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Full Length Research Paper

Genetic variability, correlation and path analysis for quantitative traits of seed yield, and yield components in chickpea (*Cicer arietinum* L.) at Maichew, Northern Ethiopia

Assefa Amare Hagos¹*, Tadesse Desalegn² and Tesfay Belay³

¹Ethiopian Institute of Agricultural Research, Mehoni Agricultural Research Center, Maichew, Tigray, Ethiopia. ²Department of Plant Science, College of Agriculture and Environmental Science, Bahir Dar University, Bahir Dar, Ethiopia.

³Tigray Agricultural Research Institute, Mekelle Agricultural Research Center, Mekelle, Ethiopia.

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Twelve chickpea genotypes were tested to assess variability, heritability, correlations and direct and indirect effects between yield and yield components. Maximum phenotypic and genotypic coefficient of variation was recorded for number of seeds per plant (33.8, 32.4), number of secondary branches per plant (30.3, 29.6), number of pods per plant (25.6, 24.7) and 100 seed weight (23.0, 22.7) respectively. High heritability coupled with high expected genetic advance as percent of mean were estimated for number of secondary branches per plant, number of pods per plant and 100 seed. Path coefficient analysis (seed yield as a dependent variable) revealed that seeds per plant followed by biomass yield, days to maturity and 100 seed weight had exerted positive direct effect on seed yield. To conclude, number of seeds per plant, biomass yield, 100 seed weight and days to maturity are important parameters for selecting maximum yielding genotypes in chickpea.

Key words: Chickpea, genetic variability, path coefficient, heritability, correlation, genetic advance.

INTRODUCTION

Chickpea ranks third among pulses, and it accounts for 12% of the world pulses production (Khan and Khan, 2011). In Ethiopia it accounts for about 14.31% (third) of the acreage and 17.28% (second) of the total production of all grain legumes grown in the country. Area of production has been increasing greatly in recent years. In the 2011 main (MEHER) season, about 232,000 ha of

cultivated land is used for the production of 400,200 tons of chickpea (CSA, 2012). Chickpea, a multi-functional crop, has an important role in the diet of the Ethiopian small scale farmers' households and also serves as protein source for the rural poor who cannot afford to buy animal products. The crop also serves as a source of cash income and plays a major role in Ethiopia's foreign

*Corresponding author. E-mail: assefaamare69@yahoo.com. Tel: +251-914-297477. Fax: +251347770021.

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exchange earnings through export to Asia and Europe. Despite its nutritional values and economic importance, the average yield of chickpea is relatively low in the country. This is primarily due to poor genetic makeup of the cultivars available, excessive vegetative growth, low tolerance to diseases and non-availability of grains of improved varieties which need immediate attention of the breeders for the evolution of maximum yielding varieties which fulfill the requirements of ever increasing population.

Genetic variability is a prerequisite for any breeding program, which provides opportunity to a plant breeder for selection of high yielding genotypes. However information on the association between yield and its various components provide the basis for the selection of improved varieties (Saleem et al., 2005). Information on the relative magnitude of the different sources of variation particularity among different genotypes for several traits helps in measurement of their range of genetic diversity and may provide evidence for identification of their relationship. The variability of a biological population is an outcome of genetic constitution of the individuals and its interaction with the prevailing environment. A survey of genetic variability with the help of suitable parameters such as genetic coefficient of variation, heritability estimates and genetic advance are absolutely necessary to start an efficient breeding program. Some of the characters are highly associated among themselves and with seed yield. The analysis of the relationships among these characters and their associations with seed yield is essential to establish selection criteria (Atta et al., 2008). Progress in any breeding program depends upon the nature and magnitude of variability present in the base population. Assessment of the extent of genetic variability within chickpea is fundamental for chickpea breeding (Qureshi et al., 2004).

Chickpea breeders should consider heritability estimates along with genetic advance because heritability alone is not a good indicator of the amount of usable genetic variability (Noor et al., 2003). The concept of heritability explains whether the differences observed among individuals arose as a result of differences in genetic makeup or due to environmental forces. Genetic advance gives an idea of possible improvement of new population through selection, when compared to the original population. The genetic gain depends upon the amount of genetic variability and magnitude of the masking effect of the environment. Information of the genetic variability, heritability and association of various characters provides a basis to the plant breeders to breed the chickpea genotypes possessing higher yield potential. Selection on the basis of grain yield, a polygenic character, is usually not very efficient, but selection based on its component characters could be more efficient. The present study was initiated with the prime objective of finding the mutual relationships of different quantitative traits and the type and extent of their

contribution to grain yield.

MATERIALS AND METHODS

Description of the experimental site

The field experiment was conducted in Maichew Agricultural College which is located at 39°32′E and 12°47′N in the Tigray National Regional State, Ethiopia. Maichew is found 123 km far from mekelle the capital city of Tigray region and 662 km north of Addis Ababa the capital city of Ethiopia. It is located at 2396 m above sea level and receives an average annual rain fall of 758.7 mm and annual mean temperature of 16.4°C.

Experimental materials and procedures

Twelve chickpea genotypes obtained from the High Land Pulse Research Program of Debre Zeit Agricultural Research Center (Table 1) were planted in a randomized complete block design with three replications. Each plot consisted of 4 lines of 4 m length by 1.2 m width (4.8 m²). The plant-to-plant and row-to-row distance was maintained at 10 and 30 cm, respectively. Agronomic practices were carried out as per recommendation.

Data collection

The following data were collected from the experiment both per plot and per plant basis.

Data recorded on plot basis

- a) Days to 50% flowering (DF)
- b) Days to 90% maturity (DM)
- c) Grain filling period (GFP)
- d) Hundred Seed weight (HSW)
- e) Biomass yield (Biological yield) (BY)
- f) Seed yield per hectare (SY)

Data recorded on plant basis

The data for the following characters were recorded from five randomly taken plants from each plot and the average value was considered per plant basis.

- a) Plant height (PH)
- b) Number of Primary Branches per Plant (NPB)
- c) Number of Secondary Branches per Plant (NSB)
- d) Number of Pods per Plant (PPt)
- e) Number of Seeds per Pod (SPo)
- F) Number of Seeds per Plant (SPt)

Statistical analysis

The phenotypic, genotypic and environmental variances and coefficient of variation is defined according to the formula suggested by Singh and Chaudhary (1985) as follows:

Environmental variance $(\sigma^2 e) = MS_e$

$$Genotypic\ Variance(\sigma^2g) = \left[\frac{MS_g - MS_e}{r}\right]$$

S/N	Variety	Year of release	Crosses/seed source	Seed color	Туре
1	DZ-10-11	1974	Collection	Light Brown	Desi
2	Dubie	1978	Collection	Grey	Desi
3	Mariye	1985	K850xF378(sel fromICCx730089)	Brown	Desi
4	Wroku	1994	(Annigeri x Chaffa) x (Rabat xF378)	Golden	Desi
5	Akaki	1995	P99 xNEC 108) x Radhey	Golden	Desi
6	Mastewal	2006	NA	Golden	Desi
7	Naatolii	2007	(ICCV88102 x ICCV10) x ICC4958	Light Green	Desi
8	shasho	1999/2000	ICCC33x(L144xE100Y(M)	White	Kabuli
9	cheffe	2004	(ICCV2xsurutato 77)xICC7344	White	Kabuli
10	Habru	2004	X85TH230/ILC3395xFlip83-13C	White	Kabuli
11	Ejeri	2005	X94TH71/FLIP87-59CxUC15	White	Kabuli
12	Teji	2005	X94TH75/FLIP87-58CxUC15	White	Kabuli

Table 1. List of genotypes considered in the study.

Phenotypic variance(σp^2) = $\sigma g^2 + \sigma e^2$

Phenotypic Coefficient of variation (PCV) = $\frac{\sigma p}{\bar{x}}x100$

Genotype coefficient of variation (GCV) = $\frac{\sigma g}{\bar{x}} x 100$

Where, \bar{x} = grand mean of character.

Broad sense heritability (H) expressed as the percentage of the ratio of the genotypic variance to the phenotypic variance will be computed on genotype mean basis as described by Allard (1960) as:

$$H = \left[\frac{\sigma_g^2}{\sigma_p^2} \right] x 100$$

Genetic advance in absolute percent of the mean (GAM), assuming selection of superior 5% of the genotypes will be estimated in accordance with the methods illustrated by Johnson et al. (1955).

$$GA = \frac{K\sigma^2 P h^2}{h^2}$$

GMA =
$$(GA)^{x}/x$$
 100

Phenotypic and genotypic correlation coefficients were estimated using the standard procedure suggested by Miller et al. (1958) from corresponding variance and covariance:

$$phenotypic\ correlation\ coefficient\ \left(r_{P_{xy}}\right) = \frac{\sigma_{F_{xy}}}{\left(\sqrt{\sigma^2 P_x * \sigma^2 P_y}\right)}$$

Genotypic correlation coefficient
$$\left(r_{g_{xy}}\right) = \frac{\sigma_{g_{xy}}}{\left(\sqrt{\sigma^2 g_{x^*} \sigma^2 g_y}\right)}$$

$$t = \frac{r}{\sqrt{\frac{1 - r^2}{n - 2}}}$$
 where, n number of genotypes

$$t = \frac{rg_{xy}}{SE_{rg_{xy}}} \quad where, SE_{rg_{xy}} = \sqrt{\frac{1 - r^2 g_{xy}}{2h^2_x * h^2_y}}$$

Phenotypic correlation coefficient was tested for their significance using the formula suggested by Sharma (1998).

$$t = \frac{r}{\sqrt{\frac{1 - r^2}{n - 2}}}$$
 where, n number of genotypes

Genotypic correlation coefficient was tested with the following formula suggested by Robertson (1959):

$$t = rac{rg_{xy}}{SE_{rg_{xy}}}$$
 where, $SE_{rg_{xy}} = \sqrt{rac{1 - r^2 g_{xy}}{2h^2_x * h^2_y}}$

 $SErg_{xy} = Standard$ error of genotypic correlation coefficient between character X and Y

 h^2x = heritability for character x and h^2y = heritability for character y. The calculated absolute t value was tested against the tabulated t-value at g-2 degree of freedom for both phenotypic and genotypic correlations.

Path coefficient analysis was estimated as suggested by Dewey and Lu (1959) using the phenotypic as well as genotypic correlation coefficients to determine the direct and indirect effects of yield components on seed yield based on the following relationship:

$$r_{ij} = P_{ij} + \sum r_{ik} P_{kj}$$

Where, $\mathbf{rij} = \mathbf{Mutual}$ association between the independent character (i) and dependent character, grain yield (j) as measured by the correlation coefficients. $\mathbf{Pij} = \mathbf{Components}$ of direct effects of the independent character (i) as measured by the path coefficients and $\mathbf{\Sigma} \ \mathbf{r_{ik}} \ \mathbf{p_{kj}} = \mathbf{summation}$ of components of indirect of a given independent character (i) on a given dependent character (j) via all other independent characters (k). The contribution of the remaining unknown factor was measured as the residual factor (PR), which is calculated as:

17.55

64.15

25.90

46.10

31.30

Trait	GV	PV	EV	PCV (%)	GCV (%)	H (%)	GA	GA (%)
DF	47.78	47.30	0.47	14.64	14.56	99	14.10	29.85
DM	11.74	10.20	1.53	2.98	2.78	87	6.13	5.34
GFP	46.18	45.28	0.90	10.06	9.96	98	13.74	20.32
PH	17.49	15.63	1.86	13.84	13.08	89	7.69	25.47
NPB	0.05	0.01	0.04	8.62	2.84	11	0.10	1.93
NSB	5.30	5.05	0.25	30.28	29.56	95	4.52	59.43
PPt	97.39	90.12	7.28	25.64	24.67	92	18.62	48.88

12.00

33.80

14.15

23.03

16.0

10.11

32.44

13.34

22.70

16.00

71

92

87

97

90

0.21

29.09

879.31

13.02

631.01

0.01

18.50

25819.15

1.20

8.73

Table 2. Genetic parameters of yield and yield components in chickpea.

0.01

216.59

205172.36

41.20

104073.23

DF = Days to 50% flowering, DM = Days to maturity, GFP = Grain filling period, PH = Plant height (cm), number of primary branches per plant, NSB = Number of secondary branches per plant, PPt = Number of pods per plant, SPo= Number of seeds per pod, SPt = Number of seeds per plant, BY = Biomass yield (kg/ha), SW = 100 seed weight (g), SY= Seed yield (kg/ha), SE=Standard error, GV= Genotypic Variance, PV=phenotypic Variances, EV=Environmental Variance, PCV= phenotypic of variability, GCV= genotypic coefficient of variability, H=broad sense heritability, GA= expected genetic advance GA% =genetic advance as percent of the mean GA%.

$$p_r = \sqrt{\left(1 - \sum r_{ij} P_{ij}\right)}$$

SPo

SPt

BY

SW

SY

0.02

235.09

230991.52

42.29

115361.95

The magnitude of PR indicates how best the causal factors account for the variability of the dependent factor (Singh and Chaudhary, 1999).

RESULTS AND DISCUSSION

It is clear from the Table 2 that The highest estimates for phenotypic coefficients of variation were recorded for number of seeds per plant (33.80), number of secondary branches per plant (30.28), number of pods per plant (25.64) and 100 seed weight (23.03). The higher phenotypic coefficients of variation values for number of pods per plant and 100 seed weight were in agreement with previous reports (Sharma and Saini, 2010). The highest genetic coefficients of variation were observed for number of seeds per plant (32.44), number of secondary branches per plant (29.56), number of pods per plant (24.67) and 100 seed weight (22.70). Similar results were reported (Sharma and Saini, 2010) who found high GCV values for secondary branches per plant, pods per plant and seeds per plant in chickpea genotypes. heritability estimate was high (>80%) for days to 50% flowering, grain filling period, 100 seed weight, number of secondary branches per plant, number of seeds per plant, number of pods per plant, seed yield, plant height, days to maturity and biomass yield. High heritability values for 100-seed weight, number of pods per plant, seed yield per plant, number of branches per plant and plant height were in accordance with previous reports by Sharma and Saini (2010).

Genetic advance as percent of mean at 5% selection intensity was high for number of seeds per pod (64.2%) followed by number of secondary branches per plant (59.4%), number of pods per plant (48.9%) and 100 seed weight (46.1%). Ali et al. (2011) found higher values of genetic advance for number of pods per plant, plant height and grain filling period. The present study revealed that high heritability coupled with high expected genetic advance as percent of mean for number of secondary branches per plant, number of pods per plant and 100 seed weight. Therefore, these characters could be improved more easily than other characters measured in this study. Genotypic and phenotypic correlations among the characters are shown in Table 3. Seed yield showed positive and significant phenotypic association with biomass yield (0.75) and plant height (0.59) Therefore, any improvement of these characters would result a substantial increment in seed yield. Similar reports were observed by Vaghela et al. (2009), Malik et al. (2010) and Kobraee et al. (2010). The correlation coefficients of seed yield with hundred seed weight were positive at genotypic level and negative at phenotypic level. Biomass yield had significant positive genotypic and phenotypic correlation with seed yield. Similar results have been reported by Ali et al. (2011). Positive genotypic correlations of biomass yield with plant height (0.53), 100 seed weight (0.44) and number of primary branches per plant (0.32) have also been observed. Hundred seed weight had positive genotypic and phenotypic correlation with plant height. It had negative and significant genotypic and phenotypic correlation with number of secondary branches per plant (r_{a=}0.83, r_{ph=} 0.81), seeds per pod ($r_{q=}0.87$, $r_{ph}=0.73$), pods per plant $(r_{g=}0.73, r_{ph=}0.73)$ and seeds per plant $(r_{g=}0.84, r_{ph=}0.79)$.

Table 3. Genotypic (above diagonal) and phenotypic (below diagonal) correlation coefficients among 12 characters.

Variables	DF	DM	GFP	PH	NPB	NSB	PPt	SPo	SPt	BY	SW	SY
DF		0.295	-0.894**	-0.286	-0.454	0.190	0.223	0.137	0.195	0.159	-0.013	0.443
DM	0.299		0.163	-0.362	-0.710**	0.212	0.381	0.610*	0.569*	0.215	-0.497	0.217
GFP	-0.881**	0.186		0.132	0.104	-0.098	0.047	0.150	0.071	-0.073	-0.224	-0.358
PH	-0.263	-0.267	0.146		0.444	-0.727**	0.253	-0.322	-0.285	0.523	0.607*	0.600*
NPB	-0.169	-0.326	0.003	0.104		0.208	0.115	0.283	-0.074	0.316	0.111	0.237
NSB	0.180	0.212	-0.081	-0.641*	0.049		0.704*	0.594	0.667*	-0.538	-0.833**	-0.370
PPt	0.213	0.344	0.038	0.177	0.002	0.675**		0.618*	0.933**	-0.372	-0.727**	0.137
SPo	0.115	0.488	0.125	-0.251	0.020	0.484	0.494*		0.848*	-0.165	-0.868**	-0.032
SPt	0.184	0.523*	0.079	-0.209	-0.040	0.640*	0.911**	0.786**		-0.323	-0.836*	0.090
BY	0.148	0.214	-0.048	0.550*	0.045	-0.471	-0.275	-0.110	-0.231		0.465	0.766**
SW	0.408	-0.478	-0.226	0.582*	0.090	-0.805**	0.676**	-0.734**	-0.790**	0.444		0.335
SY	-0.014	0.156	-0.341	0.587*	0.099	-0.316	0.180	0.034	0.143	0.748**	-0.338	

^{*, **} Indicate significance at the 0.05 and 0.01 probability levels, respectively. DF = Days to 50% flowering, DM = Days to maturity, GFP = Grain filling period, PH = Plant height (cm), number of primary branches per plant, NSB = Number of secondary branches per plant, PPt = Number of pods per plant, SPo= Number of seeds per pod, SPt = Number of seeds per plant, BY = Biomass vield (kg/ha), SW = 100 seed weight (g), SY= Seed vield (kg/ha).

Negative association between 100 seed weight indicates a compensatory relationship between them. Pods per plant had positive and significant genotypic and phenotypic correlation with number of secondary branches per plant ($r_{\alpha}=0.70$, $r_{\text{ph}=}$ 0.68) and seeds per plant ($r_{a=}0.93$, $r_{ph=}0.91$). A positive and significant genotypic and phenotypic correlation of number of pods per plant with number of secondary branches per plant agrees with the findings of Ali et al. (2011). Positive and significant genotypic and phenotypic correlation of seeds per plant with number of secondary branches per plant (r_{a=}0.67, r_{ph=} 0.64), number of pods per plant ($r_{q=}0.93$, $r_{ph=}0.91$) and seeds per pod $(r_{q=}0.82, r_{ph=}0.78)$ has been observed. Seeds per pod had significant positive genotypic and phenotypic correlation with seeds per plant $(r_{0}=0.85, r_{0}=0.79)$. Positive and significant correlation of number of secondary branches per plant with number of pods per plant (r_{a=}0.70, r_{bh=} 0.68) and number of seeds per plant was

observed at genotypic and phenotypic level. The positive and significant correlation of number of secondary branches per plant with number of pods per plant agrees with the findings of Malik et al. (2010). Plant height had positive and significant genotypic and phenotypic correlation with 100 seed weight ($r_{a=}0.61$, $r_{ph=}0.58$) and seed yield $(r_{\alpha}=0.60, r_{\text{ph}}=0.59)$, Plant height had positive genotypic correlation with biological yield and number of primary branches per plant. This is in line with the study by Ali et al. (2011) who found positive and non-significant genotypic correlation of plant height with number of primary branches per plant. Generally, positive and significant association of pairs of characters at phenotypic level and positive and high correlation at genotypic level justified the possibility of correlated response to select. The negative correlations prohibit the simultaneous improvement of those traits. Thus, correlation analysis indicated that biomass yield and plant height were found to be important yield components and these traits can be used for yield improvement in chickpea (Table 3).

Seeds per plant followed by biomass yield, days to maturity and 100 seed weight had exerted positive direct effect on seed vield. Deb and Khaleque (2005), Yucel et al. (2006) and Zali et al. (2011) reported similar results for seeds per plant. However, days to 50% flowering, grain filling period, number of secondary branches per plant, number of pods per plant, seeds per pod, plant height and number of primary branches per plant showed negative direct effect on seed yield. The high positive direct effect of 100 seed weight on seed yield was counterbalanced by its indirect effect via seeds per plant which finally resulted in positive and low genotypic correlation with seed yield. The residual (0.0315) indicates that characters which are included in the genotypic path analysis explained 96.85% of the total variation in seed yields (Table 4).

Table 4. Estimates of direct (bold diagonal) and indirect effect (off diagonal) for 12 characters.

Variables	DF	DM	GFP	PH	NPB	NSB	PPt	SPo	SPt	BY	SW	rg
DF	-1.56613	0.12957	1.58404	0.01542	0.01231	-0.07272	0.0494	-0.02597	0.18214	0.13742	-0.0029	0.44
DM	-0.46273	-0.43854	0.28917	0.01948	0.01925	-0.08130	0.0845	-0.11574	0.53241	0.18610	-0.1145	0.22
GFP	1.40073	0.07160	-1.77109	-0.00711	-0.00282	0.03761	0.0105	-0.02838	0.06652	-0.06351	-0.0515	-0.36
PH	0.44859	-0.15865	-0.23398	-0.05384	-0.01204	0.27841	0.0561	0.06119	-0.26688	0.45355	0.1397	0.60*
NPB	0.71160	-0.31147	-0.18415	0.02391	-0.02710	-0.07948	0.0254	-0.05371	-0.06917	0.27385	0.0255	0.24
NSB	0.29758	0.09316	0.17405	0.03917	-0.00563	-0.38271	0.1562	-0.11274	0.62416	-0.46645	-0.1918	-0.37
PPt	-0.34889	0.16724	0.08403	0.01363	0.00311	-0.26960	0.2217	-0.11722	0.87247	-0.32212	-0.1674	0.14
SPo	-0.21437	0.26748	-0.26484	0.01736	-0.00767	-0.22738	0.1369	-0.18976	0.79298	-0.14330	-0.1998	-0.03
SPt	0.30499	0.24964	-0.12596	0.01536	0.00200	-0.25540	0.2068	-0.16089	0.93528	-0.27975	-0.1925	0.09
BY	-0.24830	0.09416	0.12978	-0.02817	-0.00856	0.20596	0.0824	0.03137	-0.30186	0.86676	0.1070	0.77**
SW	0.01962	-0.21814	0.39627	0.03268	0.00300	0.31884	0.1612	0.16474	-0.78225	0.40297	0.2301	0.33

Residual Effect =0.032.*, ** Indicate significance at the 0.05 and 0.01probability levels, respectively . DF = Days to 50% flowering, DM = Days to maturity, GFP = Grain filling period, PH = Plant height (cm), number of primary branches per plant, NSB = Number of seeds per plant, BY = Biomass yield (kg/ha), SW = 100 seed weight (g), SY = Seed yield (kg/ha).

Conclusion

On the basis of these results it was suggested that pods per plant, primary branched per plant, secondary branches per plant and 100 seed weight may be given more importance while making selection for higher yield potential in chickpea.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

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