

Full Length Research Paper

Antioxidant and antihyperlipidemic activities of unique nicotinyl polysaccharide from the green seaweed *Ulva pertusa*

Yuzhou Wan¹, Xintao Chang², Lin Liu¹, Shaopeng Wang¹, Huimin Qi^{1*} and Fang Yan¹

¹College of Pharmacy, Weifang Medical University, Weifang, Baotong Road 7166, 261053, China.

²People's Hospital of Zhangqiu District, Jinan, Huiquan Road, No. 1920, 250200, China.

Received 16 February, 2022; Accepted 5 April, 2022

Nicotinyl derivative of polysaccharide (NU) from the green seaweed *Ulva pertusa* (Chlorophyta) has unique structure and strong antioxidant activity *in vitro*. In the present study, *in vivo* antioxidant and antihyperlipidemic activities were tested in the livers of hyperlipidemic mice. Activity levels of superoxide dismutase, glutathione peroxidase (GSH-Px), catalase, and malondialdehyde were observed. At the 500 mg/kg dose, NU showed strongest antioxidant activity compared with the hyperlipidemic mice. This dose also increased GSH-Px compared with ulvan. NU at a dose of 125 mg/kg showed the strongest antihyperlipidemic activity, significantly decreasing total cholesterol (TC), triglyceride (TG), and low-density lipoprotein levels (LDL-C), as well as elevating high density lipoprotein. The antioxidant and antihyperlipidemic mechanisms of NU may be related to the nicotinyl group in its chemical structure. NU may be effective in protecting liver tissue from the damage of a cholesterol-rich diet in mice and may be of use as a novel antihyperlipidemic agent.

Key words: *Ulva pertusa*, polysaccharide, nicotinyl ulvan, antioxidant activity, hyperlipidemic activity.

INTRODUCTION

Dyslipidemia is characterized with high levels of total cholesterol (TC), triglycerides (TG), low-density lipoprotein cholesterol (LDL-C), and a low level of high-density lipoprotein cholesterol (HDL-C) in clinical investigations. Disordered lipid metabolism is a primary cause of atherosclerosis and cardiovascular disease (Arnett et al., 2019). In China, related report indicated that the prevalence and mortality from cardiovascular disease (CVD) were significantly increased with a rise in living

standard and worsening lifestyle habits in the past 20 years (Hu et al., 2017). Reactive oxygen species (ROS) are free radicals with the center of oxygen. The massive production of ROS induces the hyperlipidemia (Miah et al., 2021; Steven et al., 2017; Zhang et al., 2016). Their extreme unstableness and ability to react very quickly with other substances and groups in the body can lead to injury of cells or tissues (Zimmerman and Case, 2019).

ROS can be scavenged by many enzymes, such as

*Corresponding author. E-mail: wfqihuimin@126.com. Tel: +86 536- 8462493.

superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GSH-Px) and peroxiredoxin (Prx) (Garcia and Blesso, 2021).

A polysaccharide is a large molecule comprised of a chain or ring of smaller monosaccharides, or simple sugars, which are linked by glycosidic bonds. The biological activities of antioxidant, anti-inflammatory and so on are reasons polysaccharides are being studied intensively (Liu et al., 2015). Much attention has been focused on polysaccharides isolated from algae (Cao et al., 2021; Li et al., 2020; Liu et al., 2019). *Ulva pertusa*, (sea lettuce), is an edible green alga. Off the eastern coast of China, it is distributed in the Bohai Sea and the Yellow Sea. *U. pertusa* is a low-calorie food containing abundant vitamins, dietary fiber, and trace elements (Tanna and Mishra, 2019). Polysaccharides extracted from *U. pertusa* show a wide range of pharmacologic activities such as antioxidant, antiviral, anti-hyperlipidemia, and anticancer properties (Li et al., 2018; Liu et al., 2019; Sun et al., 2018). Ulvan is a water-soluble polysaccharide extracted from *U. pertusa*, and is in a group of sulphated heteropolysaccharides mainly composed of rhamnose, xylose, glucose, glucuronic acid, iduronic acid, and sulfate (Gao et al., 2020; Li et al., 2020, 2018).

Synthetic antihyperlipidemic agents may be effective for treating dyslipidemia; however, their adverse effects are well known. Natural sources of antioxidants and antihyperlipidemics may potentially mitigate these side effects. It has been found that natural antioxidants possess potential antihyperlipidemic activities (Mu et al., 2021; Adigun et al., 2016). In previous research, our team has prepared the nicotinyl ulvan (NU) and demonstrated its antioxidant activities *in vitro* (Chang et al., 2017). Thus, in the current study we firstly explored the antihyperlipidemic and antioxidant activities of NU in the mice model of hyperlipidemia.

MATERIALS AND METHODS

Ethical approval

This study was approved by the Ethics Committee for Animal Studies, Weifang Medical University and carried out in accordance with the National Academy of Sciences Guide for the Care and Use of Laboratory Animals (2011).

Plant materials

U. pertusa was collected on the coast of Taiping Jiao, Qingdao, China, in October, 2015. The algae specie was identified at the Medical University, China (Figure 1) and thereafter washed, air-dried, and stored at 25°C in plastic bags before use.

Chemical reagents

Malondialdehyde (MDA), SOD, CAT, and GSH-Px assay kits were obtained from Nanjing Jiancheng Bioengineering (Nanjing, China).

Serum total cholesterol (TC), triglycerides (TG), low-density lipoprotein cholesterol (LDL-C) and high-density lipoprotein cholesterol (HDL-C) assay kits were purchased from Shanghai Rongsheng Biotech (Shanghai, China). All other chemical reagents were of analytical grade.

Preparation of nicotinyl ulvan

Ulvan isolated from *U. pertusa* (U) and nicotinyl ulvan (NU) were prepared according to the method described by Chang et al. (2017). NU was prepared using the solvent dimethylformamide (DMF). The mass ratio of ulvan to nicotinoyl was 1:2, with the reaction carried out at 125°C for 4 h. The nitrogen contents of ulvan and NU were 0.60 and 6.62%, respectively. Elemental analyzer (vario EL cube; Elementar Analysensysteme GmbH, Ronkonkoma, NY, USA) was used to measure the nitrogen content of NU.

Experimental design

Seventy (70) male Kunming mice (18 ± 2 g) were purchased from the Animal Laboratory Center, Shandong University, China (license no. SCXK (Lu) 20090001). The mice were housed under controlled conditions of 12 h light:dark cycle, room temperature, and 60% humidity. After three days of acclimatization, mice were randomly divided into 7 groups. Both the normal (NOR) and hyperlipidemia control (MOD) groups received the same daily intragastrically-administered volume (0.7 ml) of deionized water. U group was treated daily with ulvan (250 mg/kg). LNU, MNU, HNU groups were treated with low-dose (125 mg/kg), medium-dose (250 mg/kg), and high-dose (500 mg/kg) of NU in daily, respectively. Positive group received nicotinic acid daily (500 mg/kg). Treatment was for 32 days.

NOR group remained on the standard laboratory diet throughout the study. Animals in all of other groups received a high-cholesterol diet consisting of sodium cholic acid, cholesterol, lard and commercial chow, with the proportion of them being 0.3, 2.0, 8.0 and 89.7%, respectively (Qi et al., 2005). Food intake was recorded daily. During the study period, all animals were weighed every five days during the first 25 days and then weighed for the last time from days 26 to 32 to determine the final weight. After 32 days, the animals were fasted for 12 h. They were then re-weighed, animals were anesthetized with 4% chloral hydrate and blood samples (about 1 ml) were collected retro-orbitally. According to manufacturers' instructions, the levels of TC, TG, HDL-C, and LDL-C in serum were measured. Animals were then euthanized by cervical dislocation; thereafter the livers were removed and immediately stored at -80°C. MDA, SOD, CAT and GSH-Px were detected using the commercial kit following the protocols provided by manufacturer.

Statistical analysis

Data were expressed as mean ± SD and analyzed by one-way ANOVA. Differences were considered to be significant when $P < 0.05$, and very significant when $P < 0.01$.

RESULTS

Food intake and body weight of mice

Among the seven groups, there were no significant changes in food intake, though the normal control group



Figure 1. Appearance of *Ulva pertusa* collected from Qingdao, Shandong province, China.

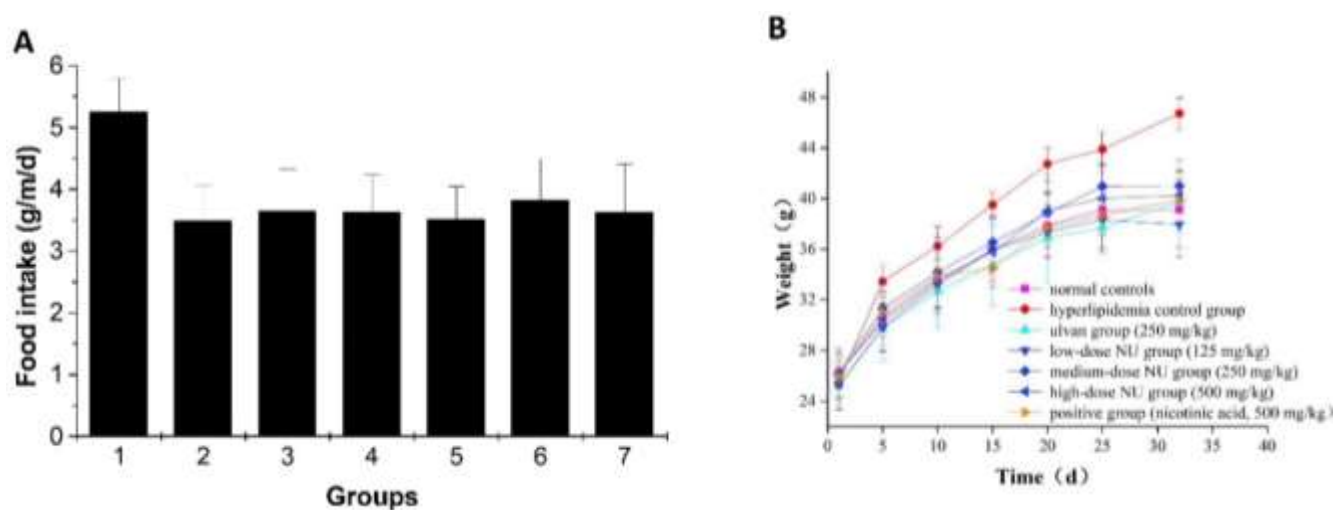


Figure 2. Effect of nicotinyl ulvan on food intake and body weight of mice (A. food intake, 1: NOR group; 2: MOD group; 3: U group (250 mg/kg); 4: LNU group (125 mg/kg); 5: MNU group (250 mg/kg); 6: HNU (500 mg/kg); 7: Positive group (nicotinic acid, 500 mg/kg); B. body weight).

did exhibit an elevated but insignificant level of food intake (Figure 2A). Hyperlipidemia is typically accompanied by an increase in body weight (BW). Throughout this study, all mice remained in good health, without mortalities. In the first 5 days, BW gain was observed in all of the groups without significant difference. From the

6th day, BW gain in the hyperlipidemia group increased rapidly, and was significantly higher compared with the normal control group ($P < 0.05$) (Figure 2B). There were no significant differences in BW gain of the other six groups. Of note, BW decreased during the last week of the study, although not significantly between low-dose

Table 1. Activities of antioxidant enzymes in high-cholesterol diet mice liver.

Group	Dose (mg/kg)	SOD (U/mg prot)	GSH-Px (U/mg prot)	CAT (U/mg prot)	MDA (nmol/mg prot)
1	-	99.47±13.80	452.39±68.00	331.49±53.30	0.86±0.27
2	-	75.32±8.84*	289.85±85.91**	188.63±47.40*	1.08±0.17*
3	250	94.99±14.41 [#]	336.76±60.34	313.54±59.00 [#]	0.84±0.24 [#]
4	125	94.13±14.04	436.28±86.56 [#]	334.91±61.80 [#]	0.87±0.17
5	250	91.58±21.35	384.11±69.21	274.56±46.10	0.87±0.16
6	500	96.37±25.32 [#]	450.59±72.34 ^{##} Δ	326.84±51.30 [#]	0.93±0.19
7	500	98.10±19.96 [#]	355.75±73.51	305.17±34.70 [#]	0.84±0.16 [#]

1: NOR group; 2: MOD group; 3: U group (250 mg/kg); 4: LNU group (125 mg/kg); 5: MNU group (250 mg/kg); 6: HNU (500 mg/kg); 7: Positive group (nicotinic acid, 500 mg/kg), *P < 0.05, **P < 0.01 vs. NOR group; [#]P < 0.05, ^{##}P < 0.01 vs. MOD group. ΔP < 0.05 vs. U group.

Table 2. Changes of serum lipid profiles in high-cholesterol diet mice.

Group	Dose (mg/kg)	TC (mmol/L)	TG (mmol/L)	LDL-C (mmol/L)	HDL-C (mmol/L)	LDL/HDL
1	-	3.06±0.56	0.98±0.04	0.26±0.08	1.00±0.30	0.26
2	-	4.70±0.84**	1.52±0.46*	0.46±0.08**	0.57±0.11*	0.81
3	250	3.56±0.45 ^{##}	1.05±0.38 [#]	0.34±0.05 [#]	0.79±0.21 [#]	0.43
4	125	3.85±0.61 [#]	0.79±0.25 ^{##} , Δ	0.30±0.10 [#]	0.84±0.32 [#]	0.36
5	250	4.23±0.27	1.04±0.29 [#]	0.41±0.07	0.88±0.02 [#]	0.47
6	500	4.37±0.51Δ	0.99±0.25 ^{##}	0.38±0.08	1.06±0.18 [#]	0.36
7	500	3.69±0.29 ^{##}	1.04±0.27 [#]	0.33±0.12 [#]	1.02±0.28 [#]	0.32

1: NOR group; 2: MOD group; 3: U group (250 mg/kg); 4: LNU group (125 mg/kg); 5: MNU group (250 mg/kg); 6: HNU (500 mg/kg); 7: Positive group (nicotinic acid, 500 mg/kg), *P < 0.05, **P < 0.01 vs. NOR group; [#]P < 0.05, ^{##}P < 0.01 vs. MOD group. ΔP < 0.05 vs. U group.

NU group (125 mg/kg) and the normal controls.

Antioxidant activities of ulvan, NU, and nicotinic acid in high-cholesterol diet mice liver

Compared with the MOD group, the HNU group exhibited more optimal effects on antioxidant enzyme (SOD, CAT, and GSH-Px). After treating with high dose of NU, the activities of these enzymes increased by 27.9, 55.5, and 73.3% (P < 0.05), respectively (Table 1). Moreover, compared with the U, HNU significantly increased GSH-Px (33.8%; P < 0.05). However, all U and three dose NU-treated groups exhibited no significant effects on MDA. Furthermore, MDA levels were not affected by U nor by the three dose levels of NU.

Effects of ulvan, NU, and nicotinic acid on serum lipid indicators in high-cholesterol diet mice

In the serum, MOD group showed significant levels of TC, TG, and LDL-C is about 1.5, 1.6, and 1.8 times higher

than that of the NOR group (P < 0.05), respectively (Table 2). Compared with the NOR group, HDL-C levels in MOD group were obviously reduced by 43.0% (P < 0.05). All above data indicated that the hyperlipidemic model was successfully established. After 32 days of treatment, levels of TG were decreased significantly in the NU groups than in the MOD group, with the LNU group exhibiting the most robust response. TG reduced by 48.0% (P < 0.01) than the MOD group and reduced by 24.8% (P < 0.05) compared with the U group. However, these declines were not in a dose-independent manner. In the LNU group, the levels of TC, LDL-C, and TG were significantly decreased by 18.1, 34.8, and 48.0% than MOD group (P < 0.05), respectively, while the level of HDL-C in serum improved by 47.4% (P < 0.05). On the other hand, compared with the MOD group, U and three NU dose groups all had significant effects on HDL-C levels. The results indicated that LNU group may be beneficial for the treatment of hyperlipidemia.

DISCUSSION

The antioxidant enzymes (SOD, CAT, and GSH-Px)

prevent against the production of ROS under oxidative stress (Irazabal and Torres, 2020). It has been demonstrated in hyperlipidemic rat model that the vigours of SOD, CAT, and GSH-Px were decreased (Li et al., 2020). The activities of these antioxidant enzymes were also decreased in the liver of hyperlipidemic mice (Ru et al., 2019; Wang et al., 2019). In the present study, ulvan and NU showed that strong activity improved the ability of these three enzymes in the livers of hyperlipidemia mice, especially under those treated with high dose of NU, which exhibited the strongest activity. SOD as crucial endocellular compound plays a role in protecting against oxidative stress caused by superoxide anion. For NU groups, only HNU (500 mg/kg) significantly improved the activity of SOD. CAT, enriched in hepatocytes, is crucial in catalyzing dismutation of toxic H_2O_2 (Hong and Park, 2021). Except for the MNU group (250 mg/kg), the activity of CAT showed significant difference between other drug administration groups and hyperlipidemic mice. MDA as the marker of oxidative stress mainly produced by endogenous lipid peroxidation, a process induced by free radicals (Mas-Bargues et al., 2021). However, in this study, as compared with the positive and ulvan groups, all three doses of NU showed lesser activity on MDA.

The beneficial effects of a serum high level of HDL-C are well known. In the liver, HDL-C plays a role in cholesterol metabolism by reverse cholesterol transport (Zhao et al., 2012). In our study, mice that received U, the three doses of NU, or the positive controls all showed significantly increased HDL-C levels, with high-dose NU exhibiting similar activity to the positive controls. The increased LDL/HDL ratio and its existence in a positively correlated manner showed that the ratio of LDL-C to HDL-C could be a predictor of atherosclerosis and cardiovascular conditions (Carr et al., 2019). Our data indicated that administration of low and high-dose NU significantly suppressed LDL/HDL ratio by 55.6% compared with the hyperlipidemic group.

Nicotinic acid (vitamin B₃/niacin) has antihyperlipidemic effects, but patients often have side effects such as facial flushing, burning sensation, and upper gastrointestinal discomfort (Meyer-Ficca and Kirkland, 2016). The results of this study revealed that LNU had the strongest antihyperlipidemic activity and HNU had the most robust antioxidant activity in the liver of hyperlipidemia mice. Many studies have demonstrated that acetylated derivative of polysaccharide, high-sulfated polysaccharide, and phosphorylated polysaccharide exhibited stronger biological activities than polysaccharide alone (Chen and Huang, 2019; Li et al., 2020; Lopes et al., 2017; Ren et al., 2020; Jiang et al., 2020). These studies also revealed that some activities can be enhanced through the structural modification.

This current study is an initial investigation of the *in vivo* antioxidative and antihyperlipidemic activity of NU from *U. pertusa*. One of the mechanisms of ulvan inhibiting

oxidative stress is that it can directly capture ROS or provide H to act on $\cdot OH$ and produce H_2O (Huang, 2017). Nicotinyl group was linked to ulvan, on the one hand, that increases the spatial volume of ulvan, thereby increasing the ability of ulvan to capture ROS, and on the other hand, increases the hydrogen-donating ability of ulvan, so that ulvan can react with more $\cdot OH$ to generate H_2O (Huang, 2017; Qiu et al., 2022; Zhou and Huang, 2021). NU reduces the occurrence of oxygen radical chain reaction by reacting with ROS, thereby reducing the consumption of metal ions (Fe^{2+} , Cu^{2+}) which are an important part of SOD (Wang et al., 2018). It may be that the mechanism of NU improves the activities of antioxidative enzymes. The mechanism of NU's modulation of lipid metabolism is not fully understood. In our previous study, we successfully prepared NU from *U. pertusa* and identified its unique structure (Chang et al., 2017). Based on studies by other investigators, nicotinyl enhanced the ion exchange capacity of ulvan, make it to better act with bile acid in gut, and furthermore decreases the serum TC (Evans, 2020). At the genetic molecular level, ulvan enhances the expression of peroxisome proliferator-activated receptor gamma (PPAR γ) in hyperlipidemic rats (Qi and Sheng, 2015). Therefore, our future investigations may focus on the effects of NU on PPAR γ gene expression. Of further consideration is that in the atherosclerotic process, oxidative stress contributes to endothelial damage (Yang et al., 2010). Moreover, lipid peroxidation can be induced by massive free radicals, leading to dysfunction of the lipid metabolism pathway in the liver and aggravation of hyperlipidemia. Therefore, research on the antioxidant properties of polysaccharides derived from natural sources such as the seaweed *Porphyra haitanensis* and sea cucumber *Apostichopus japonicus* is garnering importance (Wang et al., 2017; Lin et al., 2012). All of the results revealed that among the three different doses of NU, the high dose (500 mg/kg) showed the strongest antioxidant activity while the low dose (125 mg/kg) had the most robust antihyperlipidemic activity. The mechanisms of these effects remain to be investigated.

Conclusion

NU derivative from the green seaweed *U. pertusa* (Chlorophyta) possess antioxidant and antihyperlipidemic activities *in vivo*. NU showed the more powerful activities of antioxidant and antihyperlipidemic than ulvan. The antioxidant and antihyperlipidemic mechanisms of NU may be related to the nicotinyl group in its chemical structure. As a natural antioxidant, NU is potentially responsible for relieving or treating chronic diseases caused by oxidative stress. In addition, NU has hypolipidemic activity, as it is not only able to treat hypolipidemic conditions, but can also be a potential in preventing cardiovascular and cerebrovascular diseases

caused by hyperlipidemia.

CONFLICT OF INTERESTS

The authors have not declared any conflict of interests.

ACKNOWLEDGEMENTS

The authors thank Nissi S. Wang, for developmental editing of the manuscript. This work was financially supported by the National Natural Science Foundation of China (41206129), the Natural Science Foundation of Shandong Province (ZR2012DL13) (ZR2020MH400), the Development of Medical Science and Technology Project of Shandong Province (2016WS0665), and Weifang Medical University Teachers Domestic Scholar Program.

REFERENCES

- Adigun NS, Oladiji AT, Ajiboye TO (2016). Antioxidant and anti-hyperlipidemic activity of hydroethanolic seed extract of *Aframomum melegueta* K. Schum in Triton X-100 induced hyperlipidemic rats. South African Journal of Botany 105:324-332.
- Arnett DK, Blumenthal RS, Albert MA, Buroker AB, Goldberger ZD, Hahn EJ, Himmelfarb CD, Khera A, Lloyd-Jones D, McEvoy JW, Michos ED, Miedema M D, Muñoz D, Smith SC, Jr Virani SS, Williams KA, Sr Yeboah J, Ziaeian B (2019). 2019 ACC/AHA Guideline on the Primary Prevention of Cardiovascular Disease: Executive Summary: A Report of the American College of Cardiology/American Heart Association Task Force on Clinical Practice Guidelines. Circulation, 140(11):e563-e595.
- Cao M, Li Y, Famurewa AC, Olatunji OJ (2021). Antidiabetic and Nephroprotective Effects of Polysaccharide Extract from the Seaweed *Caulerpa racemosa* in High Fructose-Streptozotocin Induced Diabetic Nephropathy. Diabetes, Metabolic Syndrome and Obesity: Targets and Therapy 14:2121-2131.
- Carr SS, Hooper AJ, Sullivan DR, Burnett JR (2019). Non-HDL-cholesterol and apolipoprotein B compared with LDL-cholesterol in atherosclerotic cardiovascular disease risk assessment. Pathology 51(2):148-154.
- Chang XT, Li WD, Jiang NF, Liu K, Xue M, Qi HM, Liu ZJ (2017). The preparation of ulvan derivative NU and the study on antioxidant activity in vitro. Chinese Journal of Marine Drugs 36:27-33.
- Chen L, Huang G (2019). Antioxidant activities of phosphorylated pumpkin polysaccharide. International Journal of Biological Macromolecules 125:256-261.
- Evans CEL (2020). Dietary fibre and cardiovascular health: a review of current evidence and policy. Proceedings of the Nutrition Society 79(1):61-67.
- Gao X, Qu H, Shan S, Song C, Baranenko D, Li Y, Lu W (2020). A novel polysaccharide isolated from *Ulva Pertusa*: Structure and physicochemical property. Carbohydrate Polymers 233:115849.
- Garcia C, Blesso CN (2021). Antioxidant properties of anthocyanins and their mechanism of action in atherosclerosis. Free Radical Biology and Medicine 172:152-166.
- Hong YA, Park CW (2021). Catalytic Antioxidants in the Kidney. Antioxidants (Basel) 10(1):130.
- Hu S, Gao RL, Liu LS, Zhu ML, Wang W, Wang YJ, Wu ZS, Li HJ, Gu DF, Yang YJ, Zheng Z, Jiang LX, Chen WW, Sui H, Ma LY (2017). Report on cardiovascular diseases in China 2016. Beijing, China: Encyclopedia of China Publishing House.
- Huang G, Mei X, Hu J (2017). The Antioxidant Activities of Natural Polysaccharides. Curr Drug Targets 18(11):1296-1300.
- Irazabal MV, Torres VE (2020). Reactive Oxygen Species and Redox Signaling in Chronic Kidney Disease. Cells 9(6):1342.
- Jiang NF, Li BX, Wang XQ, Xu XN, Liu XL, Li WD, Chang XT, Li H, Qi HM (2020). The antioxidant and antihyperlipidemic activities of phosphorylated polysaccharide from *Ulva pertusa*. International Journal of Biological Macromolecules 145:1059-1065.
- Li BX, Xu HT, Wang XQ, Wan YZ, Jiang NF, Qi HM, Liu XX (2020). Antioxidant and antihyperlipidemic activities of high sulfate content purified polysaccharide from *Ulva pertusa*. International Journal of Biological Macromolecules 146:756-762.
- Li WD, Wang K, Jiang NF, Liu XL, Wan MH, Chang XT, Liu SM (2018). Antioxidant and antihyperlipidemic activities of purified polysaccharides from *Ulva pertusa*. Journal of Applied Phycology 30(4):2619-2627.
- Lin X, Sun ZL, Zhang MS, Meng XM, Xia XK, Yuan WP, Xue F, Liu CH (2012). Antioxidant and antihyperlipidemic activities of polysaccharides from sea cucumber *Apostichopus japonicus*. Carbohydrate Polymers 90(4):1664-1670.
- Liu J, Willfor S, Xu CL (2015). A review of bioactive plant polysaccharides: Biological activities, functionalization, and biomedical applications. Bioactive Carbohydrates and Dietary Fibre 5(1):31-61.
- Liu Z, Gao T, Yang Y, Meng F, Zhan F, Jiang Q, Sun X (2019). Anti-Cancer Activity of Porphyran and Carrageenan from Red Seaweeds. Molecules 24(23):4286.
- Lopes N, Ray S, Espada SF, Bomfim, WA, Ray B, Faccin-Galhardi LC, Nozawa C (2017). Green seaweed *Enteromorpha compressa* (Chlorophyta, Ulvaceae) derived sulphated polysaccharides inhibit herpes simplex virus. International Journal of Biological Macromolecules 102:605-612.
- Mas-Bargues C, Escrivá C, Dromant M, Borrás C, Viña J (2021). Lipid peroxidation as measured by chromatographic determination of malondialdehyde. Human plasma reference values in health and disease. Archives of Biochemistry and Biophysics 709:108941.
- Meyer-Ficca M, Kirkland JB (2016). Niacin. Advances in Nutrition 7(3):556-558.
- Miah P, Mohona SBS, Rahman MM, Subhan N, Khan F, Hossain H, Alam MA (2021). Supplementation of cumin seed powder prevents oxidative stress, hyperlipidemia and non-alcoholic fatty liver in high fat diet fed rats. Biomedicine and Pharmacotherapy 141:111908.
- Mu S, Yang W, Huang G (2021). Antioxidant activities and mechanisms of polysaccharides. Chemical Biology and Drug Design 97(3):628-632.
- Qi H, Zhang Q, Zhao T, Chen R, Zhang H, Niu X, Li Z (2005). Antioxidant activity of different sulfate content derivatives of polysaccharide extracted from *Ulva pertusa* (Chlorophyta) in vitro. International Journal of Biological Macromolecules 37(4):195-199.
- Qi HM, Sheng JW (2015). The Antihyperlipidemic Mechanism of High Sulfate Content Ulvan in Rats. Marine Drugs 13:3407-3421.
- Qiu SM, Aweya JJ, Liu X, Liu Y, Tang S, Zhang W, Cheong KL (2022). Bioactive polysaccharides from red seaweed as potent food supplements: a systematic review of their extraction, purification, and biological activities. Carbohydrate Polymers 275:118696.
- Ren YY, Sun PP, Ji YP, Wang XT, Dai SH, Zhu ZY (2020). Carboxymethylation and acetylation of the polysaccharide from *Cordyceps militaris* and their alpha-glucosidase inhibitory activities. Natural Product Research 34(3):369-377.
- Ru Y, Chen X, Wang J, Guo L, Lin Z, Peng X, Qiu B (2019). Polysaccharides from *Tetragonia hemsleyana* Diels et Gilg: Extraction optimization, structural characterizations, antioxidant and antihyperlipidemic activities in hyperlipidemic mice. International Journal of Biological Macromolecules 125:1033-1041.
- Steven S, Daiber A, Doppeide JF, Munzel T, Espinola-Klein C (2017). Peripheral artery disease, redox signaling, oxidative stress - Basic and clinical aspects. Redox Biology 12:787-797.
- Sun Y, Chen X, Song L, Liu S, Yu H, Wang X, Li P (2018). Antiviral Activity against Avian Leucosis Virus Subgroup J of Degraded Polysaccharides from *Ulva pertusa*. BioMed research international 3:1-11.
- Tanna B, Mishra A (2019). Nutraceutical Potential of Seaweed Polysaccharides: Structure, Bioactivity, Safety, and Toxicity. Comprehensive Reviews in Food Science and Food Safety 18(3):817-831.

- Wang W, Song X, Gao Z, Zhao H, Wang X, Liu M, Jia L (2019). Anti-hyperlipidemic, antioxidant and organic protection effects of acidic-extractable polysaccharides from *Dictyophora indusiata*. *International Journal of Biological Macromolecules* 129:281-292.
- Wang XM, Li WD, Liu CD, Qi HM, Zhang ZS (2017). In vivo antihyperlipidemic and antioxidant activity of porphyran in hyperlipidemic mice. *Carbohydrate Polymers* 174:417-420.
- Wang Y, Branicky R, Noe A, Hekimi S (2018). Superoxide dismutases: Dual roles in controlling ROS damage and regulating ROS signaling. *Journal of Cell Biology* 217(6):1915-1928.
- Yang XL, Yang L, Zheng HY (2010). Hypolipidemic and antioxidant effects of mulberry (*Morus alba* L.) fruit in hyperlipidaemia rats. *Food and Chemical Toxicology* 48(8-9):2374-2379.
- Zhang Y, Hu T, Zhou HL, Zhang Y, Jin G, Yang Y (2016). Antidiabetic effect of polysaccharides from *Pleurotus ostreatus* in streptozotocin-induced diabetic rats. *International Journal of Biological Macromolecules* 83:126-132.
- Zhao LY, Huang W, Yuan QX, Cheng J, Huang ZC, Ouyang LJ, Zeng FH (2012). Hypolipidaemic effects and mechanisms of the main component of *Opuntia dillenii* Haw, Polysaccharides in high-fat emulsion-induced hyperlipidaemic rats. *Food Chemistry* 134:964-971.
- Zhou S, Huang G (2021). Preparation, structure and activity of polysaccharide phosphate esters. *Biomed Pharmacother* 144:112332.
- Zimmerman MC, Case AJ (2019). Redox biology in physiology and disease. *Redox Biology* 27:101267.