academic Journals

Vol. 6(1), pp. 1-9, January 2015 DOI: 10.5897/JCECT2014.0342 Articles number: F083B8B50061 ISSN 2141-2634 Copyright © 2015 Author(s) retain the copyright of this article http://www.academicjournals.org/JCECT

Journal of Civil Engineering and Construction Technology

Full Length Research Paper

A study on arsenic and copper extraction capacity of Spirodela polyrhiza from water

Sourav Ray¹*, S. Islam¹, D. R. Tumpa¹, M. A. Kayum¹ and S. D. Shuvro²

¹Department of Civil and Environmental Engineering, Shahjalal University of Science and Technology, Sylhet, Bangladesh.

²Department of Water Resources Engineering, Bangladesh University of Engineering and Technology, Dhaka, Bangladesh.

Received 17 October, 2014; Accepted 3 December, 2014

Heavy metals such as arsenic (As), copper (Cu), chromium (Cr), Hg, lead (Pb) and cobalt (Co) cause adverse effects on living organisms by their toxic nature. To remove heavy metals, a variety of conventional treatment technologies have been tested which are not economical and user friendly. So. natural remediation method such as phytoremediation is becoming more popular where plants are used. Phytoremediation is a cost effective and eco-friendly method. This paper accounts the study to exercise the phytoremediation potential of the aquatic plant Spirodela polyrhiza for arsenic and copper removal from water. To carry out the study, six plastic bowls each carrying 1 L distilled water were taken where arsenic and copper of known concentration was added for preparing a solution, which contained 1.0, 0.9, 0.7, 0.6, 0.5, and 0.3 mg/L of arsenic and 5.0, 4.6, 4.2, 3.8, 3.4 and 3.0 mg/L of copper. 50 g of S. polyrhiza plant was placed in water of each bowl. The concentration of arsenic and copper in water was measured by spectrophotometer using Silver diethyldithiocarbamate (SDDC) and Bicinchoninate methods, respectively and test was performed for 7 days after placing plants in solution. Gross effective floating period for S. polyrhiza was found 96 h up to initial concentration of 0.6 mg/L to treat arsenic contaminated water and 3.4 mg/L to treat copper contaminated water. The extraction capacity of S. polyrhiza was found more than 80% for all concentration of arsenic and more than 60% for all concentration of copper after 96 h. The removal of arsenic and copper was found to follow the first order kinetics except copper with initial concentration of 5.0 and 4.6 mg/L.

Key words: Spirodela polyrhiza, phytoremediation, first order kinetics.

INTRODUCTION

Environmental protection and conservation are facing new challenges due to the raise of global development (Duruibe et al., 2007). Because of globalization, industrialization and urbanization are increasing rapidly which creates problems like pollution and water pollution is one of them. Consequent to water pollution, there is a scarcity of fresh water in every part of the world. Heavy metal pollution is one of the reasons of water pollution. Heavy metals can occur naturally or can be found in industrial waste. Heavy metal pollution is a widespread problem and has direct effect on human and environmental health (Hogan, 2012). Heavy metals are

*Corresponding author. E-mail: sourav.ceesust@gmail.com. Author(s) agree that this article remain permanently open access under the terms of the Creative Commons Attribution License 4.0 International License toxic at higher concentrations. Common heavy metals are cadmium (Cd), lead (Pb), cobalt (Co), zinc (Zn), chromium (Cr), copper (Cu) and arsenic (As) (Agarwal, 2009; Merrill et al., 2007).

Among the various heavy metals, arsenic and copper are well known toxic metal. Arsenic contamination is a great threat to millions of people in many countries of the world such as China, Bangladesh, Nepal, Myanmar and Thailand (Bissen and Frimmel, 2003). Arsenic creates many human health problems. Symptoms of acute arsenic poisoning are nausea, vomiting, diarrhoea, cyanosis, cardiac arrhythmia and confusion. Symptoms of chronic arsenic poisoning are less specific (Ng et al., 2003). These include depression, numbress, sleeping disorders and headaches. Arsenic related health effects are usually not acute, but mostly encompass cancer, mainly skin cancer (Wang et al., 2007). The World Health Organizations (WHO) provisional guideline of 0.01 mg/L has been adopted as the drinking water standard. However, many countries have retained the earlier WHO guideline of 0.05 mg/L as their standard including Bangladesh and China.

Copper is present in the wastewater of several industries, such as metal cleaning and plating baths, refineries, paper and pulp, fertilizer and wood preservatives (Periasamy and Namasivayam, 1996). Copper can be found in many kinds of food, water and air. Because of that, people absorb eminent quantities of copper each day by eating, drinking and breathing. The excessive intake of copper by man leads to severe mucosal irritation, widespread capillary damage, hepatic and renal damage, liver and kidney damage (Kalavathy et al., 2005). The WHO provisional guideline for copper is 1.0 mg/L which is also for the Bangladesh drinking water quality standard has been adopted worldwide.

Several techniques based on the principal of precipitation, ion exchange, electrolysis, solvent extraction, reverse osmosis, membrane and bio-sorption process (McNeill and Edwards, 1997; Tiravanti et al., 1997; Kumari et al., 2006) have been established to remove metals from water. It is very difficult to select an appropriate one. Some are effective but not economical. Some are not user friendly, technologically not sound, post treatment required, and skill manpower required. Sometimes water standard cannot be maintained.

Among these methods, phytoremediation technology has become increasingly popular. Aquatic plants and their associated microbes are used to absorb metals from surrounding water and are extremely efficient. It is considered a clean, cost effective and nonenvironmentally disruptive technology (Hannink et al., 2001).

Ebel et al. (2007) studied on *Eichhomia crassipes* and found that it has high growth rate, high tolerance to pollution and has absorption capacity of heavy metal. It is an efficient plant for wastewater treatment (Fang et al., 2007; Ebel et al., 2007). *Myriophyllum aquaticum*,

Ludwigina palustris and Mentha aquatic could effectively remove Fe, Zn, Cu and Hg from contaminated water (Kamal et al., 2004). *Lemna minor* could accumulate Cu and Cd from contaminated wastewater (Kara, 2004; Hou et al., 2007). *Myriophyllum spicatum* was an efficient plant for the metal contaminated industrial wastewater treatment (Lesage et al., 2007).

Greater duckweed (*Spirodela polyrhiza*) was tested under laboratory condition by Rahman et al. (2007, 2008) to investigate arsenic uptake efficiency and mechanisms interaction with PO_4^{3-} and Fe ions. They observed that As (v) uptake by *S. polyrhiza* was negatively correlated with phosphate uptake and positively correlated with iron uptake. Total arsenic was extracted by the plant about 56%.

Arsenic removal from water by *E. crassipes* was performed by Alvarado et al. (2008) and the results found that it had a removal rate of 600 mg arsenic ha⁻¹ d⁻¹ under field condition and a removal recovery of 18% under laboratory condition.

The potentiality of the rootless duckweed *Wolffia globosa* for arsenic accumulation and tolerance was investigated by Zhang et al. (2009). It was found that this plant can accumulate >1000 mg of arsenic kg⁻¹ dry weight (dw) and can tolerate up to 400 mg arsenic kg⁻¹ dw.

Loveson et al. (2013) studied on the efficiency of *S. polyrhiza* to improve the quality of two polluted wetland. In first wetland, percentage reduction of heavy metals such as Pb, Cu, Zn Cr, Hg, Co and Mg after 8 days treatment period was 95, 79, 66, 53, 45, 26, 20 and 7%, respectively. Again for the same treatment period of 8 days second wetland's heavy metals such as Cd, Fe, Pb, Cu, Zn and Hg reduced by 100, 98, 91, 74, 62 and 53%, respectively.

The present study establishes to develop the phytoremediation potential of the aquatic plant *S. polyrhiza* for arsenic and copper from water.

MATERIALS AND METHODS

Selection of plant material

Young aquatic plants *S. polyrhiza* (L). Schleiden were collected from a lake of Shahajalal University of Science and Technology and rinsed with tap water to remove any epiphytes and insect larvae grown on plants. The plants were placed in pots with tap water under natural sunlight for 1 day to allow them to adapt the new environment.

Sample preparation

Stock solution of arsenic and copper was prepared in the laboratory. Solution was added to each bowl containing 1 L distilled water for preparing arsenic concentration of 0.3, 0.5, 0.6, 0.7, 0.9 and 1 mg/L and copper concentration of 3, 3.4, 3.8, 4.2, 4.6 and 5 mg/L.



Figure 1. Spirodela polyrhiza in plastic bowls.

Experimental setup

The whole experiment was carried out in the water supply and sanitation laboratory of Department of Civil and Environmental Engineering of Shahajalal University of Science and Technology. Six identical plastic bowls (radius 9" and depth 4") were used in this experiment (Figure 1). Each bowl was kept in open air for growing plants naturally. Spectrophotometer and other subsidiary equipment were used to perform the work.

Experimental procedures

50 g of plants were transplanted into every pot of arsenic and copper test and allow them to take water containing arsenic and copper. Water was collected from each bowl after 1, 2, 4 and 6 h in 1st day to measure the remaining amount of arsenic and copper in water. From the next day, water was collected once a day and it continued up to 7 days. Concentration of arsenic and copper in water was measured by spectrophotometer using Silver diethyldithiocarbamate (SDDC) method (SM 3500-As B, 1999) and Bicinchoninate method (Method 8506, Hach Handbook of Water Analysis, 1979), respectively. No external agent was required during the experiment period.

Kinetic modeling

It was found out whether the extraction of arsenic and copper from water by *S. polyrhiza* follows first order kinetics or not.

RESULTS AND DISCUSSION

Experimental results for arsenic

As *S. polyrhiza* was extracting arsenic from water, the concentration of arsenic in water decreased with time and

within 6 h concentration of arsenic in water decreased from 1, 0.9, 0.7, 0.6, 0.5 and 0.3 mg/L to 0.401, 0.561, 0.296, 0.151, 0.235 and 0.087 mg/L, respectively.

Figure 2 shows a graph representing the remaining concentration of arsenic in water in first 6 h. From graph it was found that for all the water samples the plot was steep in first hour. It means extraction rate was higher in the first hour. But after that it was going from steep to flat trend which indicate that the extraction rate was decreasing with time in the first 6 h.

Figure 3 shows a graph representing the arsenic concentration in water during 7 days. From graph it was found that the concentration of arsenic gradually decreased up to 96 h (that is, up to 5th day) from planting of plants to solution for all water samples. After 96 h, concentration of arsenic in water decreases from 1, 0.9, 0.7, 0.6, 0.5 and 0.3 mg/L to 0.187, 0.149, 0.083, 0.051, 0.042 and 0.01 mg/L, respectively. But after 5th day and up to 7 days from planting of plants to solution concentration of arsenic of 1, 0.9 and 0.7 mg/L started to increase and concentration of 0.6, 0.5 and 0.3 mg/L maintained static condition. The final concentration of arsenic up to 7 days for 1, 0.9, 0.7, 0.6, 0.5 and 0.3 mg/L was found as 0.203, 0.162, 0.098, 0.048, 0.039 and 0.01 mg/L, respectively.

After 96 h, the colour of leaves of *S. polyrhiza* was changed; they began to die. It can be predicted that for this reason the concentrations of arsenic of sample 0.7, 0.9 and 1.0 mg/L were increasing. After 96 h the ability of *S. polyrhiza* to absorb high concentration of arsenic (>0.6 mg/L) were decreased and they started to discharge arsenic.

After 96 h or at 5th day it was found that some samples

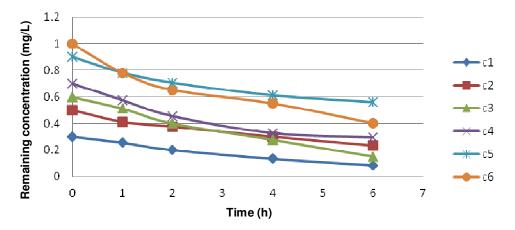


Figure 2. Remaining concentration of arsenic in water in first 6 h.

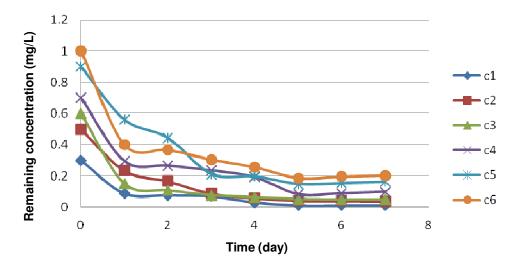


Figure 3. Rate of decreasing concentration in 7 days.

with initial concentration of 0.6, 0.5 and 0.3 mg/L reached to the margin of 0.05 mg/L or below which is the Bangladesh drinking water quality standard. But some samples with initial concentration of 1, 0.9 and 0.7 mg/L did not reach that margin. So in this case further treatment is required to ensure Bangladesh drinking water quality standard.

From the above analysis, it can be concluded that 0.6 mg/L is the boundary line of initial arsenic concentration to reach the level of Bangladesh drinking water quality standard at 96 h after phytoextraction by *S. polyrhiza* and when the initial concentration is more than 0.6 mg/L the remaining concentration of arsenic increases with time after 96 h. So, it can be said that the gross effective floating period for *S. polyrhiza* was 96 h up to initial concentration of 0.6 mg/L to treat arsenic contaminated water.

After completion of the experiment, removal percentage

was calculated and it was seen that removal percentage was more than 80% for all concentration at 5th day or after 96 h. At the same time 1st day removal percentage was also calculated and seen that percentage of removal obtained on the 1st day more than 50% except 0.9 mg/L concentration of arsenic and it was 37.66% (Figure 4). So it can be said that the major portion of all concentrations of arsenic was extracted on first day except 0.9 mg/L.

Experimental results for copper

Figure 5 shows a graph representing the remaining concentration of copper in water in first 6 h. In first hour the plot was steep which means extraction rate was higher but after that there was fluctuation of removal concentration. Within 6 h initial concentration of 3.4 and 5.0 mg/L started to increase and then again decreased.

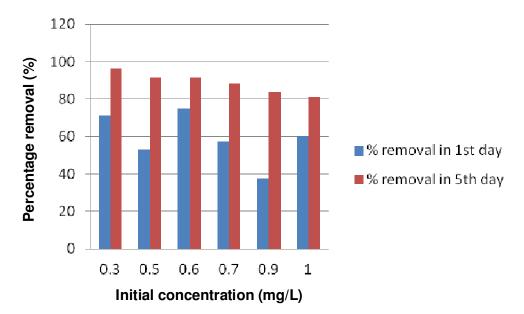


Figure 4. Percentage removal of arsenic at 1st and 5th days.

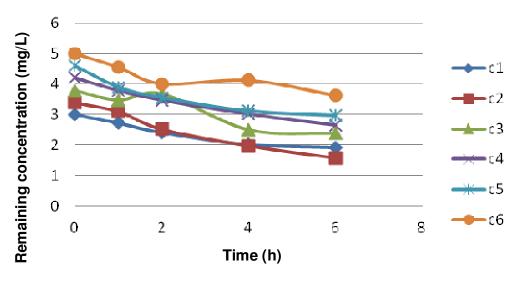


Figure 5. Remaining concentration of copper in water in first 6 h.

Concentration of copper in water decreased from 5.0, 4.6, 4.2, 3.8, 3.4 and 3.0 mg/L to 3.63, 2.97, 2.65, 2.38, 1.57 and 1.91 mg/L, respectively within 6 h.

Figure 6 shows a graph representing the copper concentration in water during 7 days. From graph it was found that the concentration of copper gradually decreased up to 5 days. But there was also fluctuation of removal concentration. All the initial concentrations of copper showed fluctuation without 3.0 mg/L. Within 5 days concentration of copper in water decreased from 5.0, 4.6, 4.2, 3.8, 3.4 and 3.0 mg/L to 1.75, 1.56, 1.3, 1.0, 0.68 and 0.45 mg/L, respectively. But after 5 days and up to 7 days concentration of copper of 5.0, 4.6, 4.2 and 3.8

mg/L started to increase and concentration of 3.4 and 3.0 mg/L maintained static condition. The final concentration of copper up to 7 days for 5.0, 4.6, 4.2, 3.8, 3.4 and 3.0 mg/L was found as 1.79, 1.62, 1.81, 1.04, 0.59 and 0.69 mg/L, respectively.

After 96 h, leaves of *S. polyrhiza* changed their colours and began to die. It can be predicted that for this reason the copper concentrations of sample 5.0, 4.6, 4.2 and 3.8 mg/L were increased. After 96 h their absorbent ability of high concentration of copper (>3.4 mg/L) were decreased and they started to discharge copper.

Some samples with initial concentration of 3.0, 3.4 and 3.8 mg/L reached to the margin of 1.0 mg/L or below

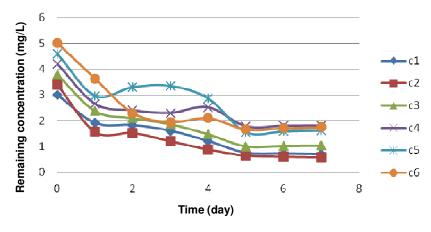


Figure 6. Rate of decreasing concentration in 7 days.

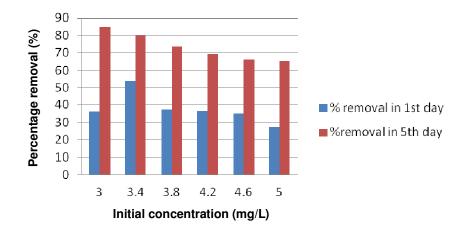


Figure 7. Percentage removal of copper after 1st and 5th days.

which is the Bangladesh drinking water quality standard and some samples with initial concentration of 4.2, 4.6 and 5.0 mg/L did not reach that margin after 96 h or at 5th day. But they can reach that margin. For that further treatment is necessary.

So, it can be said that the gross effective floating period for *S. polyrhiza* was 96 h up to initial concentration of 3.4 mg/L to treat copper contaminated water.

From Figure 7, it can be said that removal percentage was more than 60% for all concentration of copper at 5th day or after 96 h. At the same time 1st day removal percentage was also calculated and seen that percentage of removal obtained on the 1st day more than 35% except 5.0 mg/L concentration of copper and it was 27.44%.

Theory of first order kinetics

The velocity of chemical reaction is proportional to the product of the mass of active reactants present and hence for a single reactant, the velocity (dA/dt) is

proportional to the unreacted substance (A), where dA is the change in concentration of reactant over time interval dt. So, the decrease in concentration over this time period can be written as:

$$d[A]/dt = k[A]$$
(1)

Where k is the reaction rate constant.

Integrating and applying the boundary conditions, the equation of first order kinetics is found.

$$ln[A] / [A]_0 = -kt$$

$$[A] = [A]_0 e^{-kt}$$
(2)

The final Equation 2 is the exponential form of first order removal kinetics.

The removal of arsenic was found to follow the first order kinetics. According to Equation 2, In A_0/A and time t, yield a linear relationship if k is constant. The values of rate constant (k) and corresponding correlation coefficient

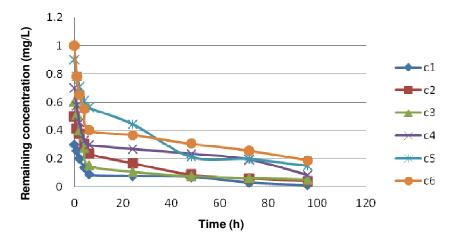


Figure 8. Arsenic concentration at different initial concentration.

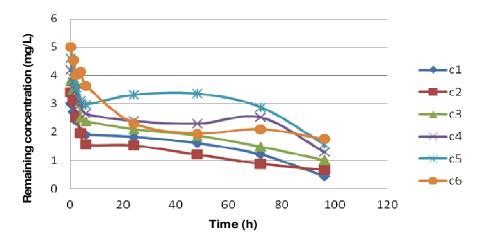


Figure 9. Copper concentration at different initial concentration.

Table 1. Equation of removal kinetics with correlation coefficient for arsenic.

Initial concentration of arsenic (mg/L)	Equation of removal kinetics	R ² Value
0.3	$y = 0.195e^{-0.02x}$	$R^2 = 0.893$
0.5	$y = 0.358e^{-0.02x}$	$R^2 = 0.944$
0.6	$y = 0.330e^{-0.02x}$	$R^2 = 0.770$
0.7	$y = 0.468e^{-0.01x}$	$R^2 = 0.824$
0.9	$y = 0.709e^{-0.01x}$	$R^2 = 0.932$
1.0	$y = 0.645e^{-0.01\times}$	$R^2 = 0.802$

 (R^2) values obtained from all experimental data are summarized in Table 1. Arsenic removal by *S. polyrhiza* can be represented by the first order kinetics as the values of R^2 is constantly greater (Figure 8).

The values of rate constant (k) and corresponding correlation coefficient (R^2) values obtained from all experimental data are summarized in Table 2. Copper removal by *S. polyrhiza* can be represented by the first order kinetics as the values of R^2 is constantly greater

except copper with initial concentration of 5.0 and 4.6 mg/L. For the initial concentration of 5.0 and 4.6 mg/L greater values of R^2 is 0.909 and 0.651, respectively. So, they did not follow the first order kinetics (Figure 9).

Conclusion

The extraction of arsenic and copper from water using

Initial concentration of copper (mg/L)	Equation of removal kinetics	R ² Value
3.0	$y = 2.580e^{-0.01\times}$	$R^2 = 0.858$
3.4	$y = 2.460e^{-0.01x}$	$R^2 = 0.853$
3.8	$y = 3.176e^{-0.01\times}$	$R^2 = 0.884$
4.2	$y = 3.432e^{-0.01x}$	$R^2 = 0.745$
4.6	$y = 3.790e^{-0.01x}$	$R^2 = 0.647$
5.0	$y = 4.061e^{-0.01x}$	$R^2 = 0.809$

Table 2. Equation of removal kinetics with correlation coefficient for copper.

S. polyrhiza was studied in this research. After 96 h the ability of S. polyrhiza to absorb high concentration of arsenic (>0.6 mg/L) and copper (>3.4 mg/L) were decreased and they started to discharge arsenic and copper. Arsenic with initial concentration (≦0.6 mg/L) and copper with initial concentration (≦3.8 mg/L) reached to the margin of Bangladesh drinking water guality standard 0.5 and 1.0 mg/L, respectively. High concentration of arsenic (>0.6 mg/L) and copper (>3.8 mg/L) need further treatment to reach that margin. Gross effective floating period for S. polyrhiza is 96 h up to initial concentration of 0.6 mg/L to treat arsenic contaminated water and 3.4 mg/L to treat copper contaminated water. The removal percentage was more than 80% for all concentration of arsenic and for copper the removal percentage was more than 65% for all concentration. It was also revealed that the major portion of all concentration of arsenic that is more than 50% was extracted on 1st day except 0.9 mg/L. But for copper it was more than 35% for all concentration except 5.0 mg/L. The removal or accumulation of arsenic and copper from water by S. polyrhiza follows first removal kineticsexcept copper with initial order concentration of 5.0 and 4.6 mg/L.

Conflict of Interest

The author(s) have not declared any conflict of interest.

ACKNOWLEDGEMENTS

The authors are grateful to the Department of Civil and Environmental Engineering, Shahjalal University of Science and Technology for providing the laboratory facilities. This is also acknowledged that this work is a version of an undergraduate thesis work of D.R. Tumpa and M.A. Kayum supervised by Sourav Ray.

REFERENCES

Agarwal SK (2009). Heavy metal pollution. APH publishing, 1-270. Alvarado S, Guédez M, Lué-Merú MP, Nelson G, Alvaro A, Jesús AC, Gyula Z (2008). Arsenic removal from waters by bioremediation with the aquatic plants water hyacinth (*Eichhornia crassipes*) and lesser duckweed (*Lemna minor*). Bioresour. Technol. 99:8436-8440. http://dx.doi.org/10.1016/j.biortech.2008.02.051; PMid:18442903

- Bissen M, Frimmel FH (2003). Arsenic A review. Part I. Occurance, toxicity, speciation, mobility. Acta Hydrochim. Hydrobiol. 31:9-18. http://dx.doi.org/10.1002/aheh.200390025
- Method 8506, Hach Handbook of Water Analysis (1979). Copper, Bicinchoninate Method, Hach Company, P. O. Box 389, Loveland, CO 80537. Available on-line at http://www.hach.com.
- Duruibe JO, Ogwuegbu MOC, Egwurugwu JN (2007). Heavy metal pollution and human biotoxic effects. Int. J. Phys. Sci. 2:112-118.
- Ebel M, Evangelou MWH, Schaeffer A (2007). Cyanide phytoremediation by water hyacinths (*Eichhornia crassipes*). Chemosphere 66:816-823. http://dx.doi.org/10.1016/j.chemosphere.2006.06.041;

PMid:16870228

- Fang YY, Yang XE, Chang HQ, Pu PM, Ding XF, Rengel Z (2007). Phytoremediation of nitrogen-polluted water using water hyacinth. J. Plant Nutr. 30:1753-1765. http://dx.doi.org/10.1080/15226510701375507
- Hannink N, Rosser SJ, French CE, Basran A, Murray JAH, Nicklin S, Bruce NC (2001). Phyodetoxification of TNT by transgenic plants expressing a bacterial nitroreductase. Nature Biotech. 19:1168-1172.
- Hogan CM (2012). "Heavy metal". In: Encyclopedia of Earth. Eds. Cutler J. Cleveland (Washington, D.C.: Environmental Information Coalition, National Council for Science and the Environment).
- Hou WH, Chen X, Song GL, Wang QH, Chang CC (2007). Effects of copper and cadmium on heavy metal polluted water body restoration by duckweed (*Lemna minor*). Plant Physiol. Biochem. 45:62-69. http://dx.doi.org/10.1016/j.plaphy.2006.12.005; PMid:17300947
- Kalavathy MH, Karthikeyan T, Rajgopal S, Miranda LR (2005). Kinetics and isotherm studies of Cu (II) adsorption onto H₃PO⁴ activated rubber wood sawdust. J. Colloid Interface Sci. 292:354-362. http://dx.doi.org/10.1016/j.jcis.2005.05.087; PMid:16040040
- Kamal M, Ghaly AE, Mahmoud N, Cote R (2004). Phytoaccumulation of heavy metals by aquatic plants. Environ. Int. 29:1029-1039. http://dx.doi.org/10.1016/S0160-4120(03)00091-6
- Kara Y (2004). Bioaccumulation of copper from contaminated wastewater by using *Lemna minor*. Bull. Environ. Contam. Toxicol. 72:467-471. http://dx.doi.org/10.1007/s00128-004-0269-4
- Khondker M, Islam AKM, Nurul, Nahar N (1993). A preliminary study on the growth rate of *Spirodela polyrhiza*. Dhaka Univ. J. Biol. Sci. 2:197-200.
- Kumari P, Sharma P, Srivastava S, Srivastava MM (2006). Biosorption studies on shelled *Moringa oleifera* Lamarck seed powder: Removal and recovery of arsenic from aqueous system. Int. J. Miner. Process. 78:131-139. http://dx.doi.org/10.1016/j.minpro.2005.10.001
- Lesage E, Mundia C, Rousseau DPL, Van de Moortel AMK, Du Laing G, Meers E, Tack FMG, De Pauw N, Verloo MG (2007). Sorption of Co, Cu, Ni and Zn from industrial effluents by the submerged aquatic macrophyte *Myriophyllum spicatum* L. Ecol. Eng. 30:320-325. http://dx.doi.org/10.1016/j.ecoleng.2007.04.007
- Loveson A, Sivalingam R, Syamkumar R (2013). Aquatic Macrophyte *Spirodela Polyrrhiza* as a Phytoremediation Tool in Polluted Wetland Water from Eloor, Ernakulam District, Kerala. Environ. Anal. Toxicol. 3:1-7.

- McNeill S, Edwards M (1997). Predicting arsenic removal during metal hydroxide precipitation. J. Am. Water Works Assoc. 89:75-82.
- Merrill JC, Morton JJP, Soileau SD (2007). Metals. In A. W. Hayes. Principles and methods of toxicology, 5th edition. CRC Press, ISBN 084933778X.
- Ng JC, Wang J, Shraim A (2003). A global health problem caused by arsenic from natural sources. Chemosphere 52:1353-1359. http://dx.doi.org/10.1016/S0045-6535(03)00470-3
- Periasamy K, Namasivayam C (1996). Removal of copper (II) by adsorption onto peanut hull carbon from water and copper plating industry wastewater. Chemosphere 32:769-789. http://dx.doi.org/10.1016/0045-6535(95)00332-0
- Rahman MA, Hasegawa H, Ueda K, Maki T, Okumura C, Rahman MM, (2007). Arsenic accumulation in duckweed (*Spirodela polyrhiza* L.): A good option for phytoremediation. Chemosphere 69:493-499. http://dx.doi.org/10.1016/j.chemosphere.2007.04.019; PMid:17509657
- Rahman MA, Hasegawa H, Ueda K, Maki T, Rahman MM (2008). Arsenic uptake by aquatic macrophyte *Spirodela polyrhiza* L.: Interactions with phosphate and iron. J. Hazard. Mater. 160:356-361. http://dx.doi.org/10.1016/j.jhazmat.2008.03.022; PMid:18430512
- SM 3500-As B (1999). Standard Methods for the Examination of Water and Wastewater, Silver Diethyldithiocarbamate method.
- Tiravanti G, Petruzzelli D, Passiono R (1997). Pretreatment of tannery wastewaters by an ion exchange process for Cr (III) removal and recovery. Water Sci. Technol. 36:197-207. http://dx.doi.org/10.1016/S0273-1223(97)00388-0

- Wang CH, Hsiao CK, Chen CL, Hsu LI, Chiou HY, Chen SY, Hsueh YM, Wu MM, Chen CJ (2007). A review of the epidemiologic literature on the role of environmental arsenic exposure and cardiovascular diseases. Toxicol. Appl. Pharmacol. 1:315-326. http://dx.doi.org/10.1016/j.taap.2006.12.022; PMid:17433393
- Zhang X, Zhao FJ, Huang Q, Williams PN, Sun GX, Zhu YG (2009). Arsenic uptake and speciation in the rootless duckweed *Wolffia globosa*. New Phytol. 182:421-428. http://dx.doi.org/10.1111/j.1469-8137.2008.02758.x; PMid:19210724