszmianski J, Czemerys R (2007). Antioxidant activity and phenolic compounds in 32 selected herbs. Food Chem. 105(3):940-
- World health organization consultation (1999). Definition, diagnoses and classification of diabetes mellitus and its complication. Part 1: diagnosis and classification of diabetes mellitus. Report of a WHO consultation, Geneva.
- Ye X, Song C, Yuan P, Mao R (2010). α-Glucosidase and α-amylase inhibitory activity of common constituents from traditional Chinese medicine used for diabetes mellitus. Chin. J. Nat. Med. 8:0349-0352.
- Zafra-Stone S, Yasmin T, Bagchi M, Chatterjee A, Vinson JA, Bagchi D, Berry B (2007). Anthocyanins as novel antioxidants in human health and disease prevention. Mol. Nutr. Food Res. 51(6):675-683.
- Zheng W, Wang SY (2001). Antioxidant activity and phenolic compounds in selected herbs. J. Agric. Food Chem. 49(11):5165-5170.