

Full Length Research Paper

Gas chromatography-mass spectrometry analysis and phytochemical screening of ethanolic root extract of *Plumbago zeylanica*, Linn.

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Plumbago zeylanica is a medicinal plant widely used in folklore medicine in Africa and Asia for the management of ailments such as parasitic diseases, scabies, ulcers, piles, diarrhoea, skin diseases, leprosy, fever or malaria, rheumatism and intestinal parasites. The present study was carried out to identify the phytochemical components present in the ethanolic root extract of the plant by phytochemical screening methods and GC-MS analysis. The phytochemical screening showed the presence of alkaloids, tannins, steroids, flavonoids, saponins, anthraquinones, cardiac-glycosides, phlobatinnins and carbohydrates. The gas chromatography-mass spectrometry (GC-MS) analysis also identified the presence of phytochemical components like phenolics (Phenol, 2,4-bis(1,1-dimethylethyl) (RT: 6.796), Cyclopentadecane (RT: 7.305), fatty acids/methyl esters, Feropfen (RT: 9.602), 1,1,1,5,7,7,7-Heptamethyl-3,3-bis(trimethylsiloxy)tetrasiloxane (RT: 8.253), Indazol-4-one, 3,6,6-trimethyl-1-phthalazin-1-yl-1,5,6,7-tetrahydro- (RT: 10.162), 2H-Indol-2-one, 1-(2,6-dichlorophenyl)-1,3-dihydro- (RT: 11.191), 2-Methyl-7-phenylindole (RT: 14.540), 1H-Indole-2-carboxylic acid, 6-(4-ethoxyphenyl)-3-methyl-4,5,6,7-tetrahydro-, isopropyl ester (11.528), and Anthranilic acid, 3-chloro-N-(o-chlorophenyl)- (RT: 12.443).

Key words: *Plumbago zeylanica*, gas chromatography, mass spectrometry, phytochemical screening.

INTRODUCTION

The use of plants in ethnomedicine is increasing around the world. The World Health Organization (WHO) has reported that approximately 80% of the world's population currently uses herbal medicines as teas, decocts or extracts with easily accessible liquids such as water, milk or alcohol (Julsing et al., 2007; Kalidass et al., 2010). In Nigeria, the use of herbs in traditional medicine is widely accepted as a significant instrument of health care. Herbal drugs are widely prescribed, even when their

biological components are not known, as a result of their effectiveness, fewer side effects and relatively low cost (Kumar et al., 2009).

Plumbago zeylanica Linn. belongs to the family Plumbaginaceae. *P. zeylanica* is a branched evergreen shrub reaching up to 2 m. It is distributed throughout the tropical and subtropical countries of the world. Four species of the genus *Plumbago* have been reported. They are *Plumbago indica*, *Plumbago rosea*, *Plumbago capensis*, and *P. zeylanica* (Arunachalam et al., 2010). *P. zeylanica* is known by different names in different parts of the world. Such names include *white leadwort* or *ceylon leadwort* (in English), *bleiwurz* or *zahnkraut* (in German), *chitrak* or *chitramol* (in India), *ensain* or *enkin* (in Arabia),

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sanza (in Swahili), and *inabiri* among the Yoruba (in South- West Nigeria). The whole plant, roots, powder of the root, leaves and stem-bark are widely used as medicinal herbs throughout Asia and Africa. In traditional herbal medicine, the root part of *P. zeylanica* is used for treatment of different ailments such as parasitic diseases, scabies and ulcers (Olagunju et al., 2006), piles, diarrhoea, skin diseases and leprosy (Uma et al., 1999), fever or malaria, rheumatism, intestinal parasites, anemia due to 'stagnant blood', internal and external trauma, toxic swelling and furunculosis scabies (Olagunju et al., 2006; Jeyachandran et al., 2009; Jianguo New Medical College, 1979; Dai et al., 2004).

The pharmacological studies of various root extracts of *P. zeylanica* have shown that the plant has antimicrobial (Ahmad et al., 2000), antioxidant and anti-inflammatory (Oyedapo, 1996; Raimi and Oyedapo, 2009), antiplasmodial (Simonsen et al., 2001), antihyperglycemic (Olagunju et al., 1999), antifungal (Mehmood et al., 1999), hypolipidaemic and antiatherosclerotic (Sharma et al., 1991), and central nervous system stimulatory, cytotoxic and anti-insecticidal properties (Arunachalam et al., 2010). Several active compounds previously identified and isolated from the root of *P. zeylanica* include five naphthoquinones (plumbagin, chitranone, maritnone, elliptinone and isoshinanolone), five coumarins (seselin, 5-methoxyseselin, suberosin, xanthyletin and xanthoxyletin), two plumbagic acid glucosides (3'-O- β -glucopyranosyl plumbagic acid and 3'-O- β -glucopyranosyl plumbagic acid methyl ester) (Lin et al., 2003), neoishoshinanolone and 1-epineo-ishoshinanolone (Jetty et al., 2010). In this study, further investigation was carried out to identify other active constituents present in the ethanolic root extract of the plant by phytochemical screening and Gas chromatography-mass spectrometry (GC-MS) analytical methods.

MATERIALS AND METHODS

Plant material

The root parts of *P. zeylanica* was collected in the second week of August, 2008 at Babajakan Village, Osun State, Nigeria through Professor O. O. Oyedapo of the Department of Biochemistry, Obafemi Awolowo University, Ile-Ife, Osun State, Nigeria. Plant identification and authentication had been done earlier by Olagunju et al. (2006) and voucher specimen with code QC 488 had been deposited at the Ife herbarium.

Preparation of extract

The roots were separated from the stalks and thoroughly but gently rinsed with tap water thrice to remove sands. These were then oven dried completely at 27 to 30°C for 1 week. The dried roots were ground to fine powder using a local grinder.

150 g of the powdered root of *P. zeylanica* was exhaustively extracted in 800 ml 99.9% (absolute) ethanol with soxhlet extractor and then concentrated using rotary vacuum evaporator. 10.0 ml extract was measured out from the concentrate for phytochemical screening while the remaining extract was evaporated to complete

dryness at 35°C given a dark-brown color solid residue [yield: 3.14% (^{w/w})]. The dried extract was stored in airtight container and placed in refrigerator.

Phytochemical screening

The extract was subjected to preliminary phytochemical screening to test for the presence or absence of phytochemical constituents using the methods described below by Evans (1996).

Alkaloids

1.0 ml extract was stirred with 5 ml dil. Hydrochloric acid on a steam bath, filtered and 1.0 ml of each Dragendoff's/Mayer's/Wagner's reagents was added to 1.0 ml separate portions of filtrate. A cloudy orange-red/slightly yellow/turbid brown color indicates the presence of alkaloids.

Tannins

1.0 ml extract was stirred with 1.0 ml Ferric chloride. A greenish black precipitate indicates the presence of tannins; 1.0 ml extract was also stirred with 1.0 ml bromine water. A reddish brown turbid color indicates the presence of tannins.

Cardiac-glycoside (Legal's test):

1.0 ml extract was dissolved in 5.0 ml pyridine, 2 drops 2% Sodium Nitroprusside and 2 drops 20% NaOH were added. A deep red color faded to brown indicates presence of cardenolide.

Kedde's test

1.0 ml extract was mixed with 1.0 ml 2% 3,5-Dinitro-benzoic acid in methanol and 1.0 ml aqueous NaOH. A violet color precipitate indicates lactone ring present in cardenolide.

Steroids (Liebermann-Burchard's test)

0.5 g extract dissolved in 2.0 ml acetic anhydride, cooled in ice and 1.0 ml conc. H₂SO₄ was carefully added. A blue-green ring indicates the presence of steroids.

Salkowski's test

0.5 g extract was dissolved in 2.0 ml chloroform and 1.0 ml conc. H₂SO₄ was carefully added. A reddish brown color indicates the presence of steroids.

Flavonoids (Ferric chloride test)

0.2 ml extract was added to 10% FeCl₃ and mixture was shaken. A woolly brownish precipitate indicates the presence of flavonoids.

Lead acetate test

0.2 ml extract was added to 0.2 ml 10% lead acetate and gently shaken. A dirty brownish precipitate indicates the presence of flavonoids.

Table 1. Results of phytochemical screening of ethanolic root extract of *P. zeylanica*

Phytochemicals	Result
Alkaloids	+
Tannins	+
Cardiac-glycosides	+
Steroids	+
Flavonoids	+
Saponins	+
Anthraquinones	+
Phlobatinnins	+
Carbohydrates	+

+ = present.

Sodium hydroxide test

0.2 ml dil. NaOH was added to 0.2 ml extract, gently shaken. A dirty yellowish brown precipitate indicates the presence of flavonoids.

Saponins (Frothing test)

0.2 ml extract was mixed with 5.0 ml distilled water, shaken for 20 min. Persistence of foams indicates presence of saponins.

Anthraquinones (Borntrager's test)

2.0 g extract in 10 ml ethanol, steamed for 5 min and filtered, 2.0 ml filtrate was added to 2.0 ml chloroform, shaken thoroughly, chloroform layer was taken off and 5.0 ml distilled water added which was shaken with 5.0 ml dilute ammonia solution. A red color in ammonia upper phase indicates the presence of anthraquinones.

Phlobatinnins test

1.0 ml extract was boiled in 2.0 ml 1% aqueous HCl. A red precipitate indicates the presence of phlobatinnins.

Carbohydrate tests

1.0 ml extract was added to 2.0 ml Fehling's solution and boiled for 5 min. A red precipitate indicates the presence of reducing sugars; 1.0 ml extract was added to 2.0 ml Barfoed' reagent and boiled for 1 min. A red precipitate indicates the presence of reducing monosaccharides; 1.0 ml extract was added to 1.0 ml Molisch's reagent and 1.0 ml conc. H₂SO₄ was carefully added. A reddish ring indicates the presence of carbohydrates.

Gas chromatography-mass spectrometry analysis

The GC-MS analysis was carried out using a Hewlett Packard Gas Chromatograph (Model 6890 series) equipped with a flame ionization detector and Hewlett Packard 7683 series injector, MS transfer line temperature of 250°C. The GC was equipped with a fused silica capillary column- HP-5MS (30 x 0.25 mm), film thickness 1.0 µm. The oven temperature was held at 50°C for 5 min holding time and raised from 50 to 250°C at a rate of 2°C /min,

employing helium gas (99.999%) as a carrier gas at a constant flow rate of 22 cm/s. 1.0 micron of extract (1 mg dissolved in 1 ml absolute alcohol), at a split ratio of 1:30 was injected. MS analysis was carried out on Agilent Technology Network Mass Spectrometer (Model 5973 series) coupled to Hewlett Packard Gas Chromatograph (Model 6890 series) equipped with NIST08 Library software database. Mass spectra were taken at 70 eV/200°C, scanning rate of 1 scan/s.

Identification of compounds was conducted using the database of NIST08 Library. Mass spectrum of individual unknown compound was compared with the known compounds stored in the software database Library.

RESULTS AND DISCUSSION

Table 1 shows the results of the phytochemical screening of the ethanolic root extract of *P. zeylanica*. This reveals the presence of alkaloids, tannins, steroids, flavonoids, saponins, anthraquinones, phlobatinnins, cardiac-glycosides and carbohydrates. The results of the GC-MS analysis identified the various compounds present in the plant (Figure 1 and Table 2). Figure 1 shows the gas chromatogram of the extract which shows 24 distinct peaks identified by GC-MS while the compounds identified through the NIST08 L. database are listed in Table 2. The major compound present in the ethanolic root extract of *P. zeylanica* as identified by GC-MS was Phenol, 2,4-bis(1,1-dimethylethyl) with RT: 6.796% of Total: 54.62 and quality: 96 (Table 2). Mass spectrum of Phenol, 2,4-bis(1,1-dimethylethyl) was shown in Figure 2. The GC-MS spectrum gives the structure of the compound (Figure 3), molecular formula as C₁₄H₂₂O, molecular weight as 206 and biological activity as antioxidant (Table 3). Other components also identified in the root of *P. zeylanica* are Cyclopentadecane (RT: 7.305), Fenoprofen (RT: 9.602), Indazol-4-one, 3,6,6-trimethyl-1-phthalazin-1-yl-1,5,6,7-tetrahydro- (RT: 10.162), Anthranilic acid, 3-chloro-N-(O-chlorophenyl)-(RT: 12.443), 2-Methyl-7-phenylindole (RT: 14.540) and Docosanoic acid, methyl ester (RT: 13.277) (Figure 1).

It has been reported that many medicinal plants are rich in varieties of secondary metabolites such as alkaloids, flavonoids, tannins and terpenoids (Lewis and Ausubel, 2006; Adekunle and Adekunle, 2009). Reports have it that secondary plant metabolites exert a wide range of biological activities on physiological systems (Olagunju et al., 2006). Kumar et al. (2010) also reported the activities of some phyto-components with compound nature of flavonoids, palmitic acid (hexadecanoic acid, ethyl ester and n-hexadecaonic acid), unsaturated fatty acid and linolenic (docosatetraenoic acid and octadecatrienoic acid) as antimicrobial, anti-inflammatory, antioxidant, hypocholesterolemic, cancer preventive, hepatoprotective, antiarthritic, antihistimic, antieczemic and anticoronary. It is therefore not unlikely that these phytochemicals found in *P. zeylanica* may play major roles in the reported biological activities and pharmacological properties of the plant.

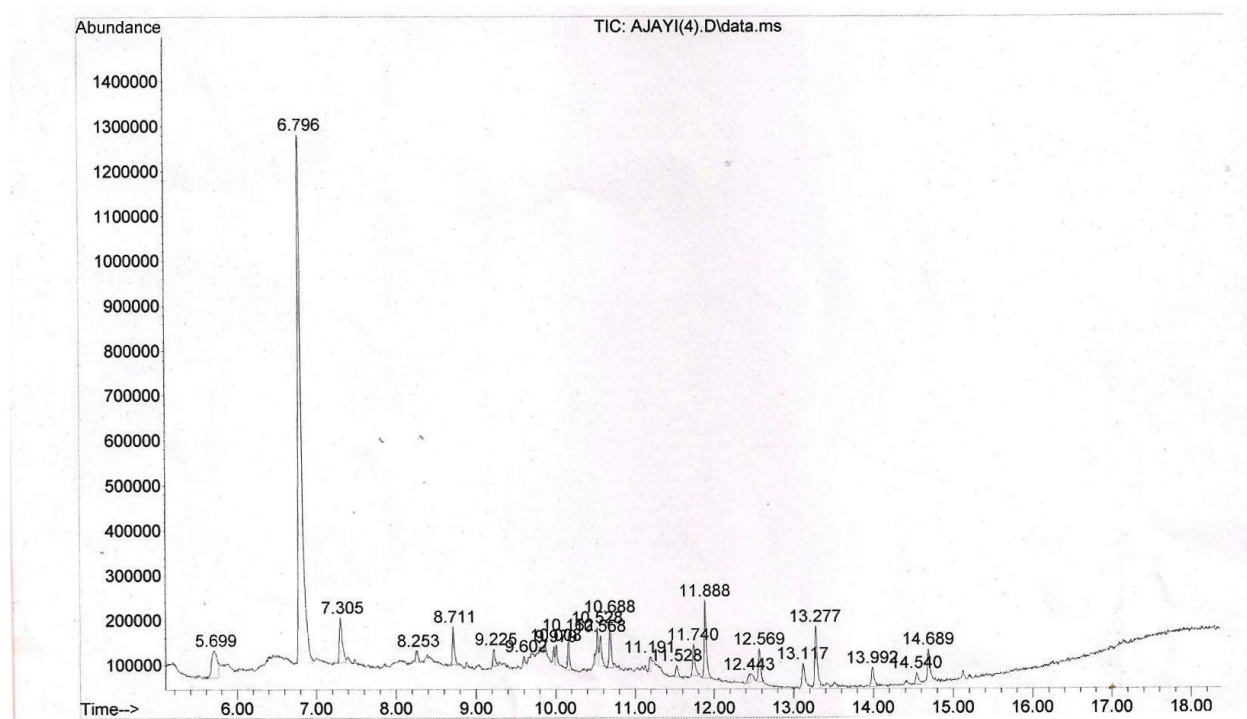


Figure 1. GC-MS chromatogram of ethanolic root extract of *P. zeylanica*. Peak 2 with retention time of 6.796 was identified as phenol,2,4-bis(1,1-dimethylethyl) and as the major phyto-component of the plant while other peaks were of the various phyto-components present.

Table 2. Phytochemical components identified in the ethanolic root extract of *P. zeylanica* by GC-MS showing their RT, % of Total and quality of compounds.

PK	RT	Identified compound	MSD	% of total	Qual
1	5.699	1-Tetradecene	56562 001120-36-1	4.602	95
2	6.796	Phenol,2,4-bis(1,1-dimethylethyl)	63980 000096-76-4	54.623	96
3	7.305	Cyclopentadecane	67239 000295-48-7	3.748	97
4	8.253	1,1,1,5,7,7,7-Heptamethyl-3,3-bis(trimethylsiloxy) tetrasiloxane	206266 038147-00-1	0.750	38
5	8.711	1-Octadecene	99556 000112-88-9	2.106	99
6	9.225	Hexadecanoic acid, methyl ester	113682 000112-39-0	0.942	96
7	9.602	Fenoprofen	91701 031879-05-7	0.633	52
8	9.979	3-Eicosene, (E)-	121344 074685-33-9	0.650	98
9	10.008	Methyl 7-methylhexadecanoate	124582 018720-85-9	1.037	64
10	10.162	Indazol-4-one, 3,6,6-trimethyl-1-phthalazin-1-yl-1,5,6,7-tetrahydro-	141172 1000310-86-5	1.647	64
11	10.528	9-Octadecenoic acid, methyl ester (E)-	133723 001937-62-8	1.909	99
12	10.568	Methyl 11-Octadecenoate	133684 1000336-35-8	1.939	99
13	10.688	Octadecenoic acid, methyl ester	135389 000112-61-8	2.783	99
14	11.191	2H-Indo-2-one, 1-(2,6-dichlorophenyl)-1,3-dihydro-	118782 015362-40-0	0.653	99
15	11.528	1H-Indole-2-carboxylic acid, 6-(4-ethoxyphenyl)-3-methyl-4-oxo-4,5,6,7-tetrahydro-, isopropyl ester	174606 1000316-17-5	0.773	22
16	11.740	cis-11-Eicosenoic acid, methyl ester	154536 1000333-63-8	2.069	99
17	11.888	Eicosanoic acid, methyl ester	156024 001120-28-1	4.689	98
18	12.443	Anthranilic acid, 3-chloro-N-(o-chlorophenyl)-	121641 099513-96-9	0.581	62
19	12.569	Heneicosanoic acid, methyl ester	165667 006064-90-0	2.095	99
20	13.117	13-Docosenoic acid, methyl ester, (Z)-	172949 001120-34-9	2.369	97

Table 2. Contd.

21	13.277	Docosanoic acid, methyl ester	174189 000929-77-1	4.519	99
22	13.992	Tricosanoic acid, methyl ester	181855 002433-97-8	1.612	99
23	14.540	2-Methyl-7-phenylindole	64822 001140-08-5	1.029	86
24	14.689	Tetracosanoic acid, methyl ester	188588 002442-49-1	2.242	99

PK=Peak, RT=Retention time, MSD=Mass spectrum data, Qual=Quality.

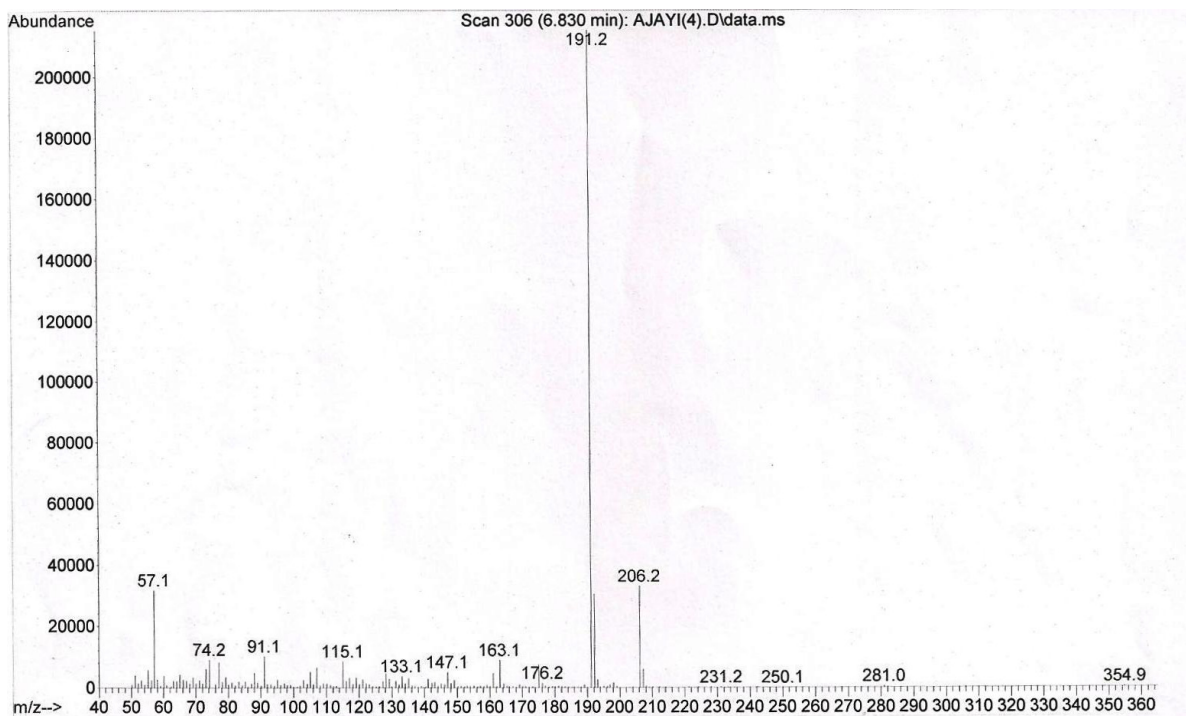


Figure 2. GC-MS mass spectrum of phenol, 2,4-bis(1,1-dimethylethyl) indicating abundant ions m/z 191.2.

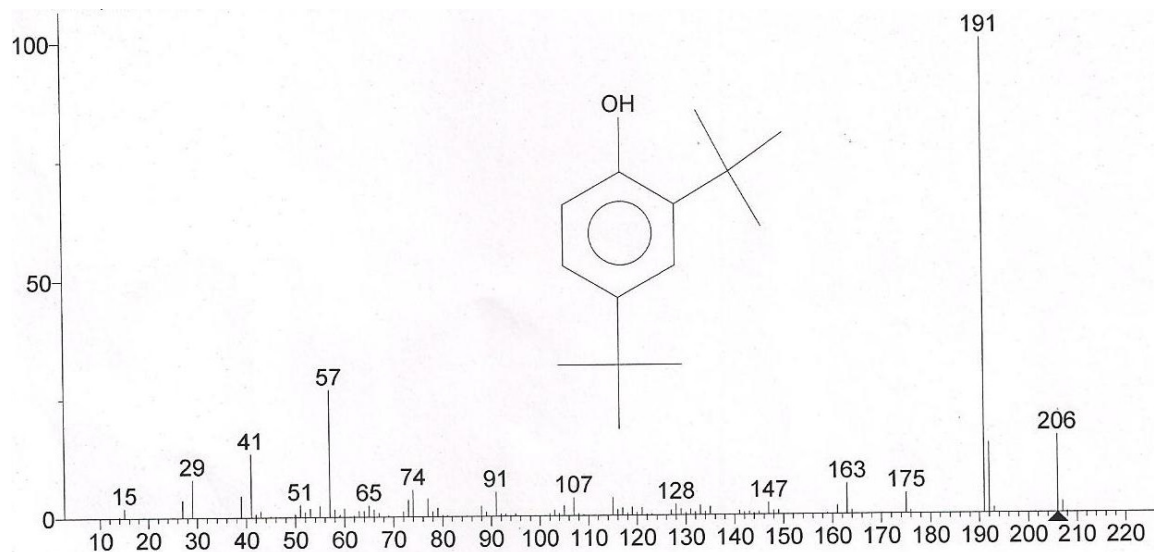


Figure 3. GC-MS mass spectrum and molecular structure of phenol, 2,4-bis(1,1-dimethylethyl).

Table 3. GC-MS indicated properties of phenol, 2,4-bis(1,1-dimethylethyl).

Properties	Given as
Molecular formula	C ₁₄ H ₂₂ O
Molecular weight	206
Biological activity	Antioxidant

Phenolic compounds have also been known as antioxidant agents, which act as free radical terminators and have shown medicinal activity as well as exhibiting physiological functions. It was reported that compounds such as flavonoids, which contain hydroxyls, are responsible for the radical scavenging effects of most plants (Omale and Okafor, 2007). The presence of these phytochemicals in *P. zeylanica* root is a significant finding in this present study.

REFERENCES

- Adekunle AS, Adekunle OC (2009). Preliminary assessment of antimicrobial properties of aqueous extract of plants against infectious diseases. *Biol. Med.*, 1(3): 20-24.
- Ahmad I, Mehmood Z, Mohammad F, Ahmad S (2000). Antimicrobial potency and synergistic activity of five traditionally used Indian medicinal plants. *J. Med. Arom. Plant Sci.*, 23: 173-176.
- Arunachalam KD, Velmurugan P, Raja RB (2010). Anti-inflammatory and cytotoxic effects of extract from *Plumbago zeylanica*. *Afri. J. Microbiol. Res.*, 4(12): 1239-1245.
- Dai Y, Hou L, Chan Y, Cheng L, But PP (2004). Inhibition of immediate allergic reactions by ethanol extract from *Plumbago zeylanica* stems. *Biol. Pharmacol. Bull.*, 27(3): 429-432.
- Evans WC (1996). *Trease and Evans Pharmacognosy*, 14th Edition, Bailliere Tindal W.B. Saunders company ltd; London, pp. 224-228, 293-309, 542-575.
- Jetty A, Subhakar C, Rajagopal D, Jetty M, Subramanyam M, Murthy MM (2010). Antimicrobial activities of neo- and 1-epineoisoshinanolones from *Plumbago zeylanica* roots. *Pharm. Biol.*, 48(9): 1007-1011.
- Jeyachandran R, Mahesh A, Cindrella L, Sudhakar S, Pazhanichamy K (2009). Antibacterial activity of Plumbagin and root extracts of *Plumbago zeylanica*. *Acta Biol. Cracoviensia Ser. Botan.*, 51(1): 17-22.
- Jiangsu New Medical College (1979). *Zhonyao Dictionary* (Encyclopedia of Chinese Materia). Scientific and Technological Press, Shanghai, pp. 711-712.
- Julsing KM, Quax JW, Kayser O (2007). *The Engineering of Medicinal Plants: Prospects and Limitations of Medicinal Plant Biotechnology*. In Oliver Kayser and Wim J Quax, ed. *Medicinal Plant Biotechnology*. WILEY-VCH Verlag GmbH and Co. KGaA, Weinheim, 978-3-527-31443-0.
- Kalidass C, Daniel A, Mohan VR (2010). Rapid propagation of *Plumbago zeylanica* L. An important medicinal plant. *J. Am. Sci.*, 6(10): 1027-1031.
- Kumar PP, Kumaravel S, Lalitha C (2010). Screening of antioxidant activity, total phenolics and GC-MS study of *Vitex negundo*. *Afr. J. Biochem. Res.*, 4(7): 191-195.
- Kumar R, Kumar S, Patra A, Jayalakshmi S (2009). Hepatoprotective activity of aerial parts of *Plumbago zeylanica* Linn against carbon tetrachloride-induced hepatotoxicity in rats. *Int. J. Pharm. Pharm. Sci.*, 1(1): 171-175.
- Lewis K, Ausubel FM (2006). Prospects for plant-derived antibacterials. *Nat. Biotech.*, 24(12): 1504-1507.
- Lin LC, Yang LL, Chou CJ (2003). Cytotoxic naphthoquinones and plumbagic acid glucosides from *Plumbago zeylanica*. *Phytochemistry*, 62: 619-622.
- Mehmood Z, Ahmad I, Mohammad F, Ahmad S (1999). Indian medicinal plants: A potential source of anticandidal drugs. *Pharm. Biol.*, 37: 237-242.
- Olagunju JA, Fagbohunka BS, Oyedapo OO, Abdul AIA (2006). Effects of an ethanolic root extract of *Plumbago zeylanica* Linn. on some serum parameters of the rats. *RPMP-Drug Dev. Mol.*, 11: 268-276.
- Olagunju JA, Jobi AA, Oyedapo OO (1999). An investigation into the biochemical basis of the observed hyperglycaemia in rats treated with ethanolic root extract of *Plumbago zeylanica*. *Phytother. Res.*, 13: 346-348.
- Omale J, Okafor PN (2007). Comparative antioxidant capacity, membrane stabilization, polyphenol composition and cytotoxicity of the leaf and stem of *Cissus multistriata*. *Afr. J. Biotechnol.*, 7(17): 3129-3133.
- Oyedapo OO (1996). Studies on the bioactivity of the extract of *Plumbago zeylanica*. *Phytother. Res.*, 13: 346-348.
- Raimi MM, Oyedapo OO (2009). Bioactivity-guided evaluation of the root extract of *Plumbago zeylanica*. *Int. J. Biol. Chem. Sci.*, 3(4): <http://ajol.info/index.php/ijbc/article/view/47151>.
- Sharma I, Gusain D, Dixit VP (1991). Hypolipidaemic and antiatherosclerotic effects of plumbagin in rabbits. *Indian J. Physiol. Pharmacol.*, 35: 10-14.
- Simonsen HT, Nordskjold JB, Smitt UW, Nyman U, Palpu P, Joshi P, Varughese G (2001). *In vitro* screening of Indian medicinal plants for antiplasmodial activity. *J. Ethnopharmacol.*, 74: 195-204.
- Uma DP, Soloman FE, Sharda AC (1999). Indian medicinal plants and their roots. *Pharm. Biol.*, 37: 231-236.