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Full Length Research Paper

Serjania erecta Radlk. improves endothelial function in isolated aortic rings

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Arterial hypertension presents a high incidence in the population, and it is a chronic degenerative disease. The roots of Serjania erecta Radlk. popularly known as "cipó-cinco-folhas", are used as a decoction against hypertension. This plant belongs to the Sapindaceae family and is plentiful in the Brazilian swamps. The present study was designed to study the effects of daily intake of S. erecta root decoction on mean arterial pressure (MAP), heart rate (HR) and vascular reactivity in thoracic aorta rings in spontaneous hypertensive rats (SHR) and Wistar-Kyoto rats (WKY). The decoction was prepared from the powder of the roots at a final concentration of 5% in solution, which is an appropriate level of administration according to some modifications of the dessert test, for 32 consecutive days. The MAP and HR were measured directly and presented no changes between groups. Vascular reactivity was tested by concentration-effect curves for noradrenaline obtained from thoracic aorta rings with (+E) and without (-E) endothelium. In SHR, treatment reduced vascular reactivity (+E) (0.92 \pm 0.18 vs. control SHR: 1.68 \pm 0.26 g), and the presence of L-NAME abolished this effect (treated SHR: 1.94 ± 0.30 g). The aorta rings (-E) exhibited no changes. In conclusion, the decoction of S. erecta roots is effective in increasing endothelial function in SHR rats. However, under these conditions, the decoction did not change the MAP and HR. Therefore, this part of the plant promotes nitric oxide (NO) production by the endothelium, which may promote blood flow to the tissues; thus, its use is justified as adjunct for the treatment of hypertension.

Key words: Serjania erecta Radlk., hypertension, endothelium function, nitric oxide.

INTRODUCTION

Hypertension is considered a chronic degenerative disease; it presents no exacerbated symptoms and

has serious consequences if not treated, targeting organs such as heart, kidney, brain and eyes. High

*Corresponding author. E-mail: jdegobbi@ibb.unesp.br. Tel: +55 (14) 3880-0319/0330. Fax: +55 (14) 3815-3744. Author(s) agree that this article remain permanently open access under the terms of the <u>Creative Commons Attribution</u> License 4.0 International License pressure is considered one of the most common causes of morbidity in adults (Nobre, 2010; Ripley et al., 2015). A major problem in the treatment of hypertension is poor patient adherence. It has been estimated that 30 to 50% of patients discontinue therapy or take only one of the medicines prescribed within the first year of the treatment (Car et al., 1991; Ross et al., 2004; Wilson et al., 2005). Generally, hypertensive elderly patients report that the need of medication is only associated with the presence of symptoms (Carvalho et al., 1998). Patients still hold beliefs considering the benefits and side effects of a prescribed medicine (Horne and Weinman, 1999; Magadza et al., 2009). Some elderly report dislike of allopathic medicines, believing that their chemicals are very bad for the body, producing many side effects (Carvalho et al., 1998). In the countryside of Sao Paulo State, Brazil, 15% of hypertensive patients reported the use of home medications for the treatment of hypertension, and 45% of these patients reported the use of teas (Boldo, lemongrass, rosemary, garlic water) as supporting treatments, especially when they realize their arterial pressure is elevated (Péres et al., 2003). In this sense, validating plants as potential hypertension treatments might be an improvement by promoting better compliance for hypertension treatment.

The use of plants for primary healthcare in underdeveloped and developing countries ranges from 65 to 80%, according to the World Health Organization. This is mainly due to poverty and lack of access to synthetic drugs (Akerele, 1993). The Mato Grosso do Sul State, with savannah and wetland, typical Brazilian biomes, has great potential to yield compounds with pharmacological activities (Proença et al., 2000). One of the species that requires attention is Serjania erecta Radlk., Sapindaceae family, known as "timbó-bravo", barkcloth, vine-of-timbó, vine-five plants or five leaves. This plant is an erect shrub, half climbing, 1 to 3 m tall, and blooms in January (Pott and Pott, 1994). The fruits of this genus are edible (Pott et al., 2004), and the oil from its seeds contains fatty acids, with a great predominance of eicosanoic acid (Mayworm and Salatino, 1996). Ethnobotanical studies, reflecting the beliefs and cultural aspects of the population, indicated that the tea from the leaves of S. erecta acts against gastric ulcers and the decoction of the roots acts against hypertension (Guarim, 1996, 2000).

Pharmacological evaluation of hydroalcoholic extracts of the stem and leaves of *S. erecta* showed topical antiinflammatory activity in mice (Gomig et al., 2008). Methanol and chloroform extracts of the stem and leaves of *S. erecta* have gastroprotective action, showing no acute toxicity (Castelo et al., 2009). However, the possible effect of the roots of this plant against hypertension has not been tested via controlled studies in laboratory animals. Therefore, the aim of this study was to explore the chronic administration of the decoction of the *S. erecta* roots on the mean arterial pressure and heart rate of spontaneous hypertensive rats (SHR) and Wistar-Kyoto rats (WKY, control for SHR), as well as the vascular reactivity of these animals. In addition, this study also evaluated these animals' exploratory behaviour in the open field and anxiety in the elevated plus maze.

METHODOLOGY

Animals

Spontaneously hypertensive rats (SHR, ~200 g initial) and their control, Wistar-Kyoto rats (WKY, ~200 g initial), acquired from the CEMIB - Multidisciplinary Center for Biological Research in the Area of Science in Laboratory Animals, UNICAMP, were used. The rats were kept at a controlled temperature $(23 \pm 2^{\circ}C)$ and subjected to a light:dark cycle of 12:12 h. They were kept in individual cages with free access to tap water and food (Biotec 9301, Biobase®). T experiments took place between 8:00 and 16:00 h. The procedures were approved by the Ethics Committee in Animal Experimentation of the Institute of Biosciences of Botucatu, UNESP (32/08-CEUA).

Plant collection

The roots of *S. erecta* were collected in the city of Aquidauana, MS, Brazil, in September 2008; they were identified by A. Pott and deposited in the Herbarium of Embrapa Beef Cattle, Campo Grande, MS. The roots were washed and dried in the shade. They were crushed in a knife mill at the Institute of Chemistry of the Federal University of Mato Grosso do Sul (UFMS), Campo Grande-MS, to produce a coarse powder.

For phytochemistry analysis, standardized tests were used to assess which secondary metabolites were present in the roots using methodology described by Mattos (1997). In the literature, there are few phytochemical studies on this species, especially for the roots. The phytochemistry concerning the decoction is still missing.

Decoction preparation and administration

The decoction was prepared according to the 2nd edition of the Pharmacopoeia of the United States of Brazil (1959), which yields a final concentration of 5% plant extract. The decoction was administered using a modified dessert test according to Johnson and Schwob (1975). The original dessert test consists of offering a condensed milk solution at 30%, which is highly palatable and quickly ingested by rats. In the present study, the decoction was prepared in the morning with a volume of water that was 15% less than that required for the final concentration. After cooling, the remaining 15% volume was made up with condensed milk, which also produces a palatable solution. Thus, the final suspension of *S. erecta* roots was adjusted to 5% plant extract.

Each animal was offered 10 ml of this solution in graduated burettes placed in front of the cages at approximately 09:00. The solution was successfully ingested in a maximum time of 15 min for the rats. The percentage of treatments successfully administered was 100%. On rare occasions (twice), a rat took 45 min to drink 10 ml of the sweetened solution. Thus, the rats ingested equal amounts of decoction and the entire portion of the 5% of the plant in a relatively short period of time. In this way, the solution was not exposed to light for a long time, limiting potential changes in its active ingredients.

Half of the rats of each strain received a decoction dissolved in

the condensed milk, and the other half received the same volume of a 15% condensed milk solution. The treatment was performed over a period of 32 consecutive days. The animals had free access to water and food at all times, and were free to choose whether to ingest the solution or not. In order to ensure that the daily condensed milk intake did not alter the blood glucose of the animals after treatment, a small sample of blood was collected prior to euthanasia (approximately 2 h after administration of the decoction) and before collection of thoracic aortic rings. Blood glucose was analysed using glucose dosage strips (Prestige Smart System®). In this study, gavage procedure was avoided to minimize any further stress in the SHR, which would increase the blood pressure, especially for a chronic treatment.

Drugs

The drugs used included norepinephrine bitartrate, acetylcholine chloride, sodium nitroprusside and methyl ester of N-nitro-L-arginine (L-NAME; all purchased from Sigma Chemical Co., St. Louis, Missouri, USA). All drugs were dissolved in Krebs-Henseleit solution prepared immediately before use.

Exploratory and anxiety tests: Open field (OF) and elevated plus maze (EPM)

The treated SHR presented an apparent docility. Another point to consider in hypertension is its association with anxiety states. Increase in anxiety enhances the risk of hypertension (Ginty et al., 2013; Garcez et al., 2014). Thus, the possibility that the decoction might act centrally as an anxiolytic was tested. Following 30 days of treatment, the rats were exposed to tests in the OF and EPM. On the morning of the test, at approximately 8:00 h, the animals received the decoction (or control) as usual and were transferred to the test room for adaptation.

The tests were conducted between 13:00 and 16:00 h. The animals were exposed for 3 min in the OF, and parameters such as rearing and exploratory activity and the number of squares that were crossed were analysed. Then, the animals were placed in the EPM, and spatiotemporal measures of avoidance of open arms (time spent in the arms) and total number of entries into the arms of the apparatus were evaluated in order to assess anxiety (Pellow et al., 1985).

Direct measurement of blood pressure and heart rate

Mean arterial pressure (MAP) and heart rate (HR) were recorded in non-anesthetized rats with free movement. A day before the recordings, the animals were anesthetized with ketamine (80 mg/kg, Dopalen®, Vetbrands) plus xylazine (20 mg/kg, Anasedan®, Vetbrands). Then, they underwent femoral artery cannulation with a PE 10 polyethylene tube (0.024" OD × 0.011" ID), which was connected to a PE 50 polyethylene tubing (0.038" Od × 0.023" ID). The catheter was conducted subcutaneously and was externalized by the animals' neck, allowing free movement.

On the next day, the PE 50 tube was connected to a pressure transducer (TSD104A, Biopac Systems) coupled to a multichannel recorder (Model M100A-CE, IC Biopac Systems, Santa Barbara, California), which was connected to computer for data acquisition. A solution of heparin (10 UI/mI) in a volume of 0.2 ml was used to flush the catheters only once in a previous connection to the transducer to facilitate the blood flow and avoid clot formation in the catheter. The recording was made for approximately 40 min.

Assessment of vascular reactivity test in vitro

After the blood pressure measurements, the animals were anesthetized with pentobarbital sodium, 50 mg/kg intraperitoneally, and killed by decapitation. The animals underwent thoracotomy and then proceeded to the isolation of thoracic aorta. After removal of the connective tissue, the aorta was divided into four rings. Two of the rings, selected randomly, had the endothelium removed mechanically. The rings were mounted in organ baths containing modified Krebs-Henseleit solution (composition in mM: NaCl 113.0; KCl 4.7, CaCl₂ 2.5, KH₂PO₄ 1.2, MgSO₄ 1.1, NaHCO₃ 25.0; glucose 11.0, ascorbic acid 0.11). The nutrient solution was maintained at 37.0 ± 0.5°C and saturated with carbogen (95% O2, 5% CO2). The rings were connected to isometric force transducers coupled to a physiograph (Ugo Basile). The settling time was 1 h under an initial tension of 1.5 g maintained throughout the experiment. A resting tension of 1.5 g is optimal in producing the maximum contraction to noradrenaline (NA) in the aorta of adult rats (Cordellini et al., 2006). The rings of thoracic aorta with endothelium (+E) and without (-E) were studied in parallel (Cordellini et al., 1990, 2006). Cumulative concentration-effect curves to NA were constructed from the tissue responses to NA. The increased doses produced remained stable over approximately 40 to 60 s (Cordellini et al., 1990, 2006).

The responses to NA were also examined in the absence and presence of L-NAME (3×10^{-4} M), an inhibitor of nitric oxide synthase, which was added to the preparations in the last 30 min of stabilization and remained in contact with them until the end of the experiments. Following completion of the determination of the concentration-effect curves, single doses of acetylcholine (10^{-6} M) and sodium nitroprusside (10^{-4} M) were used to test the integrity of the endothelial cell and smooth muscle layer, respectively. Vascular reactivity was assessed by the change of the maximum response (g voltage) and power (50% effective concentration - EC₅₀) of the NA in individual experiments. The EC₅₀ is defined as the molar concentration of agonist needed to achieve an effect equal to 50% of maximal effect.

Wet weight of the organs and microscopic examination of the liver

The animals were weighed every 2 days until the end of treatment. After euthanasia, the internal organs were removed for macroscopic verification. Immediately after the sacrifice, the organs were weighed. The livers were stored for histological evaluation and sectioned into three portions to obtain tissue samples. The samples were kept in 10% formalin solution for 24 h, washed in tap water, and stored in 70% alcohol until paraffin embedding; 20-mm thick sections were made, and the slices were stained with hematoxylin/eosin.

Statistical analysis

Data were compared by one way analysis of variance (ANOVA) followed by the Tukey's post-test for the presence of significance (Sigma Stat 3.5). For behavioural data, non-parametric Mann-Whitney test was used. Results were expressed as the mean \pm standard error of mean (SEM). The significance level was 5%.

RESULTS

Phytochemical evaluation

The qualitative phytochemical analysis of the *S. erecta* root detected the presence of saponins (foaming test),

Table 1. Parameters analyzed by the open field test.

Parameter	Treated SHR n = 10	Control SHR n = 12	Treated WKY n = 13	Control WKY n = 10
Number of squares	26 (24 - 35)	28 (22 - 42)	31 (16 - 50)	45 (34 - 45)
Rearing	18 (14 - 25)	14 (13 - 20)	18 (13 - 30)	10 (5 - 25)

Results are expressed as median and interquartile range (Q1 to Q3), Mann-Whitney test showed no differences between groups or treatments for the OF (p > 0.05).

Table 2. Parameters analyzed by the elevated plus maze.

Parameter	Treated SHR n = 10	Control SHR n = 12	Treated WKY n = 13	Control WKY n = 10
ΤΟΑ	110 (88 - 116)*	108 (45 - 146)*	17 (8 - 55)	7 (0 - 17)
TCA	170 (153 - 163)*	145 (115 - 120)*	246 (220 - 233)	248 (243 - 273)
EOA	5.5 (5 - 5.8)	7 (6.5 – 7.0)	3 (1 - 4)	3 (2 - 5.1)
ECA	6.5 (6 - 6.9)	6 (5.7 - 5.9)	2 (2 - 2)	5 (3 - 5)

TOA: Time spent in the open arms in seconds; TCA: time spent in the closed arms in seconds; EOA: entry number in the open arms; ECA: entry number in the closed arms. Results are expressed as median and interquartile range (Q1 to Q3), Mann-Whitney test demonstrated behavioural difference between strains for the elevated plus maze in relation to TOA (p = 0.01), and TCA (p < 0.05). *Different from WKY strain.

Table 3. Mean arterial pressure and heart rate in SHR and WKY, treated and control.

Group	Treated SHR n = 6	Control SHR n = 6	Treated WKY n = 5	Control WKY n = 5
MAP (mmHg)	166.1± 5.3*	170.1 ± 6.4*	125.5 ± 8.2	119.6 ± 6.3
HR (bpm)	372.2 ± 20.1	368.3 ± 18.1	356.5 ± 19.3	359.4 ± 21.3

MAP: Mean arterial pressure; HR: heart rate. Results are expressed as mean \pm standard error of the mean. One way analysis of variance (ANOVA) showed differences between strains (p < 0.01). *Different from WKY strain.

flavonoids (magnesium chip and hydrochloric acid test), tannins (reaction of ferric chloride and 2% gelatin in drip test), catechins (brown-yellow colour after acid-base reaction) and cardiac glycosides (Kedde reaction).

Administration of decoction

The animals' body weight was similar in the two groups throughout the experiment. The blood glucose levels analysed between groups were treated SHR: 97.0 ± 0.5 vs. control SHR: 96.5 ± 0.5 mg/dl; treated WKY: 96.6 ± 0.6 vs. control WKY 96.7 ± 0.6 mg/dl.

Exploratory and anxiety tests: OF and EPM

In the OF, we were interested in studying the exploratory behaviour; a decrease in this behaviour by the decoction would be considered a signal of central effects, for instance, a sedative action. There were no differences between the experimental groups (p > 0.05) (Table 1). The EPM considers the time spent in each part of the

apparatus, that is, the closed or open arms, in order to infer the level of anxiety. There were behavioural differences between strains in the time spent in the open arms (p = 0.01), and the treatment did not alter this difference (Table 2).

Blood pressure and heart rate

Table 3 presents the data obtained for treated and untreated SHR and WKY. There were differences between strains in MAP, evidencing hypertension for SHR (p < 0.01). The treatment was not able to change this difference. The heart rate was normal in both groups and was not altered by the treatment (p > 0.05).

Vascular reactivity tests

Regardless of endothelium integrity, the reactivity to NA in rings of thoracic aorta was similar in SHR and WKY. Treatment with *S. erecta* in WKY rats did not cause any change in reactivity to NA for either (+E) or (-E)

Table 4. Concentration-effect to noradrenaline in the absence or presence of L-NAME obtained in the thoracic aorta ring with or without endothelium of the WKY and SHR rats treated or not.

Aortic rings condition	Treated SHR (n = 7)	Control SHR (n = 8)	Treated WKY (n = 5)	Control WKY (n = 5)
NA (+E)	0.92 ± 0.18*	1.68 ± 0.26	1.78 ± 0.37	1.83 ± 0.49
L-NAME (+E)	1.94 ± 0.30	1.60 ± 0.13	2.03 ± 0.37	2.44 ± 0.11
NA (-E)	2.65 ± 0.45	2.10 ± 0.20	2.35 ± 0.30	3.08 ± 0.28
L-NAME (-E)	1.88 ± 0.17	1.77 ± 0.21	2.28 ± 0.15	2.73 ± 0.21

NA: Noradrenaline; (+E): with endothelium; (-E): without endothelium. Data is given in grams (g) of contraction. Results are expressed as mean \pm standard error of the mean. One way analysis of variance (ANOVA) followed by the Tukey's post test showed differences for (+E) groups.*different from control SHR and WKY strain (p = 0.03), different from NA (+E)/treated SHR (p = 0.02).

Table 5. Wet weight of the organs and body weight at the end of the treatment.

Organs	Treated SHR (n = 9)	Control SHR (n = 10)	Treated WKY (n = 7)	Control WKY (n = 6)
Heart	0.409 ± 0.019	0.407 ± 0.007	0.310 ± 0.013	0.300 ± 0.011
Lungs	0.521 ± 0.017	0.483 ± 0.032	0.461 ± 0.021	0.450 ± 0.031
Stomach	0.515 ± 0.018	0.506 ± 0.015	0.490 ± 0.029	0.548 ± 0.037
Spleen	0.189 ± 0.006	0.209 ± 0.008	0.258 ± 0.005	0.239 ± 0.002
Liver	4.193 ± 0.162	4.150 ± 0.136	4.146 ± 0.139	4.085 ± 0.190
Kidneys	0.409 ± 0.019	0.409 ± 0.019	0.789 ± 0.024	0.831 ± 0.027
Adrenal	0.023 ± 0.002	0.025 ± 0.002	0.016 ± 0.003	0.022 ± 0.002
Body weight (g)	334.4 ± 6.1	327.9 ± 4.3	405.3 ± 10.1	431.2 ± 12.8

The organ weight was corrected for each 100 g of body weight. Results are expressed as mean \pm standard error of the mean. One way analysis of variance (ANOVA) showed no difference between the weights of organs for each SHR or WKY, treated or control, p > 0.05.

(Table 4). However, after treatment, there was a decrease in the maximum response to NA in aorta (+E) in SHR (treated SHR: 0.92 ± 0.18 vs. control SHR: 1.68 ± 0.26 g, p = 0.02), which was abolished by the presence of L-NAME, an inhibitor of nitric oxide synthase (treated SHR: 1.94 ± 0.30 g, p = 0.03).

Wet weight of organs and microscopic examination of the liver

The wet weight of the organs was similar in both strains for both treated and control animals (Table 5). Macroscopically, there were no changes in internal organs after chronic administration of the decoction. Histological sections of the liver tissue of the animals showed no microscopic changes, such as fibrosis or preneoplastic lesions, after 32 days of treatment with the decoction.

DISCUSSION

The present results show that treatment with a decoction of *S. erecta* roots did not reduce blood pressure in SHR

or WKY. However, chronic treatment was able to reduce the vascular reactivity to NA in treated SHR. In addition, this reduction may have been mediated by the activation of endothelial nitric oxide (NO). NO participation was indicated by the fact that L-NAME, a NO synthase inhibitor, reversed the reactivity to NA in the rings of the thoracic aorta (+E) in treated SHR. Treatment with the decoction did not change any anxiety parameters. In addition, chronic treatment with the decoction of *S. erecta* roots produced no macroscopic changes in the organs studied in either strain. The treatment did not alter the microscopic tissue structure of the liver, the primary metabolic organ of the body, which might indicate that this part of the plant is not toxic under these conditions.

The hypertension in SHR is attributed to a polygenic effect due to either neural or vascular disorders (Lerman et al., 2005). SHR is an interesting model in which to study possible effects of decreasing blood pressure levels because many features of hypertension in this strain are similar to those found in most cases of systemic hypertension in humans, that is, essential hypertension. For instance, this model is associated with cardiac hypertrophy, endothelial dysfunction (with increased vascular reactivity) and impaired renal function (Lerman et al., 2005; Li and Bukoski, 1993; Li et al.,

Naseem, 2015: 2005). An imbalance in the production or inactivation of NO contributes to the development and persistence of high blood pressure as well as the development of complications of hypertension (Higashi et al., 2002; Naseem, 2005; Panza et al., 1995). Thus, even though the MAP of treated SHR in this study were not reduced, the decoction of S. erecta reduced vascular reactivity. The present study shows that in treated SHR, the endothelial function is restored, as the maximum response to NA is reduced in thoracic aorta rings (+E) in treated SHR without L-NAME as compared to control SHR.

The presence of L-NAME in treated SHR increased reactivity to NA in aorta rings (+E), compared to the same condition in the absence of L-NAME. No change in aortic response to NA was observed between SHR and WKY (-E). In this context, the observed alterations in the endothelial pathway of the aorta from treated SHR, but not treated WKY, were not surprising. There have been described, among other features, an increased synthesis of vasoconstrictor cyclooxygenase products and superoxide anions in vascular tissue of SHR (García-Redondo et al., 2009; Gao and Lee, 2001). In order to counterbalance these vasoconstrictor factors, an increase in NO synthesis could be observed in the vascular tissue of SHR compared to WKY (Puzserova et al., 2014). In this context, S. erecta treatment may reduce the SHR-increased vasoconstrictor factor pathway production and thus unveil the NO hyperactivation in SHR. NO is produced within the endothelium and then activated by intracellular mechanisms underneath vascular smooth muscle cells (Archer et al., 1994). The decoction of S. erecta roots must contain a factor that favours these mechanisms and consequently increases NO supply to the smooth muscle of blood vessels, contributing to the decrease in vascular reactivity.

The role of NO is attributed to regulation of the mechanical properties of the arteries, in particular the great arteries, promoting aortic compliance (Joannides et al., 1997). The increase in the compliance of the aorta favours the distribution of blood microcirculation (Joannides et al., 1997). Thus, even though the MAP was not reduced in this study, greater endothelial function in the aortic rings can translate into improved blood flow to the microcirculation, which is very beneficial for long-term tissue perfusion. It is worth noting that we used a relatively small dosage of the substance, 0.5 g of crude root administered only once a day. The administration of the decoction at a higher frequency than was used in the present study could decrease the mean arterial pressure. Normally, teas and decoctions are taken in 3 doses daily. In the future, other studies may better delineate this issue.

The Sapindaceae family is a rich source of secondary metabolites. Studies by Schmitt and Dirsch (2009) described the roles of various plant secondary meta-

bolites favouring the production of NO in endothelial and cooperating endothelium-dependent vasodilation. This action occurs predominantly because of flavonoids, such as those found in S. erecta (Chan et al., 2000). Phytochemical analysis of S. erecta roots detected the presence of saponins, flavonoids, tannins, catechins and cardiac glycosides. Recent studies of the ethanol extract of the roots of S. erecta showed the presence of flavonoid glycosides derived from kaempferol: kaempferol-3-O-α-L-rhamnopiranoside and kaempferol-3- $O-\alpha$ -L-rhamnopiranoside-(1 to 6)- β -D-glucopyranoside (Cardoso et al., 2013). In an aortic ring assay used to check the forward vascular reactivity to 5 different bioactive compounds, Leeva et al. (2010) showed that kaempferol has important vasodilator activity via activation of NO. Moreover, Gasparotto et al. (2011) found that intravenous injections of kaempferol in anesthetized SHR were not able to produce a hypotensive effect. These results are similar to the present results, where chronic treatment with S. erecta exerted a vasodilator effect in aortic rings but produced no reduction in blood pressure. Because the findings of this study result from the administration of a decoction and not an isolated extract, we cannot say which substance is responsible for the positive effect on endothelial function, or whether there is a synergistic effect of various substances found in the decoction, but kaempferol seems to be a candidate for these actions. It is worth highlighting that there is an association between enhanced anxiety and high blood pressure values (Ginty et al., 2013: Garcez et al., 2014). Because greater interactivity and a certain docility of SHR treated with the decoction were observed during the treatment, we speculated that the decoction might have an anxiolytic or sedative effect. Using a combination of the OF and EPM tests, these behaviours may be inferred.

The decoction did not cause any differences in exploratory activity; the rearing and the number of squares crossed were similar between strains and treatment. The data for EPM were also similar within each strain. The SHR did not fear spending time in the open part of the apparatus, and the treatment did not change this. However, the WKY (treated or not) spent more time in the closed arms. The SHR strain has symptoms that are similar to those of attention deficit disorder, that is, hyperactivity and impulsivity, compared to WKY (Sagvolden, 2000; Sagvolden et al., 2005). Such symptoms may explain the apparent greater interactivity and docility of animals during treatment. On the other hand, WKY exhibits behavioural instability. Thus, the decoction of S. erecta root in these conditions did not cause changes in exploratory activity parameters or anxiety. The chronic administration of the decoction did not produce any macroscopic changes in the internal organs or even any microscopic changes in the liver. Although toxicity tests are far more complex, involving measurements of the serum concentrations of some

enzymes, our results may suggest that this part of the plant has no apparent toxic effects on organ structure. These findings are a promising sign that this treatment does not have serious side effects.

Conclusion

The present results showed a reduction in vascular reactivity produced by the *S. erecta* root decoction in SHR rats, indicating that this part of the plant might promote blood flow through microcirculation. Thus, conventional wisdom seems to be accurate with regard to the use of this part of the plant against hypertension, although further study is needed to identify the active ingredients responsible for this activity.

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Conflicts of interest

The authors declare that they have no conflicts of interest.

REFERENCES

- Akerele O (1993). Nature's medicinal bounty: don't throw it away. World Health Forum 14(4):390-5.
- Archer SL, Huang JM, Hampl V, Nelson DP, Shultz PJ, Weir EK (1994). Nitric oxide and cGMP cause vasorelaxation by activation of a charybdotoxin-sensitive K channel by cGMP-dependent protein kinase. Proc. Natl. Acad. Sci. USA 91(16):7583-7.
- Car MR, Pierin AMG, Aquino VL (1991). The influence of the education process on the control of arterial hypertension. Rev. Esc. Enferm. USP 25(3):259-69.
- Cardoso CAL, Coelho RG, Honda NK, Pott A, Pavan FR, Leite CQF (2013). Phenolic compounds and antioxidant, antimicrobial and antimycobacterial activities of *Serjania erecta* Radlk. (Sapindaceae). Braz. J. Pharm. Sci. 49:775-782.
- Carvalho F, Telarolli R Jr, Machado JCMS (1998). An anthropological investigation of old age and concepts concerning arterial hypertension. Cad. Saude Publica 14(3):617-21.
- Castelo AP, Arruda BN, Coelho RG, Honda NK, Ferrazoli C, Pott A, Hiruma-Lima CA (2009). Gastroprotective effect of *Serjania erecta* Radlk (Sapindaceae): involvement of sensory neurons, endogenous nonprotein sulfhydryls, and nitric oxide. J. Med. Food 12:1411-1415.
- Chan EC, Pannangpetch P, Woodman OL (2000). Relaxation to flavones and flavonols in rat isolated thoracic aorta: mechanism of action and structure–activity relationships. J. Cardiovasc. Pharmacol. 35:326-333.
- Cordellini S, Carvalho MH, Scivoletto R, Fortes ZB, Nigro D (1990). Indirect evidence for an endothelium-derived contracting factor release in aorta of deoxycorticosterone acetate-salt hypertensive

rats. J. Hypertens. 8:53-60.

- Cordellini S, Novo R, Lanza Júnior U (2006). Exposure to stress differential vascular adaptive response in spontaneously hypertensive and Wistar rats: Role of nitric oxide, and prehypertensive and hypertensive states. Life Sci. 79:646-653.
- Farmacopéia dos Estados Unidos do Brasil (1959). Código Farmacêutico Brasileiro, 2ª Ed.
- Garcez FB, Carvalho FU, Soares AP, Goes TC, dos Santos MR, Teixeira-Silva F (2014). The influence of trait anxiety on the elevation of arterial pressure induced by L-NAME in rats. Neurosci. Lett. 583:11-5.
- García-Redondo AB, Briones AM, Beltrán AE, Alonso MJ, Simonsen U, Salaices M (2009). Hypertension increases contractile responses to hydrogen peroxide in resistance arteries through increased thromboxane A2, Ca2+, and superoxide anion levels. J. Pharmacol. Exp. Ther. 328(1):19-27.
- Gasparotto Junior A, Gasparotto FM, Lourenço EL, Crestani S, Stefanello ME, Salvador MJ, da Silva-Santos JE, Marques MC, Kassuya CA (2011). Antihypertensive effects of isoquercitrin and extracts from *Tropaeolum majus* L.: evidence for the inhibition of angiotensin converting enzyme. J. Ethnopharmacol. 134(2):363-372.
- Gao YJ, Lee RM (2001). Hydrogen peroxide induces a greater contraction in mesenteric arteries of spontaneously hypertensive rats through thromboxane A (2) production. Br. J. Pharmacol. 134(8):1639-46.
- Ginty AT, Carroll D, Roseboom TJ, Phillips AC, de Rooij SR (2013). Depression and anxiety are associated with a diagnosis of hypertension 5 years later in a cohort of late middle-aged men and women. J. Hum. Hypertens. 27(3):187-90.
- Gomig F, Pietrovski EF, Guedes A, Dalmarco EM, Calderari MT, Guimarães CL, Pinheiro RM, Cabrini DA, Otuki MF (2008). Topical anti-inflammatory activity of *Serjania erecta* Radlk (Sapindaceae) extracts. J. Ethnopharmacol. 118(2):220-4.
- Guarim Neto G (1996). Plantas medicinais do Estado de Mato Grosso. Associação Brasileira de Educação Agrícola Superior, Brasília DF.
- Guarim Neto G, Santana SR, Da Silva JVB (2000). Notas etnobotânicas de espécies de Sapindaceae Jussieu. Acta Bot. Bras. 14:327-334.
- Higashi Y, Sasaki S, Nakagawa K, Matsuura H, Oshima T, Chayama K (2002). Endothelial function and oxidative stress in renovascular hypertension. N. Engl. J. Med. 346:1954-1962.
- Home R, Weinman J (1999). Patients' beliefs about pre- scribed medicines and their role in adherence to treat- ment in chronic illness. J. Psychosom. Res. 47(6):555-567.
- Joannides R, Richard V, Haefeli WE, Benoist A, Linder L, Lüscher TF, Thuillez C (1997). Role of nitric oxide in the regulation of the mechanical properties of peripheral conduit arteries in humans. Hypertension 30(6):1465-1470.
- Johnson AK, Schwob JE (1975). Cephalic angiotensin receptors mediating drinking to systemic angiotensin II. Pharmacol. Biochem. Behav. 3(6):1077-1084.
- Lerman LO, Chade AR, Sica V, Napoli C (2005). Animal models of hypertension: an overview. J. Lab. Clin. Med. 146:160-173.
- Leeya Y, Mulvany MJ, Queiroz EF, Marston A, Hostettmann K, Jansakul C (2010). Hypotensive activity of an n-butanol extract and their purified compounds from leaves of *Phyllanthus acidus* (L.) Skeels in rats. Eur. J. Pharmacol. 649(1-3):301-13.
- Li J, Bukoski RD (1993). Endothelium-dependent relaxation of hypertensive resistance arteries is not impaired under all conditions. Circ. Res. 72(2):290-296.
- Li Q, Youn JY, Cai H (2015). Mechanisms and consequences of endothelial nitric oxide synthase dysfunction in hypertension. J. Hypertens. 33(6):1128-36.
- Magadza C, Radloff SE, Srinivas SC (2009). The effect of an educational intervention on patients' knowledge about hypertension, beliefs about medicines, and adherence. Res. Soc. Adm. Pharm. 5(4):363-75.
- Mattos FJA (1997). Introdução a Fitoquímica Experimental. 2. ed. Fortaleza: UFC: Edições UFC pp.18-40.
- Mayworm MA, Salatino A (1996). Fatty acid composition of 'Cerrado' seed oils. J. Sci. Food Agric. 72(2):226-230.

- Naseem KM (2005). The role of nitric oxide in cardiovascular diseases. Mol. Aspects Med. 26(1-2):33-65.
- Nobre F (2010). VI Diretrizes Brasiléiras de Hipertensão. Arq. Bras. Cardiol. 95(1):1-51.
- Panza JA, García CE, Kilcoyne CM, Quyyumi AA, Cannon RO 3rd (1995). Impaired endothelium-dependent vasodilation in patients with essential hypertension. Evidence that nitric oxide abnormality is not localized to a single signal transduction pathways. Circulation 91(6):1732-8.
- Pellow S, Chopin P, File SE, Briley M (1985). Validation of open:closed arm entries in an elevated plus-maze as a measure of anxiety in the rat. J. Neurosci. Methods 14(3):149-67.
- Péres DŚ, Magna JM, Viana LA (2003). Arterial hypertension patients: attitudes, beliefs, perceptions, thoughts and practices. Rev. Saúde Pública 37:635-642.
- Pott A, Pott VJ (1994). Plantas do Pantanal, Empresa Brasileira de Pesquisa Agropecuária (EMBRAPA- CPAP), Brasília, DF p 263.
- Pott A, Pott VJ, Sobrinho AB (2004). Plantas úteis à sobrevivência no Pantanal. Empresa Brasileira de Pesquisa Agropecuária (EMBRAPA), Corumbá, Brazil p 5.
- Proença C, Oliveria RS, Silva AP (2000). Flores e frutos do cerrado. Editora Universidade de Brasília.
- Puzserova A, Ilovska V, Balis P, Slezak P, Bernatova I (2014). Agerelated alterations in endothelial function of femoral artery in young SHR and WKY rats. Biomed. Res. Int. 2014:658479.
- Ross S, Walker A, MacLeod MJ (2004). Patient compliance in hypertension: role of illness perceptions and treatment beliefs. J. Hum. Hypertens. 18(9):607-13.

- Ripley TL, Brenner M, Finks S, Hough A, McConnell KJ, Parker M, Dobesh PP (2015). Key Articles and Guidelines in the Management of Hypertension: 2015 Update. J. Pharm. Pract. 28(2):146-61.
- Sagvolden T (2000). Behavioural validation of the spontaneously hypertensive rat (SHR) as an animal modelof attentiondeficit/hyperactivity disorder (AD/HD). Neurosci. Biobehav. Rev. 24:31-39.
- Sagvolden T, Russell VA, Aase H, Johansen EB, Farshbaf M (2005). Rodent models of attention- deficit/hyperactivity disorder. Biol. Psychiatr. 57:1239-1247.
- Schmitt CA, Dirsch VM (2009). Modulation of endothelial nitric oxide by plant derived products. Nitric Oxide 21:77-91.
- Wilson J, Axelsen K, Tang S (2005). Medicaid prescription drug access restrictions: exploring the effect on patient persistence with hypertension medications. Am. J. Manag. Care 11 Spec No:SP27-34.