

## Short Communication

# The influence of sterilizing compounds on the yield of viable explants of *Rhododendron* L. (Ericaceae)

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**Results are presented on the influence of sterilizing compounds upon the yield of viable explants of *Rhododendron* in sterilized culture. Yield of viable explants is dependent upon type of sterilizing compound, the type of specie the plant belongs and the type of explant. Results show, that 0.1% solution of silver nitrate is the most effective compound for sterilization of seeds of 8 *Rhododendron* species (sterilization for 5 min) and 0.1% solution of sublimate and diacid (sterilization for 8 min) are most effective for sterilization of buds of 4 *Rhododendron* species.**

**Key words:** Sterilizing compounds, seeds, buds, *Rhododendrons*.

## INTRODUCTION

The process of clonal micropropagation consists of explant isolation, sterilization and planting on nutritious medium. Cacas and Lasa (1986) investigated efficiency of 5 sterilizing compounds on yield of explants of sugar beet. Chloride of mercury is the most effective concentration 1% for 1 min. Kudina and Dovbysh (1990) in the sterilization of buds of rose sorts offered to use 70% solution of ethanol for 2 min and 10% solution of hydrogen peroxide for 15 min. Ruskauskas et al. (1989) note, that the best means for sterilization of orchid were next compounds: 70% solution of ethanol (sterilization for 2 min) and 0.1% solution of diacid (sterilization for 5 min) and 10% solution of chloramine (sterilization for 10 min). Sudhadevi and Nataraja (1987) in the sterilization of explants *Dalbergia latifolia* Roxb offered to use chloride of mercury. More effective compounds for sterilization of explants of tea (*Camellia sinensis* (L.) Kuntze) were 1 to 2% solution of hydrogen peroxide and 50 to 60% solution of ethanol (the first sterilization; 10 to 15 s) (Tvardkiladze and Mezentzev, 1987).

At the second sterilization, 0.05 to 0.2% solution of diacid was apply for 5 to 10 min. Balakrishnamurthy and Rangasamy (1988) offer to sterilize floral apex of banana by 70% solution of ethanol for 30 s, after that with 0.1% solution of sublimate for 5 min with the next washing in sterilized water.

We divide sterilizing compounds into some groups:

1. Compounds, possessed by strong disinfecting action.
2. Compounds, possessed by middle disinfecting action.
3. Compounds, possessed by weak action.

Compounds, which contain mercury (sublimate, diacid, nitric acid mercury), nitric acid silver, belong to the first group. Compounds, which contain active chlorine, sodium and potassium hypochloride, chloramine, chloride of lime, belong to the second group. Hydrogen peroxide, potassium permanganate with their oxidizing properties belongs to the third group. Chloramine and hydrogen peroxide possess by weak toxic action owing to their fast decomposition. We use these substances for sterilization of tender tissues. Combinations, which contain mercury are used in the case of uneffective action of solutions with chlorine. Chlorine active combinations (chloride of lime, chloramine) are traditional means for sterilization.

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Mechanism of destruction of microorganisms with the help of free chlorine is not cleared. Probable ways of chlorine infection are related to suppression of some important ferment reactions in microbe cell, denaturation of proteins and nucleic acids (Dychdala, 1983). Preparations, which contain oxygen (for example hydrogen peroxide) are strong oxidants, base of action of which is formation of free radicals, which injure lipid of cell membranes, DNA and another important components of microbe cell.

In spite of the synthesis of catalase by microorganisms, which protect cells from been affected by hydrogen peroxide by way of decomposition into water and oxygen, H<sub>2</sub>O<sub>2</sub> used in sterilization concentrations is allowed to overcome the present mechanism of resistance (Turner 1983). However, hydrogen peroxide has positive and negative properties.

Hydrogen peroxide in high concentrations has a wide spectrum of activity, ability to dissolve many biological combinations, has no odour, fast decomposition into non-toxic products in environment. Hydrogen peroxide has negative properties such as: high tissue toxicity, which is developed in destruction of plant pigments, which leads to tissues losing their colour. So, it is necessary to use it with care.

From group of spirits, ethyl alcohol and isopropyl alcohol are widely used in disinfection. Mechanism of their action consists of denaturation of microbe proteins (Larson, 1991).

It is necessary to note, that for every species of plants, optimum regime of sterilization, which promotes high yield of viable explants, is determined by experimental ways. Thus, according to the data of Achmedova (1999), from tested concentrations of various sterilizing compounds (nitrate of silver, chloramine, hydrogen peroxide) only 0.1% solution of nitrate of silver ensured the high yield of viable explants of sugar-beet. Analogous investigations were carried out with the plants of black currants (Atroschenko at al., 1990), aconite (Melnichuk at al., 2004), dahlia (Shumichin, 2004), barley (Rokityanskaya, 2005) and other plants.

Attention should be paid on the data of Japanese scientists Kiyosue and Kamada (1989) about investigations of infection of various sterilizing compounds and their concentrations under conditions of disinfected treatment of carrot seeds.

It was an interesting fact, that the use of potassium hypochloride in 5% concentration, calcium hypochloride in 6% concentration, sodium hypochloride in 10% concentration subsequently stimulated differentiation of somatical germs of carrot. In a case of application of calcium hypochloride solution we can reveal positive correlation between duration of treatment and frequency of formation of somatical germs.

Unfortunately in the literature there were no revealed data of investigations, according to influence of sterilizing compounds on the yield of viable explants that introduced

plants of *Rhododendron* species. For every plant optimum regime of sterilization, high yield of viable explants is determined by experimental ways. In this connection we carried out experimental investigations as for this question.

## MATERIALS AND METHODS

Objects of investigation were 12 introduced species of *Rhododendron*: *Rhododendron catawbiense* Michaux, *Rhododendron ponticum* L., *Rhododendron smirnowii* Trautv., *Rhododendron japonicum* (A.Gray) Suring, *Rhododendron brachycarpum* D.Don, (syn. *Azalea brachycarpa* D.Don), *Rhododendron kotschyi* Simonk, *Rhododendron haemaleum* Balf. f. and Forrest, *Rhododendron minus* Michaux, *Rhododendron discolor* Franch, *Rhododendron roseum* (Loisel.) Rehd., *Rhododendron fortunei* Lind., *Rhododendron schlippenbachii* Maxim.

For these 12 *Rhododendron* species we tested next sterilizing compounds: 0.1% solutions of diacid, sublimate and silver nitrate in combination with the treatment of 70% ethanol. The time of sterilization with ethanol was 5 s, with diacid and sublimate - 8 min, with silver nitrate - 5 min. We investigated and used explants buds and seeds of *Rhododendron* species in culture *in vitro*. For 4 species of *Rhododendron* (*R. japonicum*, *R. catawbiense*, *R. smirnowii*, *R. ponticum*) used top and lateral buds of young shoots as explants; for 8 *Rhododendron* species (*R. fortunei*, *R. minus*, *R. kotschyi*, *R. schlippenbachii*, *R. discolor*, *R. brachycarpum*, *R. roseum*, *R. haemaleum*) we used seeds as explants.

After sterilization we washed thoroughly plant material three times in sterilized bidistillate water for 15 min, after that this plant material was transferred on nutrient agar Andersen's medium (1975), which contain inorganic salts, vitamins, 3% (w/v) sucrose and 0.8% Difco bactoagar. The level of pH of the medium was to 4.8 before autoclaving at 1.06 kg/cm<sup>2</sup> pressure for 20 min at 121°C. 15 ml of this medium was used in a 25 × 150 mm test tube. Test tubes with transplanted explants put on the shelves, where temperature of air was 24 ± 2°C, illumination - 4000 lk, relative humidity of air was 70%, photoperiod - 16 h. Registration of infested, oxidized and viable explants was conducted daily, during 2 weeks. Experimental data are presented in Table 1.

## RESULTS, DISCUSSION AND CONCLUSION

Figures in Table 1 testify to high yield (100%) of viable seeds of investigated species of *Rhododendron* independently of type of sterilizing compound, with the exception of two species *R. minus* (85%) and *R. kotschyi* (80%), which had small sizes of seeds (0.4 × 0.1 mm).

Yield of viable buds depends on type of sterilizing compound and species belonging to the plants. We marked highest yield of viable buds of *R. japonicum*. This index is lower as for *R. catawbiense* (85%), *R. ponticum* (90%) and *R. smirnowii* (95%). It is connected with belonging of *R. japonicum* to deciduous shrubs and of another species of *Rhododendron* to evergreen shrubs. Buds of deciduous *R. japonicum* are less infected, because they were isolated from shoots, grown under conditions of glasshouse. Unfortunately, shoots of evergreen *Rhododendrons* are incapable of growth under these conditions, so their buds are more infected.

**Table 1.** Viability of explants of introduced species of *Rhododendron* depending on sterilizing compounds.

Species	Plant	Concentration of solution of sterilizing compound (% v/v)								
		Silver nitrate, 0.1			Diacid, 0.1			Sublimate, 0.1		
		Time of exposition (min)								
		5			8			8		
I	O	V	I	O	V	I	O	V		
<i>R. catawbiense</i>	Buds	0/0	3/15	17/85	0/0	0/0	20/100	0/0	0/0	20/100
<i>R. ponticum</i>	Buds	0/0	2/10	18/90	0/0	3/15	17/85	0/0	0/0	20/100
<i>R. smirnowii</i>	Buds	0/0	1/5	19/95	0/0	0/0	20/100	0/0	0/0	20/100
<i>R. japonicum</i>	Buds	0/0	0/0	20/100	0/0	0/0	20/100	0/0	0/0	20/100
<i>R. brachycarpum</i>	Seeds	0/0	0/0	20/100	0/0	0/0	20/100	0/0	0/0	20/100
<i>R. kotschy</i>	Seeds	0/0	4/20	16/80	0/0	0/0	20/100	0/0	0/0	20/100
<i>R. haemaleum</i>	Seeds	0/0	0/0	20/100	0/0	0/0	20/100	0/0	0/0	20/100
<i>R. minus</i>	Seeds	0/0	3/15	17/85	0/0	0/0	20/100	0/0	0/0	20/100
<i>R. discolor</i>	Seeds	0/0	0/0	20/100	0/0	0/0	20/100	0/0	0/0	20/100
<i>R. roseum</i>	Seeds	0/0	0/0	20/100	0/0	0/0	20/100	0/0	0/0	20/100
<i>R. fortunei</i>	Seeds	0/0	0/0	20/100	0/0	0/0	20/100	0/0	0/0	20/100
<i>R. schlippenbachii</i>	Seeds	0/0	0/0	20/100	0/0	0/0	20/100	0/0	0/0	20/100

Abbreviation: I - infected, O - oxidized, V - viable explants; Quantity of explants is in numerator (pieces); in denominator is %. Annotation: Calculation was carried out issued from 20 explants for every species.

At the base of our investigations we drew a conclusion, that yield of viable explants depends on type of sterilizing compound, species of plant and also on type of explant. 0.1% solution of silver nitrate is a most effective compound for sterilization of seeds of 8 *Rhododendron* species (sterilization for 5 min) and 0.1% solution of sublimate and diacid (sterilization for 8 min) are most effective for sterilization of buds of 4 *Rhododendron* species.

## REFERENCES

- Achmedova JV (1999). Introduction of tissues of sugar beet into culture in vitro with the aim of microclonal propagation. In: Problems of theoretical and applied genetics in Kazakhstan: Materials of republic conference. Alma-Ata. 18-22 of November: (in Russian), pp. 117-118.
- Anderson WC (1975). Propagation of rhododendrons by tissue culture. Part 1. Development of culture medium for multiplication of shoots. Combined Proceedings of the International Plant Propagator's Society, 25: 1929-1935.
- Atroschenko G, Guseva GG, Cherepanova MA (1990). Clonal micropropagation of black currants. In: Materials of conference of the young scientists and students. Leningrad Agricultural institute. Leningrad: (in Russian), pp. 43-44.
- Balakrishnamurthy G, Sree R (1988). Regeneration of Banana plantlet in vitro culture of floral apices. Curr. Sci. (India), 57(5): 270-272.
- Cacas A, Lasa J (1986). Multiplication in vitro en renirolacha azucarera (*Beta vulgaris* L.) Tipo de explante y sistema de sterilization. Estac. exp. Aula Dei., 18: 51-56.
- Dychdala GR (1983). Chlorine and chlorine compounds. In: Block S.S. (Eds.). Disinfection, sterilization and preservation. 3rd ed. Philadelphia: Lea and Febiger, pp. 157-182.
- Kiyosue T, Kamada H, Harada H (1989). Induction of Somatic Embryogenesis from Carrot Seeds by Hypochlorite Treatment. Plant Tissue Cult. Lett., 6(3): 138-143.
- Kudina GA, Dovbysh NP (1990). Microclonal propagation of hybrid rose. Industrial botany: Perspectives of evaluation: Proceedings of scientific conference of Donetsk botanical garden of HASU, 1990, Kyiv: (in Russian), pp.193-194.
- Larson EL (1991). Alcohols. In: Block S.S. (Eds.). Disinfection, sterilization and preservation. 3rd ed. Philadelphia: Lea and Febiger, pp. 191-203.
- Melnichuk MD, Pinchuk AP, Maurer VM (2004). Processes of callusformation and organogenesis of hybrids of poplar in culture in vitro. Biol. Biotechnol., (in Ukraine), 5(1): 25-30.
- Raskauskas V, Kazlauskienė R, Gudaviciene N, Sirvydyte D (1989). Sterilization materials and culture media selection method meristeminiu reproduced orchids. Scientific transactions of institutes of higher education of Lithuania. (Bulletin of biological sciences) (in Lithuanian), pp. 27: 36-42.
- Rokityanskaya LS (2005). Search of effective methods of sterilization of ripe germs of barley. In: Biology is a science of XXI century. Puschino: (in Russian), p. 190.
- Shumichin SA (2004). Optimization of separate stages of microclonal propagation of dahlia cultural: sterilization of explants. Bulletin of Permskogo university, (in Russian), 2: 61-63.
- Sudhadevi AN, Nataraja K (1987). Establishment of plantlets in hypocotyl cultures of *Dalbergia latifolia* Roxb. Indian J. For., 10(1): 1-6.
- Turner FJ (1983). Hydrogen peroxide and other oxidant disinfectants In: Block SS (Eds.). Disinfection, sterilization and preservation. 3rd ed. Philadelphia: Lea and Febiger, pp. 240-250.
- Tvartkiladze OK, Mezencev AV (1987). Method of propagation of tea in vitro: a.s. 1311673 USSR, MKI A01NZ/OD/ - N 3868024/ 30-15; Declaration. 10.01.1985. Published 12.01.1987. Inventions Bulletin. (in Russian), 19: 15.