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Spermicidal and antigonococcal effects of tannins from pomegranate rind

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The purpose of this work was to evaluate the spermicidal, antigonococcal and antifertility efficiency of tannins in pomegranate rind (TPR). Spermicidal activity was assessed by a modified version of the Sander-Cramer test and World Health Organization (WHO) standard method. A chromatographic method was applied to screen the spermicidal and antigonococcal ingredients of TPR, and the antifertility effect was investigated using rabbits. Spermicidal effect of TPR was identified both *in vitro* and in rabbit. The minimum effective spermicidal concentration of TPR *in vitro* was 1.25 mg/ml. In antigonococcal process, TPR shows moderate performance. However, TPR terminated the origination of pregnancy significantly in rabbit.

Key words: Spermicide, gonorrhoeae, tannins; pomegranate rind.

INTRODUCTION

With constant increases of global population, it is imperative that birth-control policy should be a major concerned. Even though a variety of contraceptive agents are on sale, their mechanisms of prevenception mainly include antifertilization, anti-implantation and inhibition of ovulation. Some side effects, such as colporrhagia, body weight gain and depressive disorders, which threaten female physiological and mental health, are observed in usage frequently. Owing to the risk of causing irreversible azoospermatism and other adverse reactions, it is difficult to generalize and apply for male contraceptives, such as gossypol. Therefore, it is a stimulating challenge for current pharmaceutical scientists to find and utilize natural products or derivatives as contraceptives with simplicity, low toxicity and hyper efficiency. Neisseria gonorrhoeae infection is a common bacterial of sexually transmitted disease. Though various types of preventive antibiotics have been developed over the last several years, N. gonorrhoeae strains have developed a high level of resistance to several antibiotics because of misuse and irregularity in the treatment (Mbwana et al., 1999). The

treatment has become more complicated particularly due to multiple drug resistance. Pomegranate rind is a shrub, which was distributed originally in Iran and Afghanistan, and introduced into China in the second century BC.

It is cultivated throughout the world, mainly grown in the Near East, India, Spain (southeastern), Israel and the (California). In traditional Chinese United States medicines, Pomegranate Rind has been used to treat dysentery, microbial infections, diarrhea and haemorrhage (Negi et al., 2003). The main content of Pomegranate Rind is tannin, as a kind of polyphenols, which takes 10.4-21.3% in weight. These natural tannins have a variety of biochemical and pharmacological properties (Joseph et al., 2012; Dell'agli et al., 2010). It's probable that some certain actions may occur between tannins and sperm, which based on the common tannin-protein reaction. Recently, the relevant reports showed that tannins in Pomegranate rinds exhibited inhibition on sperm motility (Parkhuyst and Stolzenbberg, 1974). The extract of Pomegranate Rind was recognized useful source of highly active antigonorrhoeal а metabolites (Gibbons, 2005). Pomegranate Rind contain plenty of tannins, thus, this assay was conducted to find scientific evidence of some the tannins for antigonorrhoeal activity. In this paper, the inhibition of

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Pomegranate Rind on sperm motility was determined and the active parts of the tannin in Pomegranate Rind inhibiting the sperm motility were screened in order to find potent inhibition and low side effects active parts, and the antifertility of Pomegranate rinds was tested in rabbits by intravaginal administration. The present study was also aimed at evaluating the antigonorrhoeal activities of six fractions of extracts from Pomegranate Rind (Silva et al., 1997).

MATERIALS AND METHODS

Plant material

Pomegranate Rind were obtained from Xinjiang Province (China) in November 2007 and identified by Professor Hong Zhang (College of Pharmacy, Wuhan University, Wuhan, China).

Apparatus and reagents

Sepabeads HP-20 balsam; Sodium Chloride; Nonoxynol-9 and tannic acid from Chinese Gall were purchased from Sigma Chemical Company St. Louis, USA. Acetone and methanol were of analytical grade. Gas chromatography (GC) agar powder (Oxiod, British), haemoglobin (Oxiod, British), Vitox supplement (Oxiod, British), beef extract, enzymatic digestion casein and amylum solubile.

Animals and gonorrhoeae

Mature New Zealand rabbits (7 to 10 months old) weighing 2.5 to 3.0 kg were purchased from Wuhan Institute of Biologic Products (Wuhan, China). Female and male rabbits were kept under standard environmental conditions, respectively, and fed with diet and water ad libitum. Three WHO control strains B (29501), W1 (29400), W2 (29403) were used in the present study.

Preparation of TPR

Pomegranate Rind was purchased from Wuhan Medicinal Material Corporation. Dried pomegranate rinds were ground in a cutting mill fitted with a 40 mesh screen at the bottom of the cutting chamber. A certain amount of comminuted pomegranate rinds was soaked in distilled water for 30 min. The residua were extracted with $8 \sim 10$ volumes of distilled water for 10 h at room temperature. Then the residua were filtrated out and concentrated by a rotary evaporator at 40°C. The partial concentrated solution was cryodesi ccated and colored solid was obtained for phenolic determination. According to the equation:

$$C = \frac{g}{v} \times 100\%$$

Where c is the concentration of TPR, g represents the weight of pomegranate rinds; v refers to the volume of TPR.

Preparation and fractionation of TPR

10.0 g of comminuted pomegranate rinds were extracted with 3 volumes of 70% acetone (v/v) for 6 h at room temperature, then in

an ultrasonic bath for 0.5 h. After that, the residua were filtrated out and the operations were repeated 3 times. The combined extracts were concentrated by using a rotary evaporator at 40°C under reduced pressure. Then the residue was separated by column chromatography (CC) on HP-20 using distilled water, 10% methanol (v/v), 20% methanol, 40% methanol, 60% methanol, 70% acetone as eluents by turns to yield 6 fractions. Fractions were cryodesiccated and phenolics determined by the method as describe, respectively.

Phenolics determination

Phenolics in the TPR and fractions of pomegranate rinds (FPR) were determined by the method of literature (Taga et al., 1984). The concentrated fractions were filtered through a 0.45 μ m Millipore filter. To 500 μ l filtrate, 500 μ l of Folin–Ciocalteau reagent (50%, v/v) and 10 ml of sodium carbonate (2%, w/v) were added and mixed, respectively. After 2 h, the absorbance of the solution was measured at 750 nm. Quantitation was based on the standard curve of gallic acid (0 to 1.0 mg/ml), dissolved in methanol/water (60: 40, v/v; 0.3% HCl). Phenolic content was expressed as milligrams per gram of equivalent of gallic acid.

Human semen collection

The semen samples provided by medical center of GEN, Renming Hospital of Wuhan University were collected by masturbation from five nondrinker and nonsmoker male volunteers (20 to 25 years) with a mean spermatozoal count of $> 20 \pm 0.5 \times 10^6$ spermatozoa/ml and $> 70 \pm 2\%$ normal sperm morphology (Reynolds and Narang, 1984). The semen characteristics, volume, pH, viscosity and sperm morphology were determined according to World Health Organization guidelines. Samples were collected into a warm glass beaker to avoid cold shock to the sperm. The fresh samples were allowed to liquefy and further investigations were carried out at 35%.

The inhibition of TPR on sperm motility

The experiment of pomegranate rinds on inhibiting the sperm motility *in vitro* was determined by following a modified version of the Sander-Cramer test (Khillare and Shrivastav, 2003). 0.2 ml of liquefied semen was kept in a water bath at 37°C. Different concentrations of TPR in physiological saline (1.0, 2.05 and 4.0%) were prepared by serial dilution and 100 µl of each was mixed with an aliquot of 20 µl of sperm suspension (2.5×10^7 ml⁻¹). Rapidly the mixtures were observed under a phase contrast microscope (×100) for 20 s and analyzed for percentage and grade of sperm motility (Shrabanti et al., 2007). NP-9 was used as reference substance.

The spermicidal activity of TPR

The experiment of inhibiting the sperm motility (Fu et al., 2003) *in vitro* was carried out according to WHO standard method (WHO, 1992). Different concentrations of TPR in sodium chloride (5.0000, 2.5000, 1.2500, 0.6250 and 0.3125 mg·ml⁻¹) were prepared by serial dilution. 0.5 ml of each was mixed with an aliquot of 0.1 of ml sperm suspension. Rapidly the mixtures were observed under a phase contrast microscope (x100) at (25 ± 2) °C for 20 to 180 s and analyzed for percentage and grade of sperm motility. Different concentrations of NP-9 (0.5000, 0.2500, 0.1250 and 0.0625 mg/ml) and sodium chloride were used as control.

Group	Concentration (mg/ml)	Time of agglutination (s)	
TPR A	1.0	30±3.16*	
TPR B	2.0	10±2.98**	
TPR C	4.0	8.0±1.12*	
NP-9	0.15	9.0±2.31*	

Table 1. Agglutination of human semen by pomegranate rinds $(\overline{X}\pm SD)$ (n=20).

Compared with the group of 1.0% concentration. *P<0.05 and **P<0.01.

Antifertility effects of TPR in rabbit by intravaginal administration

For increasing mating rate, adult female rabbits (n=50) were treated by intramuscular injection with 100 μ g of benzestrofol. Different contents of TPR in vaginal suppository (0.0, 0.1, 0.4 and 1.0 g) were put to the vaginal vault as reported (Shrabanti et al., 2007) with a specially made glass tube (n=10, each content). 50 mg NP-9 suppository was applied as positive control. After 15 min, the female rabbits mated with male rabbits only once and the vaginal scraping with plenty of prorsad-mobiling sperm was identified as successful copulation. The litter size and corepus luteum were recorded to calculate the pregnant rate after 15 days.

Culture media and strains

500 g of TM transport medium was contained by 19 g of agar powder, 5 g of haemoglobin and 10 ml of Oxiod Vitox supplements. And the MH transport medium was freshly prepared in our laboratory with 1 g of beef extract, 8.75 g of enzymatic digestion casein and 0.75 g of amylum solubile. The three WHO control strains B (29501), W_1 (29400), W_2 (29403) were dissolved in MH and then cultured on TM medium. All samples against N. gonorrhoeae strains were determined by the agar dilution method. Freeze-fried powders of each extract were dissolved after sterile under uviol lamp (Putnam et al., 1992). The solutions of tested extracts were diluted by two fold serial dilution to make a serial of different concentrations. The stock solutions of antimicrobial agents were prepared using autoclaved GC agar base mixed with the Hemoglobin solution, after cooling to 50 centigrade, supplemented with Oxiod microbial agent. In addition, 18 ml of stock solution was combined with TPR solutions together to product a series of agar plates containing different concentrations of tannin. Fresh overnight colonies were suspended and diluted in sterile beef broth to a known density by using Mc Farland standard method. The density is about 10⁷ cfu of *Gonorrhoeae neisseria* strains per milliliter. Then, 2 ul of beef broths were inculcated on each agar plates. The plates were incubated for 24 h.

Statistical analysis

The *t*-test was used to determine statistical difference. Values of P< 0.05 were considered statistically significant.

RESULTS

Phenolics in TPR

The yield of extract obtained from pomegranate rind using ethanol was 17.23% (w/w). The TPR was proved to

contain 283 ± 4 mg/g total polyphenolics standardized as equivalent of gallic acid. The polyphenolics in different fractions were 385 ± 7 mg/g (20%), 326 ± 3 mg/g (40%), 269 ± 2 mg/g (60%), 235 ± 2 mg/g (70%), 197 ± 2 mg/g (10%) and 161 ± 8 mg/g (H₂O), respectively.

The spermicidal efficacy of TPR

Pomegranate rinds were found to have the role of agglutination, causing sperm immobilization. When agglutinating, the semen aggregated randomly. The minimum concentration of Pomegranate rinds that caused 100% immobilization within 20 s with no revival of motility at 37°C was considered to be the minimal effective concentration (MEC). The MEC of Pomegranate rinds was 2.0%. While the concentration was lower than 2.0%, the sperm lost some motility; while the concentration was more than 2.0%, the sperm lost completely motility (Table 1).

The active parts of TPR

After the sperm were mixed with TPR, the microscopy imaging showed that the sperms expanded quassated or head-tail broken and agglutinated. The sperms were active at low TPR concentration, but with the concentration increasing, they became to immobilizate significantly, from active to trembling and lost the activities eventually. Statistical results indicated that the 20% CH₃OH eluting part had the highest inhibiting activity, and the MEC in 20 s was 1.25 mg/ml (Table 2). This eluting part may contain some compounds which have strong inhibition on sperm motility.

Antifertility efficiency in rabbits

All samples in control group were pregnant. The 1.0 g of TPR group was proved to process obviously contraceptive effect. Compared with control group, 100% of contraception was found in the 1.0 g of TPR and NP-9 group. The 0.4 and 0.1 g of TPR group showed low conception rate, 10 and 40%, respectively (Table 3). The minimum inhibitory concentration (MIC) was read as the

David		Action time (s)			
Drug	mg∙ml ⁻¹ ──	20	60	180	
NP-9	0.50000	-	-	-	
	0.25000		-	-	
	0.12500	±	-	-	
	0.06250	+	+	±	
	0.03125	+	+	+	
H ₂ O eluting part	5.00000	-	-	-	
	2.50000	±	-	-	
	1.25000	±	±	+	
	0.62500	+	+	+	
	0.31250	+	+	+	
10% CH ₃ OH eluting part	5.00000	-	-	-	
31 31 31 31	2.50000	±	-	-	
	1.25000	±	±	+	
	0.62500	+	+	+	
	0.31250	+	+	+	
	0.31230	Т	T	T	
20% CH₃OH eluting part	5.00000	-	-	-	
	2.50000	-	-	-	
	1.25000	-	-	-	
	0.62500	±	-	-	
	0.31250	+	±	-	
40% CH₃OH eluting part	5.00000	-	-	-	
	2.50000	-	-	±	
	1.25000	±	±	+	
	0.62500	+	+	+	
	0.31250	+	+	+	
60% CH₃OH eluting part	5.00000	_	_	-	
our of a charge part	2.50000	_	-	±	
	1.25000	±	±	+	
	0.62500				
	0.31250	+ +	+	++	
	0.31250	Ŧ	+	Ŧ	
70% CH ₃ COCH ₃ eluting part	5.00000	-	-	-	
	2.50000	-	-	-	
	1.25000	-	±	+	
	0.62500	+	+	±	
	0.31250	+	+	+	
Physiological saline	-	+	+	+	

Table 2. The effect of several eluting parts of pomegranate rind with different concentrations against the motility of the sperm (n=20).

-: Sperm is immobile; ±: 90% semen lost motility; +: sperm is mobile.

Group	Dose	Pregnant / total rabbits	No. of implantation(X±SD)	
Control	-	10/10	4.4±0.79	
Pomegranate rinds	0.1 g	4/10	0.9±1.29*	
Pomegranate rinds	0.4 g	1/10	0.2±0.63*	
Pomegranate rinds	1.0 g	0/10	0**	
NP-9	50 mg	0/10	0**	

Table 3. Effect of CP suppository on antifertility activity in rabbits.

Compared with the control group *P<0.05 and **P<0.01.

Table 4. The minimum inhibitory concentration mg/ml of tannins in Punica granatum L. on three kinds of G. neisseria strains.

Composition	H ₂ O Eluting part	10% CH ₃ OH eluting part	20% CH ₃ OH eluting part	40% CH₃OH eluting part	60% CH ₃ OH eluting part	70% CH₃OH eluting part
B (29501)	2.5	1.25	2.5	5.0	5.0	2.5
W1 (29400)	2.5	1.25	1.25	5.0	5.0	2.5
W2,3(29403)	2.5	2.5	1.25	2.5	5.0	2.5

lowest concentration of antibiotic inhibiting visible growth. The results of the antigonorrhoeal assays were recorded in Table 4.

DISCUSSION

In the spermicidal experiments, first pomegranate rind was found to inhibit the sperm motility by following a modified version of the Sander-Cramer test in vitro; then the spermicidal part of tannin in pomegranate rind was screened by WHO standard method, and the 20% CH₃OH eluting part had the highest inhibiting activity. A dose- and time-dependent reduction in sperm motility was observed by adding TPR and TPR to semen samples (Tables 1 to 3). All of them were found to inhibit sperm motility, with the concentration increasing or the time prolonging, the inhibition became stronger and stronger. The MEC of NP-9 on inhibiting sperm activity in 20 s is 0.125 mg·ml⁻¹, consistent with the reports (Chantler et al., 1992). The eluting parts of tannin in pomegranate rind were found to inhibit the sperm activity significantly and the MEC of the 20% CH₃OH eluting part in 20 s was 1.25 mg·ml⁻¹, which was less than that of NP-9. It is feasible to screen a new spermicide from pomegranate rind in further studies. However, the mechanism by which dermaseptin exerts its rapid spermicidal action of pomegranate rind is not known.

The microscopy imaging showed that the sperm quassated or head-tail broken and agglutinated. The phenomenon suggested that tannins from pomegranate rinds may kill the sperm through destructing the substructure and organization of the sperm. Furthermore, tannin-protein reaction indicated that TPR could congeal seminal plasma protein *in vivo* to inhibit mobility of semen, react with sperm membrane protein for decreasing the stability and integrity and deactivate the metabolic enzyme (both the metal center and protein) to interrupt the energy-supplying chain. Further researches would focus on the precise spermicidal mechanism of TPR by transmission electron microscope and concrete process which TPR adheres the spermatic protein (Data not shown). The results of vaginal suppository in rabbits exhibited that when the content of TPR was more than 0.4 g, It showed confirmed forceful antifertility efficiency and when the content is 0.1g, it also occurred the activity to decrease the number of implantation $[0.9 \pm 1.29, (Table$ 3)]. The contraceptive mechanism of vaginal suppository in rabbits may be: agglutination of TPR to semen; destructing seminal plasma protein; changing the internal environment which is essential to sperm; dispersing sperm to free state and decreasing the spermatic mobility. Respecting the tolerance of rabbit sperm is double compared with human semen, so it seems that satisfied effects may be observed in clinical trail.

Gonorrhoea is a sexually transmitted disease which is caused by the bacteria, N. gonorrhoeae. So far, appropriate the clinical treatment of gonorrhea mainly uses antibiotics. But disease control is also problematic, because of high rates of changes by way of both spontaneous mutation and misuse of antibiotics in settings (Simonsen et al., 2004). The data from recent report showed that by now in some cities, more than 81.7% of all N. gonorrhoeae isolates were resistant to Penicillin G. The increase has been occurring rising trends gonococcal resistance with the third-generation of cephalosporin and fluoroquinolones (Tapsall, 2005). As is reported that, the tannin in pomegranate rind can not only inhibit the sperm motility, but also has the antigonorrhoeal effects. So it is feasible to develop TPR as a new vaginal

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contraceptive, which has the role of spermicidal and antibacterial agents.

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REFERENCES

- Chantler E, Fisher H, Solanki S, Elstein M (1992). Quantification of the *in vitro* activity of some compounds with spermicidal activity. Contraception, 46: 527-536.
- Dell'agli M, Galli GV, Bulgari M, Basilico N, Romeo S, Bhattacharya D, Taramelli D, Bosisio E (2010). Ellagitannins of the fruit rind of pomegranate (*Punica granatum*) antagonize *in vitro* the host inflammatory response mechanisms involved in the onset of malaria. Malar J., 9: 208.
- Gibbons S (2005). Plants as a source of bacterial resistance modulators and anti-infective agents. Phytochem. Rev., 4: 63-78.
- Joseph MM, Aravind SR, Varghese S, Mini S, Sreelekha TT (2012). Evaluation of antioxidant, antitumor and immunomodulatory properties of polysaccharide isolated from fruit rind of Punica granatum. Mol. Med. Rep., 5(2): 489-496.
- Khillare B, Shrivastav TG (2003). Spermicidal activity of Azadirachta indica (neem) leaf extract. Contraception, 68: 225-229.
- Mbwana J, Mhalu F, Mwakagile D, Masesa J, Moshiro C, Sandstrom E (1999). Susceptibility pattern of *Neisseria gonorrhoeae* to antimicrobial agents in Dar es Salaam. East Afr. Med. J., 76: 330-334.

- Negi PS, Jayaprakasha GK, Jena BS (2003). Antioxidant and antimutagenic activities of pomegranate peel extracts. Food Chem., 80: 393-397.
- Putnam SD, Lavin BS, Stone JR, Oldfield EC, Hooper DG (1992). Evaluation of the standardized disk diffusion and agar dilution antibiotic susceptibility test methods by using strains of *Neisseria gonorrhoeae* from the United States and Southeast. Asia J. Clin. Microbiol., 30: 974-980.
- Reynolds TR, Narang BS (1984). In: Cheeseborough M, editor. Medical Laboratory manual for tropical countries. Kent: Butterworth and Co. Ltd., 2: 186-187.
- Shrabanti K, Shampa B, Debayan M, Heramba NR, Smritinath C, Syed N, Kabir, Sukdeb B, Nirup BM (2007). *Chenopodium album* seed extract: a potent sperm-immobilizing agent both *in vitro* and *in vivo*. Contraception, 75: 71-78.
- Silva O, Ferreira E, Vaz PMV, Gomes ET (1997). Guinea-Bissau's plants *in vitro* susceptibility studies on *Neisseria gonorrhoeae*. Int. J. Pharmacogn., 53: 323-328.
- Simonsen GS, Tapsall JW, Allegranzi B (2004). The antimicrobial resistance containment and surveillance approach—A public health tool. Bull. World Health Organ, 82: 928-934.
- Stolzenbberg SJ, Parkhuyst RM (1974). Spermicidal actions of extracts and compounds from phytolacca do decandra. Contraception, 23 (6): 135.
- Tapsall JW (2005). Antibiotic resistance in *Neisseria gonorrhoeae*. Clin. Infect. Dis., 41(Suppl. 4): S263-268.
- WHO (1992). The laboratory manual of the examination of human semen and semen-cervical mucus interaction. 3th ed. UK: Cambridge University Press, p. 7.
- Xiaomin F, Qianli L, Aihua J (2003). Study on the spermicidal and antibacterialeffect of Chlorhexidine Diacetate *in vitro*. Chin. J. Mod. App. Pharm., 20(20): 97-99.