

*Full Length Research Paper*

# Foliar nutrient response in some Iranian quince genotypes

M. Mirabdulbaghi\* and H. Abdollahi

Department of Horticulture, Seed and Plant Improvement Research Institute, Karaj, Iran.

27 September, 2013; 8 September, 2014

**Significant foliar nutrient response due to genetic variability was seen for N, P, K, Mg, Ca, B, Fe and Zn for some Iranian quince genotypes, which were selected from different parts of Iran (during 2006-2009). Vector analysis has been used to interpret plant nutrient status and nutrient shifts (dilution, deficiency, excess, etc) of studied quince genotypes. In our study, reference point for calculating and comparing the relative change of the three parameters (nutrient concentration, nutrient content and leaf dry weight) for studied quince genotypes was the average value of tissue concentrations, content of nutrient and leaf dry weight which were normalized to 100% to allow comparison on a common base. Vector analysis diagnoses of foliar response revealed excess “E-shifts” behavior of all studied nutrient, as compared to the control, in the genotype ASM3. Steady-status “B-shifts” and Luxury consumption “D-shifts” behaviors were not shown by any of studied nutrient among studied quince genotypes. Excess ‘E-shifts’ and Antagonism “F-shifts” behaviors were presented by most of studied nutrient among studied quince genotypes.**

**Key words:** Foliar nutrient, Iranian quince genotypes, variability.

## INTRODUCTION

Quince (*Cydonia oblonga* Mill.) belongs to the Maloideae subfamily of the Rosaceae family, which includes commercially important fruits such as apples and pears. This subfamily comprises approximately 1,000 species in 30 genera and is characterized by a distinctive fruit, pome, and a base chromosome number of 17 (Rodger and Campbell, 2002). Quinces have originated in Persia, Turkistan and the Caucasus. The quince tree shows high genetic variability, the following authors studied the genetic variability problem in this species (Scaramuzzi, 1957; Onofrio et al., 1998; Rodrigues-Guisado et al.,

2009). Some of the Iranian authors studied the within-species variability of the leaf structure of quince genotypes (Abdollahi and Ghahremani, 2011; Khoramdel et al. 2013). According to Abdollahi et al. (2013), Quince genotypes from the North of Iran with most similarity to the wild ancestors demonstrated low fruit quality, late to very late fruit maturity and high fruit set. These genotypes also clustered as the most dwarfing and showed the lowest level of leaf chlorosis in calcareous soils. Our hypothesis was that leaf mineral compositions differ among selected quince genotypes from different parts

\*Corresponding author. E-mail: mitra\_mirabdulbaghi@yahoo.com, Tel: 0098-(261)-6702541&6703772. Fax: 0098-(261)-6700908.

Author(s) agree that this article remain permanently open access under the terms of the [Creative Commons Attribution License 4.0 International License](http://creativecommons.org/licenses/by/4.0/)

of Iran within the same stand and that these differences are under genetic control. Hence, vector analysis was developed for nutritional diagnosis purposes (Timmer and Stone, 1978; Timmer and Armstrong, 1987). Vector diagnosis involves deficiency comparing nutrient (compares) concentration, nutrient content, and biomass of plants in a graphic format known as a vector nomogram. Plant tissues are sampled and usually compared to a control or reference. Based on the magnitude and direction of vectors describing response to treatment in terms of these three variables, analysis can be used to diagnose nutrient status: sufficient, deficiency, luxury consumption, excess and dilution (Garcia et al., 2005).

In this study, we examined the mineral nutrition of 28 quince genotypes by determining leaf levels of elements and diagnosing this nutrition by vector analysis and also selecting quince genotypes that possess desirable characteristics for ability to use in breeding projects, for example, propagations of quince genotypes which induce a higher tolerance to iron

## MATERIALS AND METHODS

### Plant material and experimental site

The plant material used in this investigation belonged to the breeding programs of Iranian National Quince collection from different parts of Iran (Isfahan, Khorasan Orumia, Ardebil, Astara, and Tehran) during 2006-2009. All selected quince genotypes were budded on quince seedling rootstocks in 2012, and then grown under the same environmental conditions in nursery of Seed and Plant Improvement Institute.

### Plant sampling and analysis

In the present work, leaves were sampled from twenty eight quince genotypes which were selected from Central, Central-North, North, North West and North East regions of Iran. The leaf samples were dried at 75°C for 72 h and ground to pass a 40-mesh screen, and their mass was measured. The nitrogen content estimated by the Kjeldahl method. Ca, Mg, Fe, Zn and B were determined by atomic absorption spectrophotometry. P was analyzed by the molybdovanadat method. K was analyzed by flame photometry [Association of Official Analytical Chemists (AOAC) 1980]. Nutrient concentrations in leave and fruit tissues were expressed on a dry weight (DW) basis.

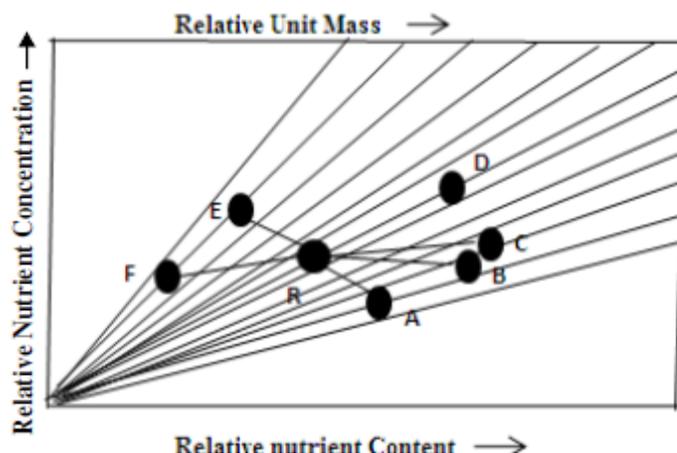
### Statistical analysis and data interpretation

The experiment was conducted in a Randomized Complete Block Design with 3 replications. The statistical evaluation was done by using analysis of variance (ANOVA). This paper would use SAS statistic computer system to calculate the surveyed data and means were evaluated using Duncan's multiple range test at  $P=0.05$ . The relationships between studied parameters were evaluated by Pearson's correlation coefficients at  $P \leq 0.05$ . Vector analysis was used to compare leaf dry weight, nutrient concentrations and nutrient content (Timmer and Stone, 1978; Imo and Timmer (1997); Weetman and Fournier, 1982; Garcia et al., 2005). Each point on the vector analysis represents the magnitude and directional shift of

each nutrient from the control. Distance from the control represents the responsiveness of the studied genotype for the nutrient being analyzed. Figure 1 shows a schematic of the approach for added nutrients. A detailed description of vector analysis can be found in Weetman and Fournier (1986) and Hasse and Rose (1995).

## RESULTS

ANOVA results showed that there were significant differences ( $p \leq 0.01$ ), between studied quince genotypes in respect to the all studied traits (Table 1). The results suggest that estimated variations of studied leaf B-content was slight, but statistically significant. The highest variability was estimated for the leaf P-content (10.56), and the lowest for leaf B-content (0.26). The results of the leaf nutrient content, leaf dry matter taken are shown in Table 2. Leaf dry weight (0.48 g) was highest for the genotype *Sahelborgmoghavem*. The leaves of genotype *KVD1* had the highest amount of leaf N-content (4.88%). The highest value of leaf P-content (0.27%) belonged to *Shai*. The highest amount of leaf K-content (4.45%) belonged to *ASP1*. The leaves of genotype *ASP2* had the highest amount of leaf Zn-content (71.94 ppm). The highest value of leaf Ca-content (2.36%), leaf Mg-content (1.57%), leaf Fe-content (57.48 ppm), leaf B-content (80.69 ppm) belonged to *NB2* (for Ca-content), *Moghavem2* (for Mg-content), *ASM3* (for Fe-content) and *NB4* (for B-content), respectively. Simple correlation analysis showed significant negative and positive correlations between studied characteristics of selected quince genotypes from different parts of Iran (Table 3). Positive correlation was observed between leaf Zn-content with leaf N-content ( $P < 0.001$ ;  $R_2 = 0.497$ ). In contrast, there were negative significant correlations between leaf Mg-content with leaf Ca-content ( $P < 0.001$ ;  $R_2 = -0.559$ ), leaf B-content with leaf Zn-content ( $P < 0.01$ ;  $R_2 = -0.466$ ) and leaf Fe-content with leaf dry weight ( $P < 0.01$ ;  $R_2 = -0.384$ ). We also observed negative significant correlation between leaf Fe-content and leaf dry weight. Vector analysis has been used to interpret plant nutrient status and nutrient shifts (dilution, deficiency, excess, etc.) of studied quince genotypes. In our study, reference point for calculating and comparing the relative change of the three parameters (nutrient concentration, nutrient content and leaf dry weight) for studied quince genotypes was the average value of tissue concentrations, content of nutrient and leaf dry weight which were normalized to 100% to allow comparison on a common on a base. The nomograms show upward, right, - and left-pointing of N, - P, - K, -, Ca, - Mg, - Zn, - B, - and Fe-vectors associated with all of studied quince genotypes, except of "*Esphehanoghaf*", compared to the control. The "*Esphehanoghaf*" showed downward of all studied nutrient as compared to the control (Figures 2 to 9). Vector analysis showed deficiency, a "C shift", for foliar N, relative to control values, in genotypes *KVD1*, *ASP2*. However, *Behtorsh* and *Moghavem1* showed N dilution effects or "A Shift"



Vector shift	Change in relative			Interpretation	Nutrient status
	Relative unit mass	Relative nutrient content	Relative nutrient concentration		
A	+	+	-	dilution	Non-limiting
B	+	+	0	Accumulation	Sufficiently, steady-status
C	+	+	+	Accumulation	Limiting, deficiency response
D	0	+	+	Accumulation	Non-limiting, luxury consumption
E	-	-,+	+	Concentration	Excess, toxic accumulation
F	-	-	-	Antagonism	Excess, antagonism

**Figure 1.** Nutrient vector analysis. Interpretation of directional changes in relative dry mass and nutrient status of plants (or plant components) contrasting in growth and/or health. The reference condition (R) is usually normalized to 100. Diagnosis (A to F) is based on shifts (increase [+], decrease [-] or no change [0]) of individual nutrient characterized in dose response curves relating plant growth (or plant unit mass), nutrient concentration, and nutrient content. Vector magnitude reflects extend or severity of the diagnosis identified (modified from Timmer, 1991). The results in this paper involve mostly vectors E and F, suggesting that the toxic accumulation of nutrient E antagonistically induced a deficiency of nutrient F.

**Table 1.** Variance analysis for leaf nutrient content, leaf dry matter of selected quince genotypes from different parts of Iran.

Sourced variation	df	MS								
		Leaf-N (%)	Leaf-P (%)	Leaf-K (%)	Leaf-Ca (%)	Leaf-Mg (%)	Leaf-Zn (ppm)	Leaf-Fe (ppm)	Leaf-B (ppm)	Leaf dry weight (g)
Treatments	29	1.67	0.01	0.56	0.46	0.43	279.42	183.91	227.61	0.01
Error	54	0.01	0.0001	0.01	0.005	0.0003	15.37	0.009	0.02	0.001
CV (%)		4.28	10.56	3.61	4.60	2.29	8.67	0.29	0.26	6.3

Significance is at  $p \leq 0.001$ .

(Figure 2). Vector analysis for *Khosro*, *Sahelborgmoghavem* and *ET1* displayed a "C" deficiency shift for foliar P (Figure 3). Vector analysis for *Behtorsh*, *ET1* and *ASP1* displayed a "C" deficiency shift for foliar k. N dilution effects or "A-Shifts" for foliar k was observed in *KVD1* (Figure 4). *ET1* and *ASP1* displayed a "C" deficiency shift for foliar Ca, relative to control values. "A-Shifts" relative to the controls were shown for *Khosro* and *Esphehanoghaf* (Figure 5). Mg deficiencies, a "C-shift", were observed for *ET1*, *unknown*, *Behtorsh*, *ASP2*, *NB3*,

*NB4* and *Moghavem1*, relative to the control (Figure 6). Nutrient vector analysis showed a "C-shift", of Zn for *unknown*, *Behtorsh* and *ASP2*. *KVD1* and *NB3* showed a Zn dilution "A-Shift" (Figure 7). *KVD3*, *Sahelborgmoghavem*, *ASP1*, *NB4* and *ET1* expressed B deficiency "C-Shifts" relative to the control. B dilution effects, an "A-Shift", were observed on *Khosro*, *Esphehanoghaf* (Figure 8). Fe deficiency "C-Shifts" were also produced by *KVD3*, *ET1* and *Behtorsh* relative to the control. Fe dilution effects, an "A-Shift", on the

**Table 2.** Leaf nutrient content, leaf dry matter and leaf surface of selected quince genotypes from different parts of Iran.

Quince genotype	Leaf-N	Leaf-P	Leaf-K	Leaf-Ca	Leaf-Mg	Leaf -Zn	Leaf-Fe	Leaf-B	Leaf dry weight (g)
	(%)						(ppm)		
KVD2	2.97	0.09	2.62	1.37	0.72	37.93	32.78	64.29	0.31
ASM3	2.57	0.14	2.67	2.13	0.72	46.76	57.48	64.03	0.32
KVD3	2.13	0.097	2.62	1.37	0.72	42.51	37.72	66.11	0.39
SVS2	3.55	0.15	2.56	1.29	0.42	35.32	36.77	64.55	0.33
KVD4	1.55	0.11	2.67	1.6	0.35	37.93	29.17	59.61	0.35
Khosro	1.818	0.13	2.57	1.6	0.23	35.97	22.23	62.21	0.43
PH2	1.95	0.1	2.67	2.36	0.46	40.88	32.4	58.57	0.32
Sahe Lbor gmoghavem	2.62	0.13	2.31	1.52	0.62	36.62	35.53	78.61	0.48
ET1	2.16	0.11	2.97	1.75	0.95	41.2	39.14	65.07	0.38
NB2	2.04	0.13	2.82	2.36	0.46	44.47	27.65	61.69	0.35
KM1	2.48	0.09	3.28	1.98	0.12	42.18	23.85	62.73	0.36
ASP1	2.62	0.05	4.45	1.98	0.69	41.86	28.22	61.17	0.38
Esphehanoghaf	2.22	0.06	2.11	1.98	0.06	40.22	18.15	62.21	0.47
SVS1	2.08	0.07	2.77	1.37	0.95	35.97	41.04	59.09	0.37
ASM1	2.39	0.18	2.26	2.05	0.65	47.74	49.02	62.21	0.33
Unknown	2.39	0.06	2.36	1.22	1.41	57.88	32.97	47.37	0.42
KVD1	4.88	0.07	2.72	1.22	0.6	48.72	36.39	56.48	0.45
ASM2	3.1	0.06	2.62	1.82	0.46	41.86	35.06	52.32	0.33
PK2	2.26	0.04	2.11	1.14	1.34	48.07	34.77	44.25	0.31
Behtorsh	2.88	0.07	3.02	0.91	1.13	52.32	40.85	51.8	0.42
ASP2	3.59	0.09	2.36	1.29	0.9	71.94	26.98	51.54	0.41
SHA1	2.75	0.27	2.77	1.29	0.67	41.2	31.26	47.37	0.35
NB3	2.66	0.1	2.41	1.14	0.99	47.74	28.22	46.07	0.423
NB4	2.39	0.12	2.62	1.22	1.41	42.18	26.7	80.69	0.42
AS2	1.82	0.08	2.26	1.14	0.76	45.13	25.65	48.94	0.37
Moghavem1	2.71	0.09	2.46	1.37	0.97	43.82	34.39	47.37	0.41
Moghavem2	4.48	0.19	3.02	1.52	1.57	60.5	32.97	54.4	0.37
Gardandar	3.5	0.11	2.57	1.9	0.3	69	40.76	51.8	0.33
LSD5%	0.19	0.02	0.16	0.12	0.03	6.40	56.28	0.25	0.04

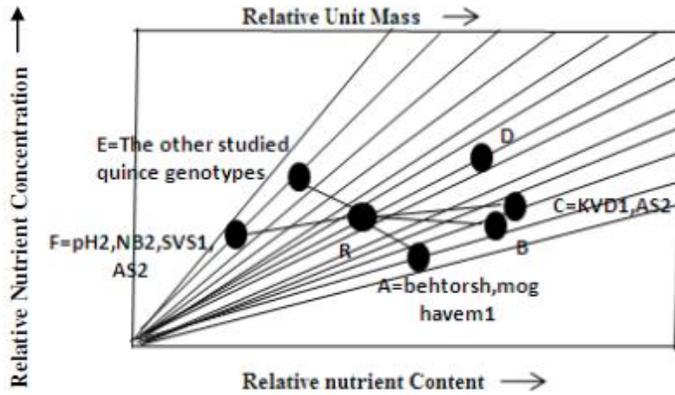
**Table 3.** Similarity coefficient between studied characteristics of selected quince genotypes from different parts of Iran.

Parameter	Leaf-N (%)	Leaf-P (%)	Leaf-K (%)	Leaf-Ca (%)	Leaf-Mg (%)	Leaf-Zn (ppm)	Leaf-Fe (ppm)	Leaf-B (ppm)	Leaf dry weight (g)
Leaf-N (%)	1								
Leaf-P (%)	0.127	1							
Leaf-K (%)	0.106	-0.039	1						
Leaf-Ca (%)	-0.235	0.111	.254	1					
Leaf-Mg (%)	0.184	-0.028	-0.033	-.559 <sub>(P&lt;0.01)</sub>	1				
Leaf-Zn(ppm)	0.497 <sub>(P&lt;0.01)</sub>	-0.038	-0.101	-.122	0.348	1			
Leaf-Fe(ppm)	0.178	0.184	-0.040	.083	0.205	0.126	1		
Leaf-B(ppm)	-0.164	0.137	0.140	.292	-0.214	-0.466 <sub>(P&lt;0.01)</sub>	.035	1	
Leaf dry weight(g)	0.088	-0.194	-0.102	-.332	0.079	.016	-.384 <sub>(P&lt;0.01)</sub>	.186	1

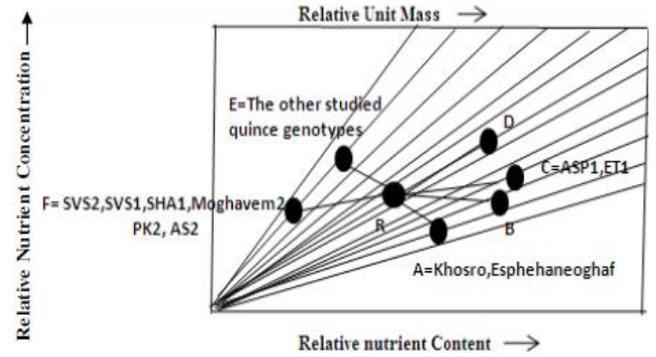
N=28\*\*.

*Sahelborgmoghavem*, *KVD1*, and *Moghavem1* were observed, respectively (Figure 9). Vector analysis

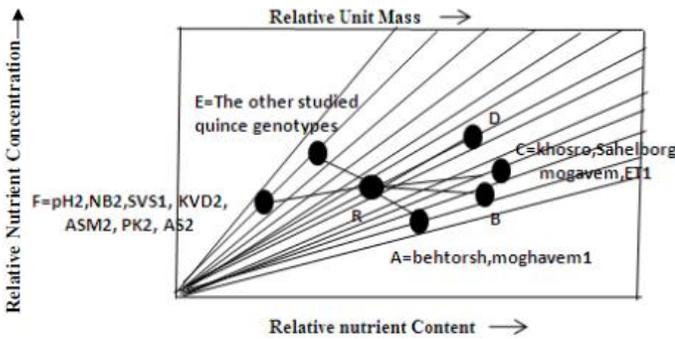
diagnoses of foliar response revealed excess "E-shifts" behavior of all studied nutrient, as compared to the



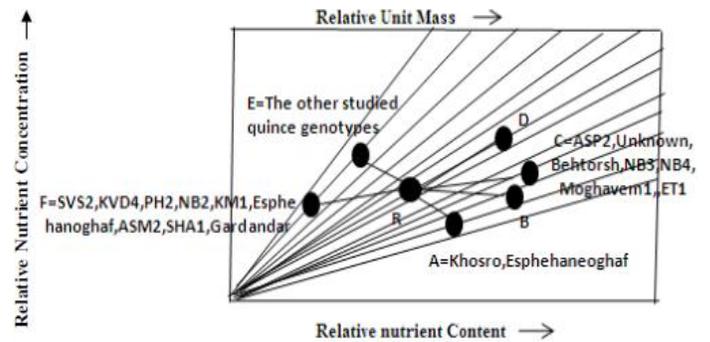
**Figure 2.** Graphical vector shifts for N concentration, content and leaf dry weight by studied quince genotypes.



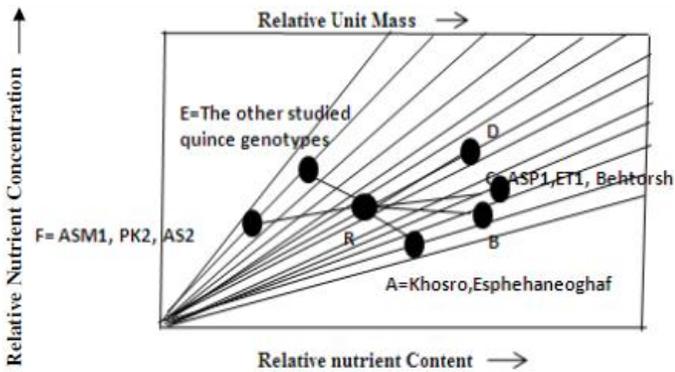
**Figure 5.** Graphical vector shifts for Ca concentration, content and leaf dry weight by studied quince genotypes.



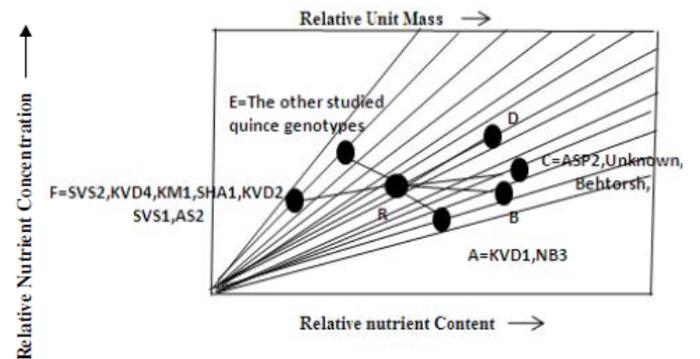
**Figure 3.** Graphical vector shifts for P concentration, content and leaf dry weight by studied quince genotypes.



**Figure 6.** Graphical vector shifts for Mg concentration, content and leaf dry weight by studied quince genotypes.



**Figure 4.** Graphical vector shifts for K concentration, content and leaf dry weight by studied quince genotypes.

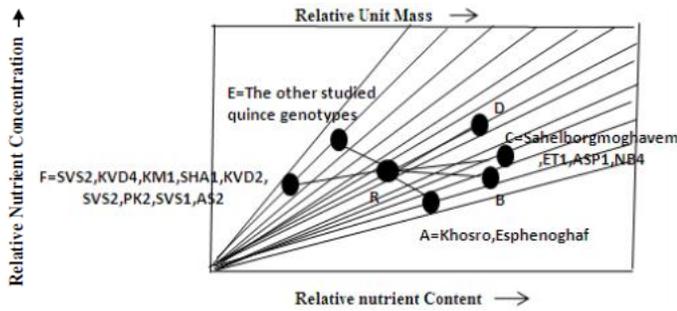


**Figure 7.** Graphical vector shifts for Zn concentration, content and leaf dry weight by studied quince genotypes.

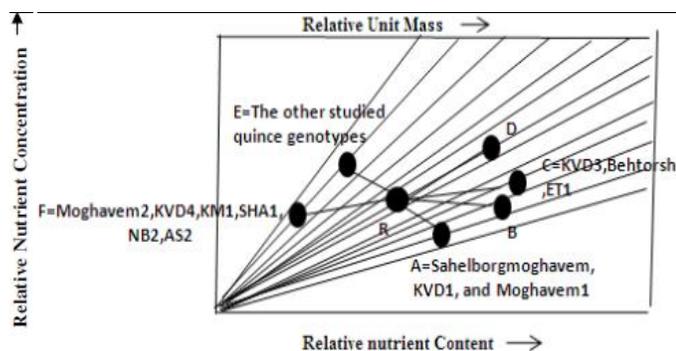
control, in the genotype ASM3. Steady-status “B-shifts” and Luxury consumption “D-shifts” behaviors were not shown by any off studied nutrient among studied quince genotypes. Excess ‘E-shifts’ and Antagonism “F-shifts” behaviors were presented by most of studied nutrient among studied quince genotypes.

**DISCUSSION**

The results suggest that estimated variations of all studied parameters were significant. The highest variability among genotypes was estimated for leaf P-content (10.56%). Somewhat lower variability was obtained for the leaf Zn-content (8.67%) and leaf dry weight (6.3%). These quantitative differences illustrated



**Figure 8.** Graphical vector shifts for B concentration, content and leaf dry weight by studied quince genotypes.



**Figure 9.** Graphical vector shifts for Fe concentration, content and leaf dry weight by studied quince genotypes.

intra species variability of parameters studied among studied quince genotypes. According to Castro-Diez et al. (1997), the within-species variability of leaf morphology and nutrient may improve plant performance, allowing species to maintain their fitness in resource availability. Also, our results of the leaf nutrient content, leaf dry matter taken is shown that leaf dry weight (0.48 g) was highest for the genotype *Sahelborgmoghavem* and the lowest for the genotype *KVD2* (0.31g). The leaves of genotype *Esphehanoghaf* had the lowest amount of leaf K-content (2.11%); leaf Mg-content (0.06%) and leaf Fe-content (18.15ppm), respectively. In similarity to Prado and Vara (2011), we have also observed negative significant correlation between leaf Fe-content and leaf dry weight. Plant nutrient test results have been shown to vary between different species or even between different ecotypes of the same species-interior (Van den Driessche, 1984; Kaufmane et al., 2002, Bussoti et al., 2000). Vector analysis in leaves, which used leaf dry weight values and the content and concentration of nutrients in selected quince genotypes, enables the interpretation of the nutritional status of studied plants to be assessed by taking as reference the average value of three parameters (tissue concentrations, content of nutrient and leaf dry weight) which were normalized to

100% to allow comparison on a common on a base.

However, any diagnosis obtained with the method cannot, strictly speaking, be generalized. It can only conclude that there are signs of deficiency, excess, antagonism, etc., compared with a reference treatment and if a different treatment is used as reference other conclusions might be reached (Garcia et al., 2005). Figure 1 demonstrates the application of the vector analysis method of Timmer and Stone (1978). In our case, the vectors for the most of studied nutrient among quince genotypes showed a general excess and antagonism values compared with control. In addition, "Steady-status" and "Luxury consumption" were not shown by any of the vectors of studied nutrient among quince genotypes.

## Conclusions

Our results approved the intra species variability of foliar nutrient response due to genetic variability among 28 quince genotypes, which were selected from different parts of Iran (during 2006-2009) and after budding on quince seedling rootstocks in 2012, grown under the same environmental conditions in nursery of Seed and Plant Improvement Institute. This information enabled to select quince genotypes possessing desirable characteristics for possible use in breeding projects. Our result demonstrated that propagations of quince genotypes *Sahelborgmoghavem*, *KVD1*, and *Moghavem1*, which Fe dilution effects on these genotypes were observed, seems to respond well in Clay-loam soils which induced chlorosis.

## Conflict of Interest

The authors have not declared any conflict of interest.

## REFERENCES

- AOAC (1980). Official Methods of Analysis Association of Official Analytical Chemists. 13th Edn. Washington, D.C.: Association of Official Analytical Chemists.
- Abdollahi H, Gahremani Z (2011). The role of chloroplasts in the interaction between *Erwinia amylovora* and host plants. *Acta Horticult.* 896:215-221.
- Abdollahi H, Alipour M, Khorramdel-Azad M, Ghasemi A, Adeli M, Atashkar, M, Akbari, M, Nasiri J (2013). Establishment and primary evaluation of quince germplasm collection from various regions of Iran. *Acta Horticult.* 976:199-206.
- Garcia AL, Gallego J, Fuentes V, Nicolas N, Madrid R (2005). Mineral Nutrition of Prunus Rootstocks: Leaf Concentrations and Diagnosis by Vector Analysis. *Horticult. Sci.* 40(6):1670-1674.
- Bussoti F, Borghni F, Celesti C, Leonzio C, Bruschi P (2000). Leaf morphology and macronutrients in broadleaved trees in central Italy. *Trees* 14:361-368. <http://dx.doi.org/10.1007/s004680000056>
- Castro-Diez P, Villar-Salvador P, Perez-Rontome C, Maestro-Martinez M, Mometesert G (1997). Leaf morphology and leaf chemical composition in three *Quercus* (Fagaceae) species along a rainfall gradient in NE Spain. *Trees* 11:127-134.

- Garcia AL, Gallego J, Fuentes V, Nicolas N, MADRID R (2005). Mineral Nutrition of Prunus Rootstocks: Leaf Concentrations and Diagnosis by Vector Analysis. *HortScience* 40(6):1670-1674.
- Hasse DL, Rose R (1995). Vector analysis and its use for interpreting plant nutrient shifts in response to silvicultural treatments. *Forest Sci.* 41:54-66.
- Imo M, Timmer VR (1997). Vector diagnosis of nutrient dynamics in mesquite seedlings. *For. Sci.* 43:268-273.
- Kaufmane E, Ikase L, Trajkovski V, Lacin G (2002). Evaluation and characterization of plum genetic resources in Sweden and Latvia. *Acta Horticult.* 577:207-213.
- Khoramdel, M, Jabernaisiri A, Abdollahi H (2013). Genetic Diversity of Selected Iranian Quinces Using SSRs from Apples and Pears. *Biochemical Genetics.* 51(5-6):426-442. <http://dx.doi.org/10.1007/s10528-013-9575-z> PMID:23430114.
- Onofrio CD, Morini S, Bellocchi G (1998). Effect of light quality on somatic embryogenesis of quince leaves. *Plant cell, Tissue and organ culture*, 53:91-98. <http://dx.doi.org/10.1023/A:1006059615088>.
- PradoRM, and Alcantara-Vara E (2011). Tolerance to iron chlorosis in non-grafted quince seedlings and in pear grafted onto quince plants. *J. Sci. Plant Nut.* 11 (4): 119-128.
- Rodger CE, Campbell CS (2002). The origin of the apple subfamily (Maloideae; Rosaceae) is clarified by DNA sequence data from duplicated GBSSI genes. *Am. J. Bot.* 89:1478-1484. <http://dx.doi.org/10.3732/ajb.89.9.1478> PMID:21665749
- Rodriguez-Guisado I, Odriguez-Guisado FCA, Hernandez P, Melgarejo P, Legua R, Martinez R, Marinez JJ (2009). Chemical, morphological and organoleptical characterisation of five Spanish quince tree clones (*Cydonia oblonga* Miller). *Scientia Horticult.* 122(3):491-496. <http://dx.doi.org/10.1016/j.scienta.2009.06.004>.
- Scaramuzzi F (1957). Contributo allo Studio delle cultivar di cotogno da frutto. *Rivista di Ortoflorofruitticoltura Italiana* 41(11-12):575-615.
- Timmer VR, Stone EL (1978). Comparative foliar analysis of young balsam fir fertilized with nitrogen, phosphorus, potassium, and lime. *Soil Sci. Soc. Am. J.* 42:125-130. <http://dx.doi.org/10.2136/sssaj1978.03615995004200010027x>
- Timmer VR, Armstrong G (1987). Diagnosing nutritional status of containerized tree seedlings: comparative plant analysis. *Soil Sci. Soc. Am. J.* 51:1082-1086. <http://dx.doi.org/10.2136/sssaj1987.03615995005100040048x>
- Van den Driessche R (1984). Relationship between spacing and nitrogen fertilization of seedlings in the nursery, seedling mineral nutrition, and outplanting performance. *Can. J. Forest Res.* 14:431-436. <http://dx.doi.org/10.1139/x84-076>
- Weeetman GF, Fournier RM (1986). Construction and interpretation of foliar graphical diagnostic technique. In: Interior forest fertilization workshop, pp. 55-76. Kamloops, British Columbia.
- Yamamoto T, Kimura T, Soejima J, Sanda T, Ban Y, Hayashi T (2004). Identification of quince varieties using SSR markers developed from pear and apple. *Breed. Sci.* 54:239-244. <http://dx.doi.org/10.1270/jsbbs.54.239>