academicJournals

Vol. 9(35), pp. 918-921, 17 September, 2015

DOI: 10.5897/JMPR2015.5786 Article Number: FAB4AF955341

ISSN 1996-0875 Copyright © 2015 Author(s) retain the copyright of this article http://www.academicjournals.org/JMPR Journal of Medicinal Plants Research

Full Length Research Paper

Isoliquiritigenin and β-sitosterol from *Cissus polyantha*Tuber Glig and Brandt

Y. M. Sani¹, A. M. Musa¹, N. Tajuddeen^{3*}, S. M. Abdullahi¹, M. I. Abdullahi², U. U. Pateh¹ and A. Y. Idris¹

¹Department of Pharmaceutical and Medicinal Chemistry, Ahmadu Bello University, Zaria, Nigeria. ²Department of Pharmaceutical and Medicinal Chemistry, Usmanu Danfodio University, Sokoto, Nigeria. ³Department of Chemistry, Ahmadu Bello University, Zaria, Nigeria.

Received 2 March, 2015; Accepted 9 August, 2015

Chemical investigation into the composition of *Cissus polyantha* tuber, a plant used in Northern Nigerian ethno medicine for managing inflammatory conditions and conjunctivitis has led to the isolation of a chalcone from the ethyl acetate and a steroid from the hexane soluble fractions of its methanol extract. The structures of these compounds were elucidated by extensive analysis of their spectroscopic data, including 1 and 2D nuclear magnetic resonance (NMR). This is the first report of the isolation of the compounds from the plant and they may be responsible for the previously reported anti-inflammatory effects.

Key words: Cissus polyantha, vitaceae, 2D nuclear magnetic resonance (NMR), chalcone, steroid.

INTRODUCTION

Medicinal plants are known to provide a rich source of raw materials used in traditional medicine practice in Africa and other parts of the developing world. Also, research and development from traditional medicinal preparations have led to the discovery of many potent drugs which are used in modern clinical practice (Burkill, 2000). Previous studies have shown that 61% of some 877 drugs introduced worldwide can be traced or were inspired by natural products (Cseke et al., 2004). Cissus polyantha is a semi-woody climber; found usually in closed-forest from Sierra Leone to Southern Nigeria and also from Eastern Cameroun to Ubangi (Burkill, 2000). C. polyantha has been used in the management of inflammatory conditions and diseases related to bacterial

infections (Burkill, 2000). Previous biological studies on the plant have established its analgesic, antiinflammatory and antimicrobial activities (Sani et al., 2013). In furtherance of our phytochemical investigations into *C. polyantha*, this paper reports the isolation of a chalcone from the ethyl acetate and a steroid from the hexane soluble fractions of the methanol extract of the tuber of the plant.

MATERIALS AND METHODS

General procedures

Nuclear magnetic resonance (NMR)-spectra were recorded on a

*Corresponding author. E-mail: ntajuddeen@yahoo.com. Tel: +2347036585444.

Author(s) agree that this article remain permanently open access under the terms of the <u>Creative Commons Attribution</u>
License 4.0 International License

Bruker AVANCE spectrometer (400 MHz) for $^1\text{H-}$ and $^{13}\text{C-NMR}$ (100 MHz). Residual solvent signal with chloroform was used as the internal standard and chloroform was used as the solvents. Melting points were determined on a Yanaco MP-400 micro melting point apparatus. For thin layer chromatography (TLC) analysis, silica gel 60 F₂₅₄ (Merck) was used, column chromatography was performed using Merck silica gel (60 to 120) mesh, while gel filtration chromatography was performed using Sephadex LH-20 (Sigma, Spruce street, St. Louis, USA). Spots on TLC plates were visualized by spraying with 10% H₂SO₄ followed by heating at 100°C for 2 min.

Plant

Whole plant of *C. polyantha* growing in the wild was collected from Turunku village in Igabi Local Government Area, Kaduna State, Nigeria, in the month of June, 2009. It was authenticated by Malam U. S. Galla of the herbarium unit of the Biological Sciences Department, Ahmadu Bello University, Zaria, Nigeria, by comparing with an existing specimen (voucher No. 616). The tuber was separated from the plant, washed, sliced, air-dried and ground into powder using pestle and mortar, and subsequently referred to as powdered plant material.

Extraction and isolation

The powdered plant (1000 g) was extracted with 75% methanol in a Soxhlet apparatus for 48 h, the solvent was removed in-vacuo to yield a residue (80 g) referred to as crude methanol extract of the tuber of C. polyantha (CMET). The methanol extract (70 g) was suspended in water and partitioned successively with hexane, chloroform and ethyl acetate to obtain the hexane and ethyl acetate soluble fraction used for the study. The ethyl acetate fraction (7.0 g) was chromatographed on silica gel (60-120 mesh), and the packed column was eluted using gradient solvent systems of n-Hexane 100%, hexane/chloroform mixtures, chloroform 100%, and chloroform/ethyl acetate mixtures. 42 fractions of 100 ml each were collected. The fractions were pooled together based on their TLC profile to give 5 sub fractions. Repeated Sephadex LH-20 (MeOH) gel filtration chromatography of sub fraction 3 led to the isolation of compound 1 (6.2 mg). Similarly, the hexane fraction (7.0 g) was chromatographed on a silica gel column eluting with 100% hexane, hexane/chloroform mixtures and 100% chloroform as solvent systems to give 37 fractions. The 37 fractions were pooled together based on similarities in their TLC profiles to give 5 sub fractions. Preparative TLC of sub fraction 3 led to the isolation of compound 2 (7.4 mg).

RESULTS AND DISCUSSION

Compound 1 was isolated as a yellow solid with a melting point of 202 to 204°C; it gave a positive result to the Shinoda test for flavonoids. The ¹H NMR spectrum of compound 1 revealed signals for a pair of ortho coupled protons at $\vec{\omega}_{\rm H}$ 7.9 (d, 1H, J = 8.94 Hz, H-5'), $\vec{\omega}_{\rm H}$ 6.43 (dd, 1H, J = 8.9, 2.2 Hz, H-6') and a third signal at $\vec{\omega}_{\rm H}$ 6.25 (d, 1H, J = 2.2 Hz, H-3') which was a meta coupled to $\vec{\omega}_{\rm H}$ 6.43 (dd, 1H, J = 8.9, 2.2 Hz), assignable to the tri-substituted benzene ring A. The signals at $\vec{\omega}_{\rm H}$ 7.63 (d, 2H, J = 8.4 Hz, H-2, 6) and $\vec{\omega}_{\rm H}$ 6.85 (d, 2H, J = 8.4 Hz, H-3, 5) were

assigned to the 1,4 di substituted ring B. Two transcoupled olefinic proton at $\sqrt[3]{H}$ 7.8 (d, 1H, J = 15.4 Hz, H- α) and $\delta_H 7.6$ (d, 1H, J = 15 Hz, H- β) were also assigned. These proton NMR signal assignments were supported by H¹-H¹ COSY correlations of H-5'/H-6', H-2/H-3 and Hα/H-β. The ¹³C spectrum of compound 1 revealed a total of 12 signals, which were assigned to a ketone carbonyl at δ_C 193.30, the remaining signal were all for unsaturated carbon atoms at $\delta_{\rm C}$ 102.55, 115.54, 116.99, 119.30 126.45, 130.39, 131.93, 144.08, 160.00, 160.20 and 165.00. They were assigned to two benzene rings and a two carbon atoms unsaturated olefinic system. The downfield signals at δ_C 144.08, 160.00 and 165.00 suggested that they are oxygenated carbon atoms. These ¹H and ¹³C signals were very similar to those of isoliquiritigenin and a comparison of these data with those reported in the literature for isoliquiritigenin showed very close agreement (Markham and Ternai, 1976; Aida et al., 1990; Sato et al., 2007), hence the structure of compound 1 was determined to be isoliquiritigenin (Figure 1).

Compound 2 was isolated as a white amorphous solid with a melting point of 137 to 139°C; it gave a positive result to the Salkowski's test for steroid/triterpenes. Its ¹H-NMR spectrum revealed three regions typical of the steroidal nucleus at 0.5 to 2.5 ppm representing overlapping methyl, methylene and methine protons; an oxymethine proton at 3.5 ppm was assigned to the position 3 of the steroidal nucleus and a single unsaturated proton signal at 5.8 ppm. The ¹³C NMR spectrum of compound 2 revealed a total of 29 carbon signals, 6 of which were methyl signals, 11 were methylene carbon signals, 9 were methine and there were 3 quaternary signals, these data is typical of βsitosterol (Rowshanul et al., 2007; Pateh et al., 2009; Li et al., 2009; Hamada et al., 2012). The signal between 11.8 and 56.7 ppm represents a region of overlapping methyl, methylene and methine carbon atoms, an oxymethine signal at 71.8 was typical of position 3 of βsitosterol, and finally the unsaturated carbon signals at 121.7 and 140.7 ppm were assigned to a two carbon olefinic system. These NMR data are very similar to the data for β-Sitosterol and a comparison with data reported for β-sitosterol showed good agreement (Rowshanul et al., 2007; Pateh et al., 2009; Li et al., 2009; Hamada et al., 2012). Also, Co-TLC analysis of compound 2 with standard sample of β -sitosterol showed them to have the same R_f value. Therefore, the structure of compound 2 was determined to be β -sitosterol (Figure 2).

A lot of studies on the biological activities of isoliquiritigenin and β -sitosterol have been reported in the literature; isoliquiritigenin has been reported to have several biological activities, including anti inflammatory, anti oxidant, anti tumor and anti spasmodic activities (Sato et al., 2007; Li et al., 2009; Hamada et al., 2012; Kim et al., 2008). β -sitosterol in addition to its well known

Figure 1. Isoliquiritigenin.

Figure 2. β-sitosterol.

anti-inflammatory effect has been reported to have analgesic, anti-oxidant, anti-diabetic, anthelminthic and anti-mutagenic activities (Saeidnia et al., 2014). These two compounds might therefore be partly responsible for the observed biological activities previously reported for *C. polyantha*, and their presence justifies the ethno medicinal claims on the plant.

Conclusion

The isolation of these two compounds, well known for their anti inflammatory and analgesic activities lends full justification to the ethno medicinal use of the plant in the treatment of inflammatory conditions.

Conflict of interest

The authors declare that they have no conflict of interest.

ACKNOWLEDGEMENT

The authors are grateful to Mr Dilip Jagjivan of the School of Chemistry and Physics, University of Kwa-Zulu Natal, Durban South Africa for assisting with running the NMR spectroscopy.

REFERENCES

Aida K, Tawata M, Shindo H, Onaya T, Sasaki H, Yamaguchi T, Chin

- M, Mitsuhashi H (1990). Isoliquritigenin: a new aldose reductase inhibitor from *glycyrrhizae radix*. Planta Med. 56:254-258.
- Burkill HM (2000). The Useful plants of West Tropical Africa, 5th edn. Royal botanic garden Kew, UK p 206.
- Cseke LJ, Kaufuman BP, Podila GK, Tsai CJ (2004). Molecular and cellular Methods in Biology and Medicine, 2nd edn. CRC press, Boca Roton, Florida p 95.
- Hamada H, Soumia M, Catherine L, Mohammed B, (2012). Chemical constituents of *Centaurea omphalotricha* Coss and Durieu ex batt and Trab. Rec. Nat. Prod. 6:292-295.
- Kim JY, Park SJ, Yun KJ, Cho YW, Park HJ, Lee KT, (2008). Isoliquiritigenin isolated from the roots of *Glycyrrhiza uralensis* inhibits LPS-induced iNOS and COX-2 expression via the attenuation of NF-kappa B in RAW 264.7 macrophages. Eur. J. Pharmacol. 584:175-184.
- Li D, Wang Z, Chen H, Wang J, Zheng Q, Shang J, Li J (2009). Isoliquiritigenin induces monocytic differentiation of HL-60 cells. Free Radic. Biol. Med 46:731-736.
- Markham KR, Ternai B (1976). 13C NMR of flavonoids II: flavonoids other than flavones and flavonol aglycones. Tetrahedron 32:2607-2612
- Pateh UU, Haruna AK, Garba M, Iliya I, Sule IM, Abubakar M, Ambi AA (2009). Isolation of stigmasterol, sitosterol and 2-hydroxyhexadecanoic acid methyl ester from the rhizome of *Stylochiton lacifolius* Pyer and Kostchy (Araceae). Niger. J. Pharm. Sci. 7(1):19-25.

- Rowshanul MH, Farjana N, Matiar R, Ekramul MH, Rezaul MK (2007). Isolation of Stigmasterol and β-Sitosterol from Methanolic Extract of Root Bark of *Calotropis gigantea* (Linn). Pak. J. Biol. Sci. 10:4174-4176.
- Sani YM, Musa AM, Abdullahi SM, Atiku I, Abdullahi MS, Hanwa UA, Sani MB, Hussaini AO (2013). Phytochemical and antimicrobial studies of methanol crude extract of the leaves of *Cissus polyantha* (vitaceae). Niger. J. Pharm. Sci. 12:33-38.
- Sato Y, He J, Nagai H, Tani T, Akao T (2007). Isoliquiritigenin, one of the antispasmodic principles of Glycyrrhiza ularensis roots, acts in the lower part of intestine. Biol. Pharm. Bull. 30:145-149.
- Saeidnia S, Manayi A, Gohari AR, Abdollahi M (2014). The Story of Beta-sitosterol- A. Review. Eur. J. Med. Plants 4(5):590-609.