

Full Length Research Paper

Preliminary study on the establishment of reference intervals for serum prolactin for women in Zaria, Nigeria

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Received 13 March, 2015; Accepted 30 November, 2015

The interpretation of prolactin results is difficult as race related reference intervals are scarce, especially in Nigerian women. Serum prolactin reference intervals were determined from 120 subjects. Subjects were randomly selected from antenatal clinics and general outpatient clinics in one private hospital and three government hospitals in Zaria, Northern Nigeria. They consisted of six groups made up of women that were A-non pregnant, B-first trimester of pregnancy, C-second trimester of pregnancy, D-third trimester of pregnancy, E-labour, and F-puerperium. Their blood samples were collected via venipuncture between 09.00 and 13.00 h after which they were centrifuged and the sera stored at -8°C for further analysis. The samples were analyzed at the Department of Chemical Pathology, Ahmadu Bello University Teaching Hospital, Shika. Serum prolactin was determined with Prolactin quantitative test kits using the principle of solid phase enzyme-linked immunosorbent assay. The concentration of prolactin is proportional to the color intensity of the samples as measured spectrophotometrically at a wavelength of 450 nm. Informed consent and ethical clearance was obtained before commencement of the study. Results were presented as mean \pm standard deviation (SD) and data analyzed using one-way analysis of variance, while post-hoc test was carried out. A level of significance of $p < 0.05$ was considered statistically significant, while the reference range was defined as 95% confidence interval. Serum prolactin results showed a significant increase in pregnant women as compared to the non-pregnant, not lactating subjects (20.24 \pm 3.41 ng/ml; 95% Confidence Interval (CI): 18.84-21.65 ng/ml). There were also significant trimester related differences in serum prolactin with prolactin levels increasing during the first trimester though not significantly (29.84 \pm 5.08 ng/ml). A sharp increase was observed during the second trimester (96.09 \pm 18.82 ng/ml; 95% CI: 89.41 to 102.76 ng/ml). There was a further increase at the third trimester (171.11 \pm 32.92 ng/ml; 95% CI: 159.03-183.18 ng/ml), a peaking during labour (198.37 \pm 15.81 ng/ml; 95% CI: 186.22-210.52 ng/ml) with a significant decline during the first week of the puerperium (197.19 \pm 22.11 ng/ml; 95% CI: 183.83-210.55 ng/ml). Apparent changes in serum prolactin levels were similar to that of previous studies; although the ranges for all groups were observed to be narrower as compared to those of other studies.

Key words: Serum prolactin, reference ranges, enzyme immunoassay, Zaria.

INTRODUCTION

Prolactin is a versatile peptide hormone, best known for its role in lactation. Prolactin also has widespread effects

amongst which are its profound effects on angiogenesis, leucopoiesis, hematopoiesis, modulation of T- and B- cell

function and as an anti-apoptotic factor (Reuwer et al., 2012; Terra et al., 2011; Ben-Jonathan et al., 2008; Welniak et al., 2001; Nagy and Berczi, 1989). Prolactin has been reported to play a role in the aetio-pathogenesis of autoimmune diseases like systemic lupus erythromatosis, rheumatoid arthritis and multiple sclerosis (Da Costa et al., 2011; Jara et al., 2011; Orbach and Shoenfeld, 2007; Jara et al., 2001; Neidhart et al., 1999). Altered prolactin levels associated with either T-helper cell type 1 and type 2 dominance often characterize autoimmune diseases (Orbach and Shoenfeld, 2007). Pro-carcinogenic effects of prolactin has also been reported (Alhaj, 2012; Manjer et al., 2003). Causes of hyperprolactinemia, include physiological states such as increased prolactin levels in the morning, after exercise, after meals and following sexual intercourse. Increased prolactin also occurs following epileptic seizures, in stress, in expectant fathers as well as in pregnancy and lactation (Kruger et al., 2012; Rojas et al., 2012; Díaz et al., 2013; Ben-Menachem, 2006; Van Cauter and Spiegel, 1997). A circadian and ultradian rhythm is responsible for these fluctuations (Fujimoto et al., 1990; Van Cauter, 1990). The ubiquitous distribution of prolactin receptors and the extrapituitary synthesis of prolactin is responsible for its numerous roles (Marano and Ben-Jonathan, 2014). Assays of prolactin are carried out as part of investigations of hypogonadism in males, erectile dysfunction in males, infertility in women and some metabolic diseases in children (Catli et al., 2012; Seaborg, 2012; Zeitlin and Raifer, 2000).

Guidelines determining prolactin reference range for men and women have been defined (Seaborg, 2014; Katayev et al., 2010; Jeffcoate et al., 1986). However, different assays and methods for measuring prolactin are employed by different laboratories. As such, the serum reference range for prolactin is often determined by the laboratory performing the measurement. Prolactin levels also vary with several variables, such as age, sex, menstrual cycle stage, lactation, pregnancy as well as racial and ethnic variability has been demonstrated (Kim et al., 2012; Tanner et al., 2011; Sachidhanandam et al., 2009; Jeffcoate et al., 1986).

Thus, accurate interpretation of the measurement is dependent on the circumstances around the given measurement.

Reference intervals estimate the expected values that would contain the 95% of the subjects of the considered population (Baron et al., 1989). Reference ranges utilized in Nigerian laboratories are as specified in the kits without further development of local laboratory reference range. However, there is a need for developing one due to the need to compensate for calibration errors in equipment, compensating for errors in the laboratory methodology,

errors in the statistical method used and other variability as suggested by Katayev et al. (2010), Nakayama (1992) and Gaines Das and Cotes (1979).

A previous study on the establishment of reference ranges for some reproductive hormones in Nigerian women including prolactin assays (Amballi et al., 2007). Their study using enzyme immune assay excluded pregnant women, women in labour and the puerperium. This study thus aimed at determining possible difference in the reference ranges for serum prolactin among Nigerian women who were non-pregnant, pregnant, in labour, and the puerperium in Zaria, Northern Nigeria.

MATERIALS AND METHODS

Study site

The study was conducted among women in Zaria, Northern Nigeria. Zaria is a town located within latitude 11°3'N and longitude 7° 42'E. It is comprised of 2 local government areas with an altitude of 610 m, an annual rainfall of 1056.6 mm and a mean annual temperature of 27°C (Mortimore, 1970).

Study design

This study is a cross-sectional, multicenter study. Women were recruited from antenatal clinics, delivery rooms and lying-in wards of five hospitals in Zaria. They consisted of Primary Health Centre, Samaru; Salama Hospital and Maternity, Kwangila; Sabon Gari Comprehensive Health Centre; St Luke's Hospital, Wusasa and Ahmadu Bello University Health Centre, Samaru, Zaria. While the control subjects consisted of individuals from Ahmadu Bello University (students and staff). Approval for the study was obtained from the Ethical Committee on Human Study of the Kaduna State, Ministry of Health. All participants provided informed consent.

Study subjects

A total of one hundred and twenty healthy female subjects participated in the study. Twenty-five subjects served as control for the study. They were non-pregnant, non-lactating women in their reproductive age who were not on hormonal contraceptives. A total of ninety-five women were either pregnant, in labour or in the puerperium. All pregnancies were dated to the last menstrual period. Nine women were in their first trimester, 33 women were in their second trimester while thirty-one women were in their third trimester. Nine women were in labour and 13 were women in puerperium; eight of which had spontaneous vaginal delivery (SVD) and five had a caesarean section (CS). Subjects who were diabetic or hypertensive (and thus on medication) were excluded from the study. A questionnaire was administered to all participants to obtain bio-socio-demographic data while weight, height, blood pressure and blood samples were collected.

Weight, height and body mass index assessment

The weights of the subjects were measured while wearing light

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clothing to the nearest 0.2 kg with a calibrated weighing scale. Height (without shoes and head attire) was measured to the nearest 0.5 cm with a stadiometer. The body mass index was calculated as weight (kg)/height (m²) (Guyton and Hall, 2006)

Blood pressure measurement

The arterial blood pressure in the brachial artery was measured after seating the subjects for 5 min with legs uncrossed. A mercury sphygmomanometer (Acosson, A. C. Cossor & Son (surgical) LTD, London) cuff was wrapped round the arm of the subjects and inflated while a 3M Littmann Classic II S.E. Stethoscope (3M Health Care, U.S.A) was used to auscultate. Auscultatory blood pressure measurements were determined using the onset of the first and the disappearance of the fifth Korotkoff sounds denotes the systolic and diastolic blood pressure, respectively (Pickering et al., 2005).

Urinalysis

A fresh mid-stream urine sample was collected from the participating subjects into sterile, clean and dry bottles. The urine samples were immediately tested by inserting a multistrip (3) urinalysis strip affixed with chemically specific reagent pads. Detectable levels of protein and glucose and the urine pH was determined by visually comparing the color reaction with the included color chart to determine the level of each chemical factor.

Sample collection

Participants were instructed about the procedure on arrival at the health facility. They rested quietly for a minimum duration of 30 min; other parameters were also obtained on the same day (for some participants they proceeded for their antenatal visits thereafter). The blood samples were taken between 09.00 and 12.00 h. Five milliliters of venous blood was collected at the cubital fossa using a 5 ml syringe and a 23G needle. The sample was then transferred into a clean, plain and dry bottle which was centrifuged using a bench centrifuge at 3000 rpm for 5 min. The sera were transferred into plain bottles and frozen at <8°C till assay was carried out.

Prolactin assay

Serum prolactin concentration was determined for the samples using Prolactin Quantitative Test Kits. The principle of the test was based on solid phase enzyme linked immunosorbent assay. The level of prolactin was determined by measuring the detectable color change using a Microwell reader at 450 nm (Batzner, 1980). The assay was carried out at the Department of Chemical Pathology, Ahmadu Bello University Teaching Hospital, Shika, Zaria.

Data analysis

Results were presented as frequencies, percentages and mean \pm standard deviation (SD), while data was analyzed using one-way analysis of variance using SPSS version 22, followed by a Tukeys' Post-hoc test, while the reference range (defined as 95% confidence limits) was determined as arithmetic mean \pm 2 SD. Results were considered statistically significant with $p < 0.05$.

RESULTS

The result is for 120 subjects comprising non-pregnant

controls, women in their first, second, and third trimesters, women in labour and the puerperium. The data presented includes the mean \pm SD and the central 95 percentile of the results of the apparently healthy Nigerian women in Zaria (Tables 1, 2 and 3).

Women below 25 years constituted the highest proportion of the study subjects (52%). While a large proportion of the subjects were Hausa women (49%). All the women had some form of education with the tertiary education group constituting the highest proportion (38%). Most of the subjects were housewives (35%) while the least were artisans (5%).

The mean age of the women in labour and the puerperium was significantly higher than that of the control ($p < 0.05$). There was also a significant increase in weight in women in labour as compared to the non-pregnant control, pregnant women and women in puerperium ($p < 0.05$). There was no significant difference in the height and BMI of the women across all groups ($p > 0.05$). Mean systolic blood pressure significantly increased among women in labour and the puerperium as compared to the non-pregnant control ($p < 0.05$). A significant decrease in the mean arterial pressure and diastolic blood pressure was observed in pregnant women and while women in labour had an observed significant increase in the mean arterial pressure and diastolic blood pressure as compared to the control non-pregnant women ($p < 0.05$).

There was a significant increase in mean serum prolactin concentration in women in the 2nd trimester, 3rd trimester, labour and puerperium as compared to the control non-pregnant group ($p < 0.05$). However, there was no significant difference in mean serum prolactin concentration between the non-pregnant control women and the first trimester pregnant women nor between the women in labour and the puerperium ($p > 0.05$).

DISCUSSION

Pregnancy, labour and the puerperium induces a number of physiological variations in most body systems. Some of these changes are marked and hence a need for reference ranges for women across these transitional phases of their reproductive life. Prolactin hormone plays a role in establishing and maintaining lactation amongst its several functions (Freeman et al., 2000). A preliminary cross-sectional study to establish reference ranges of serum prolactin was undertaken in this study.

A large proportion of the women in the study were below 25 years. This is as a result of the region of the study area (Northern Nigeria) which is reputed for high rates of teenage marriages and child bearing which has resulted in a high total fertility rate of over 7 births per woman (National Population Commission, 2009; Wolf et al., 2008).

Most of the women were of Hausa ethnicity (49.2%). The

Table 1. The socio-demographic details of the participants; non-pregnant, pregnant, in labour and in the puerperium women.

Variable	Group								Totals	
	Non-pregnant (n=25)		Pregnant(n=73)		Labour(n=9)		Puerperium(n=13)			
	Frequency	Percentage (%)	Frequency	Percentage (%)	Frequency	Percentage (%)	Frequency	Percentage (%)		
Age group (years)										
<20	5	20	19	76	0	0	1	4	25	
21-25	10	27	22	59.5	1	2.7	4	10.8	37	
26-30	4	13.8	22	79.9	2	6.9	1	3.4	29	
>31	6	20.7	10	34.5	6	20.7	7	24.1	29	
Ethnicity										
Hausa	2	3.4	43	72.9	5	8.5	9	15.3	59	
Igbo	3	30	5	50	2	20	0	0	10	
Yoruba	9	52.9	7	41.2	0	0	1	5.9	17	
Others	11	32.4	18	52.9	2	5.9	3	8.8	34	
Educational status										
Qur'an	0	0	19	76.0	3	12	3	12	25	
Primary	0	0	3	37.5	2	25	3	37.5	8	
Secondary	3	7.5	28	70	4	10	5	12.5	40	
Tertiary	22	47.8	23	50	0	0	1	2.2	46	
Nil	-	-	-	-	-	-	-	-	-	
Occupation										
Civil Servant	8	53.3	7	46.7	0	0	0	0	15	
Trader	0	0	17	73.9	2	8.7	4	17.4	23	
Student	17	50	16	47.1	0	0	1	2.9	34	
House Wife	0	0	30	71.4	5	11.9	7	16.7	42	
Artisan	0	0	3	50	2	33.3	1	16.7	6	
Total	100	358.8	292	988.5	36	165.6	51	190.2	479	

reason being that Zaria is located in Northern Nigeria, where one of the major ethnic groups is Hausa, a language used as a lingua franca in most of the region. An appreciable number of the women (38.3%) had attained tertiary education (Table 1). This is a marked improvement from the

study of Okereke et al. (2013), where less than 5% of the female respondents participating in their study (site also in Northern Nigeria) had attained tertiary education. Other researchers have observed a lower educational status for females in Northern Nigeria as compared to other regions of

the country (Korode, 2008; African Girl-Child Education Initiative, 2001; Umar, 1996). Their study was carried out in a rural setting. The findings of a high percentage of the women in our study area attaining a tertiary education (Table 1), is due to the fact that Zaria is an urban

Table 2. The age, anthropometric measurement and blood pressure of the subjects.

Parameter (mean±SD)	Non-pregnant women (n=19)	Pregnant women (n=46)	Women in labour (n=9)	Women in puerperium (n=12)
Age (years)	26.05±5.44	24.87±5.01	32.78±5.12*	30.75±6.34*
Height (cm)	163.73±8.47	161.76±11.35	165.50±1.41	161.13±2.43
Weight (kg)	58.84±11.12	63.39±13.22	78.38±3.11*	65.60±5.52
BMI (kg/m ²)	21.93±4.92	24.36±5.60	28.25±0.07	25.33±2.42
Systolic B.P (mmHg)	112.63±12.40	109.33±11.16	135.71±19.88*	133.00±20.03*
Diastolic B.P (mmHg)	77.89±9.18	70.11±9.80*	87.14±11.13*	91.00±13.70
Mean Arterial B.P (mmHg)	89.47±9.18	83.19±9.64*	103.33±13.19*	105.00±15.65

*P<0.05.

Table 3. Mean serum level of prolactin for all subjects; non-pregnant, pregnant, in labour and in puerperium women.

Group	Mean ± SD of serum prolactin concentration (ng/ml)	95% confidence interval for mean reference range (ng/ml)
Non-pregnant (n=25)	20.24±3.41	18.84 – 21.65
1 st Trimester (n=9)	29.84±5.08	25.94 – 33.75
2 nd Trimester (n=33)	96.09±18.82	89.41 – 102.76
3 rd Trimester (n=31)	171.11±32.92	159.03 – 183.18
Labour (n=9)	198.37±15.81	186.22 – 210.52
Puerperium (n=13)	197.19±22.11	183.83 – 210.55

educational town, with more than 30 educational and research institutions. This finding corroborated the study of a similar urban town in Northern Nigeria, where the number of women with an educational attainment of tertiary education (all females) constituted 58.3% of the respondents (Abdullahi and Tukur, 2013).

There was no significant difference in the height and BMI of the women across all groups. The finding infers that the women were BMI matched. This is an unanticipated feature, since pregnancy is associated with a significant increase in weight gain, due to contributions from the fetus, uterus, placenta and maternal fluid retention (Suitor, 1997; Ota et al., 2011).

A significant decrease in diastolic blood pressure and mean arterial blood pressure during pregnancy was observed (Table 2). These findings are physiological variations associated with pregnancy. The decrease is associated with a decrease in systemic vascular resistance. Pregnancy hormones (estradiol-17b, progesterone and relaxin) have been implicated in the vascular change (Conrad, 2011; Kristiansson and Wang, 2001). Relaxin especially, is said to cause a induce a rapid and sustained vasodilation by activation of endothelial nitric oxide synthase and increases in arterial gelatinase (s) activity; which still subsequently activates the endothelial (ET(B) receptor/nitric oxide vasodilatory pathway.

Blood pressure (systolic and diastolic) was understandably significantly higher in women during labour (Table 2). It has longed been recognized that

blood pressure rises during labour by 15 to 50 mmHg (Radcliffe, 1944). The rise occurs during uterine contractions and aids in maintaining a satisfactory placental circulation (Cooks and Briggs, 1903; Edwards, 1958).

In Nigeria, hyperprolactinemia is quite common in infertile patients (Meraiyebu et al., 2012; Olootu et al., 2012). It presents with galactorrhea, amenorrhea and sterility. Attempts have been made at establishing normal reference ranges for the Nigerian female population (Amballi et al., 2007; Iwamoto et al., 2004; Dada et al., 1981). These studies excluded women in other transitional stages of their reproductive cycle (pregnancy, labour, and the puerperium). Their studies entailed using radioimmunoassay method, comparing that with an enzyme immunoassay method, determining prolactin levels in men and women at various phases of their menstrual cycles. Pregnancy is associated with profound changes in laboratory markers amongst which are the reproductive hormones. A local reference range that describes the change across the non-pregnant, pregnant, labour and puerperium stages was the interest of this study.

In this study, serum prolactin levels in non-pregnant women were significantly lower than in other groups (Table 3).

Pregnant women in all trimesters presented with a higher concentration of serum prolactin as compared to their non-pregnant control group. The increase was with each increasing trimester. Estradiol levels, rising

throughout pregnancy, act at the hypothalamic level to increase prolactin secretion. The increase is about 10 to 20. During pregnancy, prolactin increases arginase activity, stimulate ornithine decarboxylase activity and enhance the rate of transport of polyamines into the mammary gland. The resultant effect is increased spermine and spermidine synthesis required for milk production (Voogt et al., 2001). From our study, non-pregnant women had a reference range of 18.84 to 21.65 ng/ml, while the pregnant women had a reference range of 25.94 to 33.75, 89.41 to 102.76, and 159.03 to 183.18 ng/ml for the first, second and third trimester, respectively. These pattern of the reference range values are similar to the review of Abbassi-Ghanavati et al. (2009) who obtained a reference range of serum prolactin for non-pregnant women of 0 to 20 ng/ml, and for pregnant women as 36 to 213, 110 to 330, and 137 to 372 ng/ml for first, second, and third trimesters, respectively). Though, the reference ranges for the subjects in our study area yielded lower values and narrower ranges. It was observed that the values were also similar to reference values (in non-pregnant females) provided in a study by Amballi et al. (2007), his study however, excluded pregnant women and women in their puerperium. Reference ranges for serum prolactin determined by Dada et al. (1981), had values that compared reasonably with those in our study.

In our study, there was a significant increase in serum prolactin in labour and puerperium as compared to the non-pregnant women. However, there was a slight decrease in mean serum prolactin among the women in puerperium when compared with those in labour that was however not statistically significant. Prolactin increases during pregnancy and reaches a peak 10 h before delivery (Onur et al., 1989; Biswas and Rodeck, 1976; Fang and Kim, 1975). In the first week postpartum, prolactin levels usually decline 50%. The decline is due to the abrupt drop of estrogen as well as progesterone allowing prolactin to induce lactation (Speroff and Fritz, 2005). Prolactin levels subsequently increase due to sucking which activates mechanoreceptors around the nipple resulting in stimulation of the hypothalamus and consequent increased pituitary secretion of prolactin (Diaz et al., 1989; Andersen et al., 1981). The findings are similar to the study by Godo et al. (1988) and Vemer and Roland (1981). The slight decrease in the puerperium and labour is due to the ineffectiveness of suckling during the early puerperium, the loss of estrogen and progesterone following delivery. The decrease in serum prolactin is more likely in our subjects who were all within the first week of puerperium when suckling is not so established. Subsequently, serum prolactin levels are known to increase as suckling becomes more established and eventually dwindle to near non-pregnant level by the sixth month of lactation (Riordan, 2005; Walker, 2006).

Limitations include inability to carry out a longitudinal study as opposed to the cross-sectional study undertaken; having its flaws. This study was a randomized study,

carried out in the Northern region of the country and thus the population might not necessarily be representative of the general population of women in Nigeria. Due to the small sample size and site of the study, we were unable to examine ethnicity differences in serum prolactin levels in major Nigerian tribes.

Conclusion

This is a preliminary cross-sectional study of reference intervals of serum prolactin spanning through pregnancy, labour and puerperium in Nigerian women in Zaria. Apparent changes in serum prolactin levels for all groups were observed to be lower and narrower as compared to those of other studies. This suggests that there may be racial differences in serum prolactin levels in women. It is envisaged that these intervals will aid in the monitoring of women in the different transitional phases of their reproductive period.

RECOMMENDATIONS

Further studies were recommended utilizing a larger sample size. A multi-centre study recruiting participants across the regions of the country will aid in deriving a reference range for Nigerian women. Studies including menopausal women in the study are also being suggested (due to the implication of serum prolactin women in breast cancer risk especially in menopausal women). More pertinent is the need for a longitudinal study following up the serum changes in prolactin from a non-pregnant state, through pregnancy and the puerperium in Nigerian women. Serum prolactin reference ranging in Nigerian children has also not been previously elucidated and is being suggested as considerations of another study. A study comparing serum prolactin levels in Caucasians, Nigerians and other Africans, utilizing the same laboratory is also being suggested.

Conflict of Interests

The authors have not declared any conflicts of interests.

ACKNOWLEDGEMENTS

The authors wish to appreciate the participants of the study, the various health professionals at the respective health services and the ethical committee of the Ministry of Health, Kaduna for their co-operation in the study.

REFERENCES

Abbassi-Ghanavati M, Greer LG, Cunningham FG (2009). Pregnancy and laboratory studies: a reference table for clinicians. *Obstet Gynecol.*

- 114(6):1326-1331.
- Abdullahi H, Tukur J (2013). Sexual stimulants and their effects on women of reproductive age group in Kano, Northern Nigeria. *Niger. J. Basic Clin. Sci.* 10(1):13-16.
- African Girl-Child Education Initiative (2001). Yearly Technical Report, Abuja Nigeria.
- Alhaj A (2012). Serum Prolactin Level in Yemeni Females with Breast Cancer Yemeni. *J. Med. Sci.* 6:1-6.
- Amballi AA, Dada OA, Adeleye AO, Salu J (2007). Evaluation of the determination of reference ranges for reproductive hormones (prolactin, FSH, LH, and testosterone) using enzyme immuno assay method. *Sci. Res. Essay* 2(4):135-138.
- Andersen AN, Tabor A, Hert JB, Schioler V (1981). Abnormal prolactin levels and pituitary-gonadal axis in the puerperium. *Obstet. Gynecol.* 57(6):725-729.
- Baron DN, Whicher JT, Lee KE (1989). Reference ranges, normality and abnormal results; Short textbook of Chemical Pathology. 5th edition. 5-8.
- Batzer FR (1980). Hormonal evaluation of early pregnancy. *Fert. Steril.* 34(1):1-13.
- Ben-Jonathan N, LaPensee CR, LaPensee EW (2008). What can we learn from rodents about prolactin in humans? *Endocr. Rev.* 29:1-41.
- Ben-Menachem E (2006). Is prolactin a clinically useful measure of epilepsy? *Epilepsy Curr.* 6(3):78-79.
- Biswas S, Rodeck CH (1976). Plasma prolactin levels during pregnancy. *Br. J. Obstet. Gynaecol.* 83:683-687.
- Catli G, Abaci A, Altincik A, Demir K, Can S, Buyukgebiz A, Bober E (2012). Hyperprolactinemia in children: clinical features and long term results. *J. Pediatr. Endocrinol. Metab.* 25(11-12):1123-1128.
- Conrad KP (2011). Maternal vasodilation in pregnancy: the emerging role of relaxin. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 301(2):R267-R275.
- Cooks HW, Briggs JC (1903). Clinical observations on blood pressure. *Johns Hopkins Hosp. Rep.* 11:451-455.
- Dada OA, Ladipo OA, Osinusi AO, Osoimein BO, Nduka EV (1981). Circulating blood levels of gonadotropins and prolactin in the normal menstrual cycle. *Int. J. Gynaec. Obstet.* 19:291-294.
- Da Costa R, Szyper-Kravitz M, Szekanecz Z, Csepány T, Danko K, Shapira Y, Zandman-Goddard G, Orbach H, Agmon—Levin N, Shoenfeld Y (2011). Ferritin and prolactin levels in multiple sclerosis. *Isr. Med. Assoc. J.* 13:91-95.
- Díaz L, Díaz-Muñoz M, González L, Lira-Albarrán S, Larrea F, Méndez I (2013). Prolactin in the Immune System. In: Nagy GM, Toth BE, editors. E-book- Obstetrics and Gynecology, Prolactin – ISBN 978-953-51-0943-3. Under CC BY 3.0 license. Copyright the Authors.
- Diaz S, Seron-Ferre M, Cardenas H, Schiappacasse V, Brandeis A, Croxatto HB (1989). Circadian variation of basal plasma prolactin, prolactin response to suckling, and length of amenorrhea in nursing women. *J. Clin. Endocrinol. Metab.* 68(5):946-955.
- Edwards EM (1958). The Blood pressure in Normal. *Br. J. Obst. Gynecol.* 65(3):367-370.
- Fang VS, Kim MH (1975). Study on maternal, fetal and amniotic human prolactin at term. *J. Clin. Endocrinol. Metab.* 41:1030-1034.
- Fujimoto VY, Clifton DK, Cohen NL, Soules MR (1990). Variability of serum prolactin and progesterone levels in normal women: the relevance of single hormone measurements in the clinical setting. *Obstet. Gynecol.* 76(1):71-78.
- Freeman ME, Kanyicska B, Lerant A, Nagy G (2000). Prolactin: Structure, Function, and Regulation of Secretion. *Am. Physiol. Society* 20(4):1524-1585.
- Gaines Das RE, Cotes PM (1979). International Reference Preparation Of Human Prolactin For Immunoassay: Definition Of The International Unit, Report Of A Collaborative Study And Comparison Of Estimates Of Human Prolactin Made In Various Laboratories. *J. Endocrinol.* 80:157-168.
- Godo G, Koloszar S, Daru J, Falkay G, Sas M (1988). Prolactin release during nursing in early puerperium. *Acta. Med. Hung.* 45(2):171-177.
- Iwamoto T, Yanase T, Koh E, Horie H, Baba K, Namiki M, Nawamata H (2004). Reference Ranges of Total Serum and Free Testosterone in Japanese Male Adults. *Nippon Hinyokika Gakkai Zasshi* 95(6):751-760.
- Jara LJ, Medina G, Saavedra MA, Vera-Lastra O, Navarro C (2011). Prolactin and autoimmunity. *Clin. Rev. Allergy Immunol.* 40(1):50-59.
- Jara LJ, Vera-Lastra O, Miranda JM, Alcalá M, Alvarez-Nemegyei J (2001). Prolactin in human systemic lupus erythematosus. *Lupus* 10:748-756.
- Jeffcoate SL, Bacon RRA, Beastall GH, Diver MJ, Franks S, Seth J (1986). Assays for Prolactin: Guidelines for the Provision of a Clinical Biochemistry Service. *Ann. Clin. Biochem.* 6:838-851.
- Katayev A, Balciza C, Secombe DW (2010). Establishing Reference Intervals for Clinical Laboratory Test Results is there a Better Way? *Am. J. Clin. Pathol.* 133:180-186.
- Kim C, Golden SH, Mather KJ, Laughlin GA, Kong S, Nan B, Barrett-Connor E, Randolph JF (2012). Racial/Ethnic Difference in Sex Hormone Levels among Postmenopausal Women in the Diabetes Prevention Program. *J. Clin. Endocrinol. Metab.* 97(11):4051-4060.
- Korode AGK (2008). Socio-economic Factors and Girl-child's Access to Secondary Education in Borno State. Unpublished M.Ed. Thesis Federal University of Technology, Yola, Nigeria.
- Kristiansson P, Wang JX (2001). Reproductive hormones and blood pressure during pregnancy. *Human Reproduction* 16(1):13-17.
- Kruger TH, Leeners B, Naegli E, Schmidlin S, Schedlowski M, Hartmann U, Egli M (2009). Prolactin secretory rhythm in women: immediate and long-term alterations after sexual contact. *Hum. Reprod.* 27(4):1139-1143.
- Manjer J, Johansson R, Berglund G, Janzon L, Kaaks R, Agren A, Lenner P (2003). Postmenopausal breast cancer risk in relation to sex steroid hormones, prolactin and SHBG (Sweden). *Cancer Causes Control* 14:599-607.
- Marano RJ, Ben-Jonathan N (2014). Minireview: Extrapituitary prolactin: an update on the distribution, regulation and functions. *Mol. Endocrinol.* 28(5):622-633.
- Meraiyebu A, Akintayo CO, Offiah NV (2012). A Study on Prolactin Hormone and Female Infertility in National Hospital Abuja, Nigeria. *IOSR J. Dental Med. Sci. (JDMS)* 2(2):38-41.
- Nagy E, Berci I (1989). Pituitary dependence of bone marrow function. *Br. J. Haematol.* 71:457-462.
- National Population Commission (NPC) (2009). [Nigeria]: Nigeria Demographic and Health Survey 2008. Abuja, Nigeria: National Population Commission and ICT Macro.
- Neidhart M, Gray RE, Gay S (1999). Prolactin and prolactin-like polypeptides in rheumatoid arthritis. *Biomed. Pharmacother.* 53:218-222.
- Okereke E, Aradeon S, Akerele A, Tanko M, Yisa I, Obonyo B (2013). Knowledge of safe motherhood among women in rural communities in northern, Nigeria: implications for maternal mortality reduction. *Reprod. Health* doi:10.1186/1742-4755-10-57
- Olootu WE, Adeleye AO, Amballi AA, Mosuro AO (2012). Pattern of Reproductive Hormones (Follicle Stimulating Hormone, Luteinizing Hormone, Estradiol, Progesterone, and Prolactin) Levels in Infertile Women in Sagamu South Western Nigeria. *Der Pharm. Lett.* 4(2):549-553.
- Onur E, Ercal T, Karslioglu I (1989). Prolactin and cortisol levels during spontaneous and oxytocin induced labour and the effect of meperidine. *Arch. Gynecol. Obst.* 244(4):227-232.
- Orbach H, Shoenfeld Y (2007). Hyperprolactinemia and autoimmune diseases. *Autoimmun. Rev.* 6:537-542.
- Ota E, Haruna M, Suzuki M, Anh DD, Tho LH, Tam NTT, Thiem VD, Anh NTH, Isozaki M, Shibuya K, Ariyoshi K, Murashima S, Moriuchig H, Yanaih H (2011). Maternal body mass index and gestational weight gain and their association with perinatal outcomes in Vietnam Bull. World Health Organ. 89:127-136.
- Pickering TG, Hall JE, Appel LJ, Falkner BE, Graves J, Hill MN, Jones DW, Kurtz T, Sheps SG, Roccella EJ (2005). Recommendations for Blood Pressure Measurement in Humans and Experimental Animals. Part 1: Blood Pressure Measurement in Humans: A Statement for Professionals from the Subcommittee of Professional and Public Education of the American Heart Association Council on High Blood Pressure Research. *Hypertension* 45:142.
- Reuwer AQ, Nowak-Sliwinska P, Mans LA, van der Loos CM, von der Thusen JH, Twickler MT, Spek CA, Gofin V, Griffioen AW, Borensztein KS (2012). Functional consequences of prolactin signaling in endothelial cells: a potential link with angiogenesis in pathophysiology? *J. Cell. Mol. Med.* 16(9):2038-2048.

- Riordan J (2005). Breastfeeding and human lactation, 3rd edn. Boston and London: Jones and Bartlett pp. 75-77.
- Rojas VS, Hollmann W, Struder HK (2012). Influences of exercise and training on the circulating concentration of prolactin in humans. *J. Neuroendocrinol.* 24(3):395-402.
- Sachidhanandam M, Singh SN, Ray US, Salhan AK (2009). Impact of prolonged high altitude exposure on plasma prolactin in men: effect of age and ethnicity. *High Altern. Med. Biol.* 10(4):343-348.
- Seaborg E (2011). The Hormone Foundation's Patient guide to Hyperprolactinemia Diagnosis and Treatment. *Endocrine News* 36(3):16-21.
- Speroff L, Fritz MA (2005). *Clinical Gynecologic Endocrinology and Infertility*. 7th edition, Wippincott Williams and Wilkins, Philadelphia. pp. 580-581.
- Suitor CW (1997). Maternal Weight Gain: A report of an Expert Work Group. Arlington VA. National Centre for Education in Maternal and Child Health.
- Tanner MJ, Hadlow NC, Wardrop R (2011). Variation of female prolactin levels with menopausal status and phase of menstrual cycle. *Austr. New Zealand J. Obstet. Gynaecol.* 51(4):321-324.
- Terra LF, Garay-Malpartida MH, Wailemann RA, Sogayar MC, Labriola L (2011). Recombinant human prolactin promotes human beta cell survival via inhibition of extrinsic and intrinsic apoptosis pathways. *Diabetologia* 54:1388-1397.
- Umar LKK (1996). Education for the Girl-child in Northern Nigeria: A Case for Counseling. *Canceller* 14(2):77-83.
- Van Cauter E (1990). Diurnal and ultradian rhythms in human endocrine function: a minireview. *Horm Res.* 34:45-53.
- Van Cauter E, Spiegel K (1997) Hormones and metabolism during sleep. In: Schwartz W, editor. *Sleep Science: Integrating Basic Research and Clinical Practice*. Basel: Karger pp. 144-174.
- Vemer HM, Rolland R (1981). The dynamics of prolactin secretion during the puerperium in women. *Clin. Endocrinol.* 15(2):155-163.
- Voogt JL, Lee Y, Yang S, Arbogast L (2001). Regulation of prolactin secretion during pregnancy and lactation. *Prog. Brain Res.* 133:173-185.
- Welniak LA, Richards SM, Murphy WJ (2001). Effects of prolactin on hematopoiesis. *Lupus* 10(10):700-705.
- Walker M (2006). *Breastfeeding Management for the Clinician: Using the evidence*. Boston: Jones and Bartlett. pp. 63-66.
- Wolf M, Abubakar A, Tsui S, Williamson NE (2008). Child spacing attitudes in northern Nigeria. VI: FHI: Arlington.
- Zeitlin SI, Raifer J (2014). Hyperprolactinemia and Erectile dysfunction. *Rev. Urol.* 2(1):39-42.