

Full length Research Paper

Genetic polymorphism of growth hormone locus and its association with bodyweight in Grati dairy cows

Sucik Maylinda

Faculty of Animal Husbandry, Brawijaya University, Malang – Indonesia. E-mail: sucikmaylinda@yahoo.com

Accepted 29 June, 2011

The aim of this research was to study genetic polymorphism in growth hormone (GH) locus and its association with body weight of 43 Grati dairy cows ranging from 2 to 4 years old. Polymorphism of GH locus was ascribed using a PCR-RFLP method involving restricted enzyme *Msp1* on 2% agarose gel. All data were subjected to statistical analysis based on one way classification model using a statistical software package of Genstat 200 version 2. The results showed that frequencies of normal allele (*Msp1*⁺) and mutant allele (*Msp*⁻) were 0.34 and 0.66 respectively with 0.22 polymorphism was found in this locus. There was significant relationship between *Msp1*⁺/*Msp1*⁺, *Msp1*⁺/*Msp1*⁻ and *Msp1*⁻/*Msp1*⁻ genotypes and body weight ($P < 0.05$) but *Msp1*⁺/*Msp1*⁺ and *Msp1*⁺/*Msp1*⁻ genotypes had a stronger correlation to the higher body weight than *Msp1*⁻/*Msp1*⁻ genotype.

Key words: Grati cows, polymorphism, growth hormone, body weight.

INTRODUCTION

Grati cattle (GC) is regarded as indigenous breed that was a result of crossing between Friesian Holstein (FH) and Ongole crossbred cattle in Pasuruan regency of East Java since 19th century (Huitema, 1986). They have been adapted to relatively harsh environment especially to hot and humid climate and low-quality feed to produce milk and power to plough a farm land prior to planting. The cow yields milk between 2,500 to 4,500 kg/lactation which is far below the potential of FH dairy cows. Nevertheless, GC plays a significant role in the income generation for smallholder dairy farms in the region. Recently, selection for better performance of such important indigenous breed has received more attention especially with the advancement of genetically molecular biotechnology. Cheong et al. (2006) and (Kish, 2008) stated that growth hormone (GH) plays a vital role in post-natal growth and general metabolism including for lactation. Thus it is not surprising if GH has been the most intensive object of studies in ruminant animals to associate mutation of GH with the productive traits (Lara et al., 2002; Zhou et al., 2005; Khatami et al., 2005; Ferraz et al., 2006; Pawar et al., 2007; Mouzavisadeh et al., 2009). Yardibi et al. (2009) reported a significant effect of GH hormone gene polymorphism on milk yield of Eastern and South Anatolian Red breed cows. More

recently, Komisarek et al. (2011) carried out a study on 209 dairy cows in Poland and found a significant decrease in milk production upon GHR-F279Y and shorter calving interval upon GHL127V polymorphism, respectively. Growth is a complex trait, which is controlled by multiple genes.

During the peak lactation milk is often synthesised at the expense of tissue catabolism, especially when the intake of feed energy is inadequate and a decline in body weight results. Under field conditions smallholder dairy farmers usually use body weight for selection criterion due to the lack of proper milk recording. This study aims at investigation of GH locus polymorphism in Grati cows and its association with body weight that potentially can be used as a marker assisted selection.

MATERIALS AND METHODS

Animals

43 Grati dairy cows that were kept at a dairy stall belonging to Suka Makmur Agricultural Farming Cooperative at Grati district, Pasuruan regency of East Java were observed during 2004 to 2005. The cows aged between 27 and 30 months old and varied between 1 and 3 lactation periods. All cows received a diet consisting of elephant grass (*Pennisetum purpureum*) and local concentrates.

Table 1. Band pattern of the fragment after being digested by Msp1 enzyme.

Length of DNA of particular band (bp)	Identified as allele	Genotype
224 105	Normal allele (Msp1 ⁺ *)	Msp1+ /Msp1+
329	Mutant allele (Msp1 ⁻ **)	Msp1+ /Msp1-
224 105 329	(Msp1 ⁺) and (Msp1 ⁻)	Msp1- /Msp1-

Cut by enzyme ; **) uncut by enzyme.

The bodyweight was estimated from the chest girth (CG) prior to blood collection using a School formula:

$$BW_{kg} = (CG_{cm} + 22)^2 / 100 \text{ (Sudono et al., 2003)}$$

The reason for estimating body weight by chest girth measurement was poorly due to the unavailability of weighing scale in the site of study, and based on the many reports that chest girth is the best predictor for body weight (ILRI, 2011)

Genotyping for GH

Approximately 10 ml of blood was taken from jugular vein of each cow using venojet spuit and mixed with EDTA 10% in the Falcon tube and stored at -25°C. Following the genomic DNA isolation, the cows were genotyped for GH locus using PCR-RFLP (polymerase chain reaction-restriction fragment length polymorphism) and 2% agarose gel electrophoresis (Sulandari and Zein, 2003). Amplification of fragment of 329 bp at intron 3' (Dybus, 2002) was done with PCR using primer F: 5'- CCCACGGGCAAGAATGAGGC - 3' dan R:5' - TGAGGAAGTGCAGGGGCCCA - 3'. To digest this fragment, a protocol of RFLP (restricted fragment length polymorphism) with restriction enzyme Msp1 was used to recognize the particular site of CC↓GG. The protocol for PCR analysis is stated in Table 1. PCR protocols to amp lily that fragment were by predenaturation at 94°C for 5 min for 1 cycle, denaturation at 94°C for 40 s, annealing at 60°C for 40 s and elongation at 72°C for 30 s for 30 cycles (Dybus, 2002).

Alleles identification

Following the end of PCR and RFLP processes, the products were then subsequently electrophorated using 2% agarose gel to identify polymorphism of alleles based on the length of the band (Table 1).

Statistical analysis

One-way analysis of variance was carried out to compare polymorphism level using a statistical software Genstat 2000 version 2. Polymorphism level was calculated using formula of Budak et al. (2003) that is:

$$PIC_i = 1 - \sum p_{ij}^2$$

where PIC_i is polymorphic information content at i-th locus, p_{ij} is the frequency of j-th allele and i-th locus.

RESULTS AND DISCUSSION

Growth hormone gene polymorphism

The number cows that met the criteria for this study was only 43. This mainly due to the large variation in farm size and husbandry practise between smallholder dairy farms in the village under study. Figure 1 indicates the number of dairy cows showing polymorphism at GH locus bearing Msp1⁺/Msp1⁺, Msp1⁺/Msp1⁻ and Msp1⁻/Msp1⁻ genotype were 7, 15 and 21, respectively (Table 2). It was found that the frequency of Msp1⁺ and Msp1⁻ alleles was 0.34 and 0.66. The level of polymorphism at that locus was 0.22 indicating that Grati dairy cows are polymorphic as has been generally accepted that the minimum value of polymorphism is 1% (Hyperdictionary, 2000; Dorak, 2006). This finding suggests that in Grati dairy cows there was high variability in the GH locus and offers opportunity to use GH genotype for selection criteria. From the previous study of Jakaria et al. (2007) on 132 indigenous cattle breed (Pesisir cattle) in West Sumatra, the Msp1-GH gene showed significant relationship with bodyweight as estimated by chest girth measurement. Based on the Chi Square test (Table 2), it was found that alleles frequency were under genetic equilibrium.

Similar findings were reported in dairy cattle by Pawar et al. (2007) that polymorphism occurred in exon 5 of GH1 gene. Allelic frequency in genetic equilibrium means allele frequency will never change resulting in alleles and genotypic frequency stability.

Relationship of GH locus polymorphism with body weight

Table 3 shows the significant association (p < 0.05) between genotypes and body weight. However, the Msp1⁻/Msp1⁻ genotype gave less influence on body weight than those of Msp1⁺/Msp1⁺ and Msp1⁺/Msp1⁻ genotypes. Another locus in growth factor such as somatomedine or IGF1 gene has also been reported plays an important role in growth performance of beef

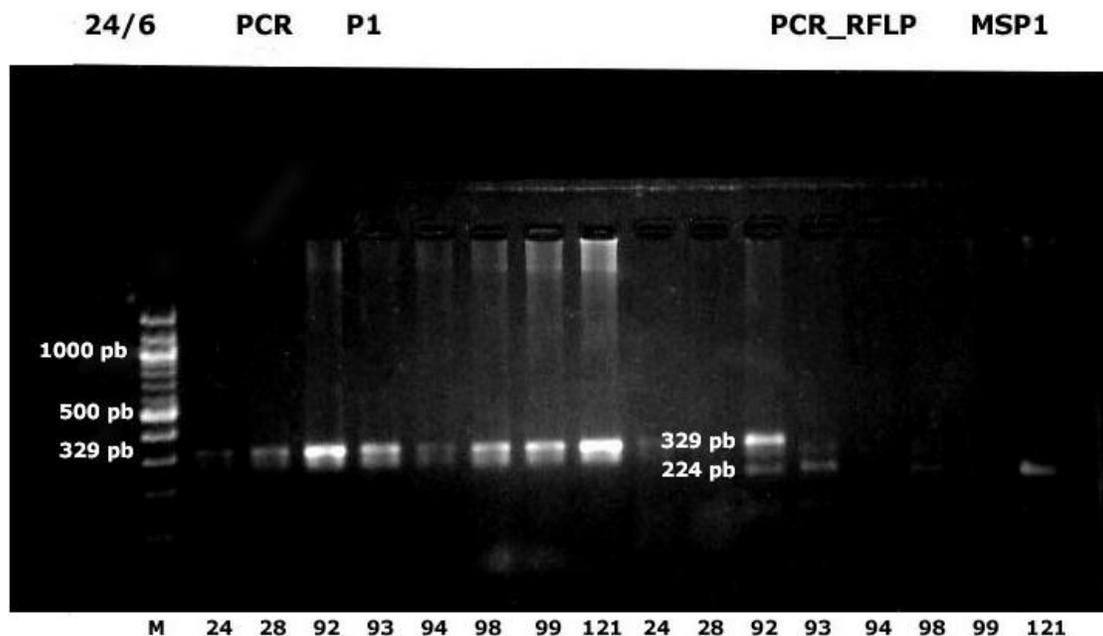


Figure 1. Electrophoresis result of PCR-RFLP products.

Table 2. Frequency of alleles Msp1+ and Msp1 at GH1 locus.

Allele frequency	Genotypic frequency
(Msp1 ⁺) = 0.34 *	Msp1+ /Msp1+ = 7
(Msp1 ⁻) = 0.66 *	Msp1+ /Msp1- = 15
	Msp1- /Msp1- = 21

*) The values denote that population were in genetic equilibrium based on the Chi square test ($p < 0.05$).

Table 3. PCR-RFLP product and mean of body weight of dairy cows in each genotype.

Genotype	N (**)	Mean of body weight (kg)
Msp1+ /Msp1+	7	448.8 ± 48.60 ^b
Msp1+ /Msp1-	15	447.7 ± 45.77 ^b
Msp1- /Msp1-	21	408.8 ± 50.83 ^a
Total	43	428.9 ± 51.58

*) Body weight was estimated using School formula, **) number of dairy cow. Values bearing unlike superscript in the same column differ significantly ($p < 0.05$).

cattle (Reyna et al., 2010). This notion supports the previous findings in other species of animals that GH gene can be promoted as candidate gene to improve animal performance such as body weight and carcass weight in poultry (Thakur et al., 2006) and Angus cattle (Zhao et al., 2004). As has been known that the growth of animals is under the hormonal control of GH, growth hormone receptor (GHR) and insulin-like growth factor I (IGF-I). Polymorphism that occurs in the regulatory region

(for example promoter region) and coding region (exons) of the genes responsible for those three hormones will influence the expression of the genes and the function of protein during the translation process (Gee et al., 1999). There is strong indication that in all animals the level of blood GH reflects the GH genotype. Current knowledge in dairy biology indicates that genetically superior animals differ from lesser animals mainly in their regulation of nutrient utilization and that growth hormone (GH) release

(Bauman, 1992). The report of Sørensen et al. (2002) demonstrated that induction of growth hormone releasing hormone (GHRH) in Danish Jersey calves affected the GH Leu/Leu genotype to increase the level of GH in blood as compared with their counterparts with GH Val/Val genotype. Such an effect was only intermediate when applied to GH Leu/Val genotype.

Conclusion

It was concluded that dairy cow population at Grati is polymorphic at locus GH with 22% polymorphism level. $Msp1^+/Msp1^+$ and $Msp1^+/Msp1^-$ genotypes can be used as the candidate genes in dairy cow selection to improve body weight.

ACKNOWLEDGEMENTS

The author wishes to express sincere gratitude to Prof. Muladno of Bogor Institute of Agriculture for his tangible help during the study and most particularly his skillful assistance in DNA analysis. I am deeply indebted to the manager and staff of Suka Makmur, Agricultural Farming Cooperative for warm and friendly welcome during the study that enabled me to accomplish data collection as scheduled. Thanks are due to the Dean of the Faculty of Animal Husbandry, Brawijaya University for providing valuable information to find the research grant and the Rector of Brawijaya University for his valuable support to publish this result. Last but not least, my special thank goes to Prof. Hendrawan Soetanto for his comment and willingness to discuss on the manuscript until he was ready to submit it for publication.

REFERENCES

- Bauman DE (1992). Bovine somatotropin: review of an emerging animal technology. *J. Dairy Sci.*, 75: 3432-3451.
- Budak H, Pedraza F, Cregan PB, Baenziger PS, Dweikat I (2003). Development and utilization of SSRs to estimate the degree of genetic relationships in a collection of Pearl Millet germplasm. *Crop Sci. Am.*, 43: 2284-2290.
- Dorak MT (2006). Basic Population Genetics. www.dorak.info/genetics/popgen.html
- Dybus A (2002). Associations of growth hormone (GH) and prolactin (PRL) genes polymorphism with milk production traits in Polish Black-and-White cattle. *Anim. Sci. Papers Reports*, 20(4): 203-212.
- Huitema H (1986). Peternakan di daerah tropis: Arti ekonomi dan kemampuannya. Penerbit Yayasan Obor Indonesia dan P.T. Gramedia, Jakarta.
- Hyperdictionary (2000). Meaning of Polymorphism. Copy Right © 2000-2003 Webnox Corp.
- ILRI (2011). Cattle linear measurements. In: Research on farm and livestock productivity in the central Ethiopian highlands. FAO Corporate Document Repository.
- Jakaria J, Duryadi D, Noor RR, Tappa B (2007). The Relationship of *Msp1* Growth Hormone Gene Polymorphism and Body Weight and Body Measurements of West Sumatera Pesisir Cattle. *J. Ind. Trop. Anim. Agric.*, 32(1): 33-40.
- Kish S (2008). Exploring How Growth Hormones are Released in Animals. National Institute of Food and Agriculture. USDA.
- Komisarek J, Michalak A, Walendowska A (2011). The effects of polymorphisms in *DGAT1*, *GH* and *GHR* genes on reproduction and production traits in Jersey cows. *Anim. Sci. Papers Reports*, 29(1): 29-36.
- Pawar RS, Tajane KR, Joshi CG, Rank DN, Bramkshtri BP (2007). Growth hormone gene polymorphism and its association with lactation yield in dairy cattle. *Indian J. Anim. Sci.*, 77(9): 884-888.
- Reyna XFDLR, Montoya HM, Castrellon VV, Rincon AMS, Bracamonte MP, Vera WA (2010). Polymorphisms in the *IGF1* gene and their effect on growth traits in Mexican beef cattle. *Genet. Mol. Res.*, 9(2): 875-883.
- Sørensen P, Grochowska R, Holm L, Henryon M, Løvendahl P (2002). Polymorphism in the Bovine Growth Hormone Gene Affects Endocrine Release in Dairy Calves. *J. Dairy Sci.*, 85: 1887-1893.
- Sudono A, Rosdiana F, Setiawan BS (2003). Intensive Dairy Cattle Keeping. 1st Ed. Agromedia Publisher.
- Sulandari S, Zein MSA (2003). Protocols in DNA Laboratory, Center of Biology Research, The Indonesian Institute of Sciences. Pp. 23-45.
- Thakur MS, Parmar SNS, Tolengkomba TC, Srivastava PN, Joshi CG, Rank DN, Solanki JV, Pillai PVA (2006). Growth Hormone gene polymorphism in Kandaknath breed of poultry. *Ind. J. Biotechnol.* 5: 189-194.
- Yardibi H, Hosturk HT, Paya I, Kaygisiz F, Ciftioglu G, Mengi A, Oztabak K (2009). Associations of Growth Hormone Gene Polymorphisms with Milk Production Traits in South Anatolian and East Anatolian Red Cattle. *J. Anim. Vet. Adv.*, 5: 1040-1044.
- Zhao Q, Davis ME, Hines HH (2004). Associations of polymorphisms in the *Pit-1* gene with growth and carcass traits in Angus beef cattle. *J. Anim. Sci.*, 82: 2229-2233.