

Full Length Research Paper

Morphological diversity and association of traits in ethiopian food barley (*Hordeum vulgare* L.) landraces in relation to regions of origin and altitudes

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One hundred and two barley accessions and five checks were evaluated using augmented design consisting of four complete blocks in 2012 main cropping season at holetta agricultural research center. Ten quantitative and six qualitative characters were recorded. Analysis of variance showed significant difference ($p < 0.01$) among accessions for plant height, awn length, peduncle extrusion, thousand seed weight, number of seeds per spike, days to 50% flowering and days to maturity. Cluster analysis grouped accessions in to five distinct classes with maximum number of accessions (44) in cluster I and minimum (2) in cluster V. Principal component analysis showed that variances of 30, 17, 15 and 10% were extracted from the first four principal components, respectively, which contributed 72% of the total variation among accessions. Estimates of genetic diversity index based on qualitative characters showed high diversity index among characters at Arsi, Wellega and Wello, and diversity index increased in altitude between 2001 and 3000 m.a.s.l and decrease at altitude >3000 . Phenotypic diversity was very high for kernel row number ($H' = 0.99$), grain color ($H' = 0.90$) and spike attitude ($H' = 0.85$) and low for lemma color ($H' = 0.48$). Days to flowering, days to maturity and numbers of seed per spike, from quantitative characters and kernel row number, grain color and spike attitude from qualitative characters were the most characters which contributed variances among accessions.

Key words: Cluster analysis, diversity index, principal component analysis, qualitative characters, quantitative characters.

INTRODUCTION

Barley (*Hordeum vulgare* L.) is an annual cereal crop which belongs to the genus *Hordeum* in the Tribe Triticeae of grass family Poaceae which contains about 350 wild species (Amanda, 2008). It is thought that barley

has to be originated in the Fertile Crescent area of the Near East from the wild progenitor *Hordeum spontaneum* over 10,000 years ago (Badr et al., 2000; Blattner and Badani, 2001; Grando and Helena, 2005; Azhaguvel and

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Komatsuda, 2007; Dai et al., 2012). Barley is a progenitor *Hordeum spontaneum* over 10,000 years ago (Badr et al., 2000; Blattner and Badani, 2001; Grando and Helena, 2005; Azhaguvel and Komatsuda, 2007; Dai et al., 2012). Barley is a major crop, grown worldwide and in a wide range of climatic conditions; despite its importance as a crop species, little is known about the population genetics of barley and the effects of bottlenecks, adaptation, and gene flow on genetic diversity within and between landrace populations (Leino and Jenny, 2010; Tanto et al., 2010). The crop successfully grows in the arid climates of the Sahara, the Tibetan plateaus, the highlands of the Himalayas, and the Andean countries, the tropical plains of India and the mountains of Ethiopia (Grando and Helena, 2005).

Ethiopia is an important primary and secondary gene center for many field crop species, including barley, which were introduced centuries ago and since then adapted and developed wide genetic diversity (Abdi, 2011). Landraces represent over 90% of the barley cultivated in Ethiopia (Tanto et al., 2010). In Ethiopia barley is the fifth most important cereal crop both in area coverage and production, with around 1,013,623.72 ha and 18,155,830.29 qt respectively (CSA, 2012). It is grown both in *Mehar* (June-September) and *Belg* (March-April) seasons. The diversity in soils, climate, altitude and topography, together with geographical isolation for long periods, are considered the main factors influencing the large diversity in Ethiopian barley (Harlan, 1976); social factors also play an important part in the diversification, thus, the morphological, biochemical and molecular groups in Ethiopian barley are the result of accumulated long-term mutations, hybridization, gene recombination and natural and human selection in heterogeneous environments (Lakew and Alemayehu, 2011).

In our country Ethiopia to conserve plant genetic resources including barley, the Plant Genetic Resources Centre of Ethiopia (PGRC/E), now the Institute of Biodiversity Conservation (IBC) was established in 1976. The primary mandates of IBC include the preservation of genetic diversity of crop plants, their wild relatives, and native species important to Ethiopian agriculture and biodiversity. Over 65 000 accessions from more than 120 plant species have been collected across the country and preserved *ex situ* at IBC. This germplasm collection includes a principal base collection of barley with >15,000 accessions (Abdi, 2011). However, most of collected and preserved landraces at the Gene Centre are not yet studied for their morphological diversity (Alemayehu and Parlevliet, 1997; Abdi, 2011). Therefore, this study is proposed with the following objectives:

1. To assess the extent of morphological variation in barley accessions in respect to regions of origin and altitudes of collection.
2. To cluster the accessions into relatively homogenous groups and to identify the major characters contributing to

the overall diversity of the germplasm.

MATERIALS AND METHODS

Experimental materials

A total of 102 barley accessions were obtained from the Institute of Biodiversity Conservation, Addis Ababa, Ethiopia. The accessions were selected based on their region of origin and altitude (Table 1). Five standard checks (controls) (HB-42, Ardu, Shege, HB1307 and Balami), that were obtained from the Holetta Agricultural Research Center were included (a total of 107 genotypes were used in this study). The region of collection and altitude range is given in Table 1. Five gram of seed (100 kg ha⁻¹) was obtained from IBC for each accession.

Experimental site

The experiment was conducted at the Holetta Agricultural Research Center, Ethiopia, during the main cropping season of 2012 under rain fed condition. Holetta Agricultural Research Center is located at 9° 3'N, 38° 30'E with an altitude of 2400 m.a.s.l. It is 28 km west of Addis Ababa on Ambo road of and characterized with annual rainfall of 1044 mm, mean relative humidity of 60.6% and mean maximum and minimum temperature of 22.1 and 6.2°C, respectively (Figure 1).

Experimental procedures

The experiment was laid out in augmented randomized complete block design (Federer and Ragavarao, 1975) consisting of four blocks in which the 102 accessions were planted in un-replicated plots and the five checks were replicated four times (ones in each block) to estimate an error variance. The plot size used was one row with 2.5 m length, and 0.4 m between rows. Seeds were planted by hand with a seeding rate of 100 kg/ha. Plots were kept free from weeds.

Data collection

Based on the IPGRI descriptor list (IPGRI, 1994); ten quantitative and six qualitative characters were recorded (Tables 2 and 3). For each accession, 10 randomly selected individual plants were used for recording quantitative characters, except days to 50% flowering, days to maturity and thousand seed weight, which were recorded on plot basis.

ANALYSIS OF VARIANCE

Quantitative traits

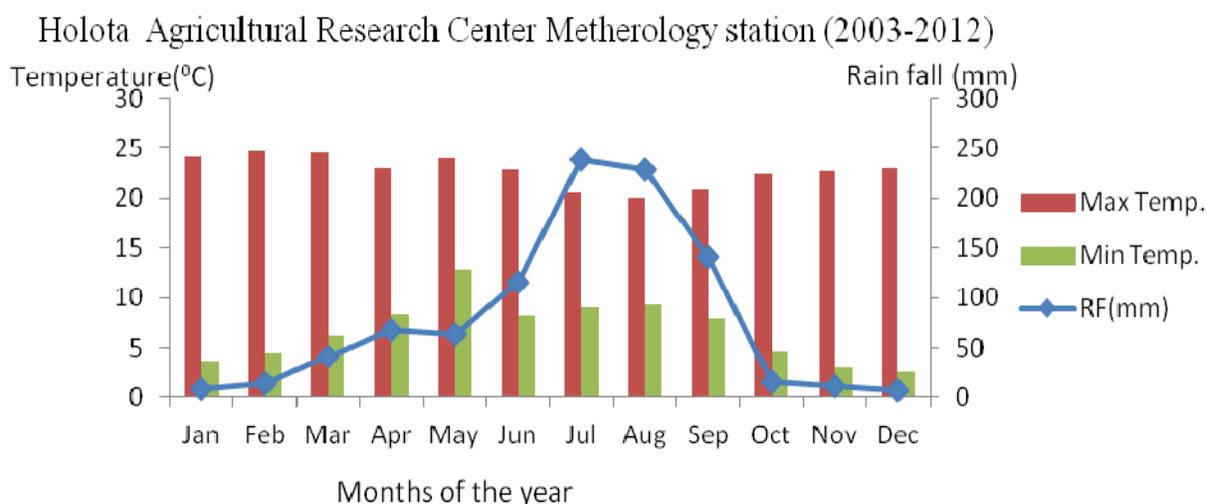
All quantitative data were analyzed using SAS v 9.1.3 Software (SAS, 2004). A mixed model in which standard checks effect were considered as fixed, and accessions effect as random effect, was adopted as:

$$Y_{ij} = \mu + \alpha_i + \beta_j + e_{ij}$$

Where: Y_{ij} is response variable; μ is general mean, α_i is the fixed effect of i^{th} standard checks and random effect of accessions, β_j is the random effect of j^{th} block and e_{ij} is random errors. Mean squares

Table 1. Region of origin, altitude, and number of accessions used for this study

Region	Number of accessions by altitude groups(m.a.s.l)				Total number of accessions
	Group I (1500-2000)	Group II (2001-2500)	Group III (2501-3000)	Group IV (3001-3500)	
Arsi	3	3	3	3	12
Bale	3	3	2	3	11
Gojam	2	4	3	2	11
Gonder	2	0	7	2	11
Shewa	3	4	3	2	12
Sidamo	3	3	4	1	11
Tigray	3	4	4	1	12
Wellega	3	4	4	0	11
Welo	3	2	3	3	11
Total	25	27	33	17	102

**Figure 1.** Climatic data of the experimental sites at Holetta agricultural research center minimum and maximum temperature and monthly rainfall.

were calculated as shown in Table 4. Estimates of σ_e^2 , σ_g^2 and σ_b^2 were obtained by equating the obtained sum of squares to their expectancies, and solving the resulting system equations:

$$\sigma_g^2 = \frac{\text{GenotypesMS} - \text{ErrorMS}}{\text{Blocks}}$$

$$\sigma_c^2 = \frac{\text{ControlsMS} - \text{ErrorMS}}{\text{Blocks}}$$

$$\sigma_t^2 = \frac{\text{test(Accessions)MS} - \text{ErrorMS}}{\text{Block}}$$

Where genotypes = accessions + checks (controls).

Cluster analysis

Before undertaking multivariate analysis of variance in which two or more variables were analyzed at a time, the data was standardized to mean of zero(0) and a variance of one(1) to avoid differences in scales. One hundred two accessions and nine regions of origin were grouped into respective classes. The values of pseudo F statistic (PSF) and Hotellin's pseudo T² statistic were used for defining optimum number of clusters. Cluster analysis was made using the hierarchical cluster analysis. The PROC CLUSTER Procedure of SAS V9.1.3 (SAS, 2004) using Unweighted Pair Group Method using Arithmetic Average linkage (UWPGMA) was employed.

Principal component analysis

The principal component analysis (PCA) was computed to reduce the number of variables into a few correlated components that can explain much of the variability. It was performed using the

Table 2. List of quantitative characters recorded along with their code and definition.

Characters	Code	Character definition
Awn length (cm)	AWL	Distance from the tip of the spike to the end of the awn
Days to 50% flowering (count)	DFL	Number of days from planting to the day when 50% of the heads fully flower (heading) emerge from the boot of flag leaf in each row
Days to maturity (count)	DMA	Number of days starting from planting to the days when peduncles of the spikes in each row become complete yellow and mature
Number of fertile tillers per plant (count)	NFTPP	Number of fertile tillers (spike bearing) of randomly selected plants per plant, counted at maturity
Number of seeds per spike (count)	NSPS	Number of seed per spike on randomly selected plants counted at maturity
Peduncle extrusion length (cm)	PEDext	Distance from the auricle of flag leaf to the base of spike
Peduncle length (cm)	PDL	Distance from last node to base of the spike
Plant height (cm)	PLH	Length of randomly selected plants measured from the ground to the tip of the spike excluding awns at maturity
Spike length (cm)	SPL	length measured from base of spike to top of spikelets excluding the awns at maturity
Thousand seed weight (g)	TSW	The weight of 1000 seeds taken from each row in gram

Table 3. List of qualitative characters recorded along with their codes and descriptions.

Characters	Code	Character descriptions
Awn color	ACO	White (1), Brown (3), Black (5), Reddish (4)
Grain color	KCO	White (1), Red(2), Black(4), Purple (3)
Kernel row number	KRN	Two row (1), Six row(5), Irregular (3)
Lemma color	LMC	White (1), Black(4), Red(2), Purple (3)
Spike attitude	SPA	Erect (1), Horizontal(5), Semi-recurved (7)
Spike density	SPD	Lax (3), intermediate (5), dense (7)

correlation matrix to define the patterns of variation among landraces based on the mean of quantitative characters. And also helps to identify characters that load the most in explaining the observed variation. The PROC PRINCOMP Procedure of SAS V9.1.3 (SAS, 2004) was used for principal component analysis.

Qualitative traits

Estimate of diversity index

The Shannon-Weaver diversity index (H') was used to compute the phenotypic frequencies to assess the phenotypic diversity for each character for all accessions, based on qualitative traits. It is used in genetic resource studies as a convenient measure of both richness and evenness using phenotypic data:

$$H = -\sum_{i=1}^n p_i \ln(p_i)$$

$$H' = H/H_{max}$$

$$H_{max} = \ln(n).$$

Where: H' = standardized relative diversity index; n = is the number of phenotypic classes per characters; P_i = is the proportion of the total number of entries in the i^{th} class; \ln = natural logarithm.

RESULTS AND DISCUSSION

Analysis of variance

Analysis of variance indicated significant difference ($p < 0.01$) among genotypes, accessions, controls and accessions vs. controls for all quantitative characters except awn length in controls, peduncle length in accessions vs. controls, spike length in genotypes, accessions and accessions vs. controls, number of fertile tiller per plant in controls and accessions vs. controls, and days to maturity in controls (Table 5). Hence, the result indicated the existence of high morphological variation in Ethiopian food barley landraces, in their regions of origin

Table 4. ANOVA table for sum of squares and their expectancies for the statistical genotypic model (Federer and Ragavarao, 1975)

Source of variation	Degree freedom	Mean square	Expected mean square
Blocks(b)	b-1	MSb	-
Genotypes(g)	g-1	MSg	$\sigma_e^2 + \sigma_g^2$
tests (accessions) (t)	t-1	MSt	$\sigma_e^2 + \sigma_t^2$
controls (c)	c-1	MSc	$\sigma_e^2 + \sigma_c^2$
tests vs. controls (t vs.c)	1	MSt vs.c	$\sigma_e^2 + \sigma_t^2 + \sigma_c^2$
Error	(b-1)(c-1)	MSe	σ_e^2
Total	n-1	-	$\sigma_e^2 + \sigma_g^2 + \sigma_b^2$

MSg = mean square of genotypes, MSb = mean square of blocks, MSt = mean square of test (accessions), MSc = mean square of controls, MSt vs.c = mean square of tests vs. controls, MSe = mean square of error, σ_e^2 =expected error variance (MSe), σ_g^2 = Genotypic variance component, σ_t^2 = accessions variance component.

Table 5. Analysis of variance for ten quantitative characters.

Source of variation	DF	Mean square									
		PLH	AWL	PDER	PDL	SPL	TSW	NSPS	NFTPP	DFL	DMA
Block	3	470.87	0.16	4.10	43.48	1.57	70.17	79.13	0.53	50.61	19.51
MSg	106	107.45**	0.86**	5.83**	46.31*	1.05 ^{ns}	28.09**	185.81**	0.39*	86.20**	130.72**
MSt	101	98.66**	0.87**	5.47**	45.87*	0.88 ^{ns}	26.60**	162.52**	0.40*	69.52**	101.43**
MSc	4	302.99**	0.56 ^{ns}	11.54**	58.81*	5.29**	63.63**	555.81**	0.25 ^{ns}	14.84*	1.12 ^{ns}
MSt _{vs.c}	1	214.27**	1.09*	19.37**	42.07 ^{ns}	1.44 ^{ns}	36.79**	1056.57**	0.59 ^{ns}	2047.01**	2240.60**
MSE	12	19.24	0.22	1.24	18.41	0.62	1.52	15.39	0.16	3.32	1.60
Cort'd total	121	13033.45	95.24	645.75	5260.68	124.13	3206.77	20118.48	45.83	9329.60	13934.57
Mean		104.78	11.45	13.89	37.69	8.09	45.23	37.63	3.71	78.82	125.80
SE		4.70	0.51	1.19	4.60	0.93	1.32	4.20	0.44	1.95	1.35
CV (%)		4.18	4.17	8.02	11.38	9.79	2.73	10.42	11.04	2.31	1.01

*, **, ns indicates significance at P=0.05 level, P=0.01 and non-significant, respectively. MSg = mean square of genotypes (controls + accessions), MSt = mean square accessions, MSc = mean square of control, MSt_{vs.c} = mean square of accessions vs. control MSE = mean square of error (error variance), Cort'd total = corrected total, SE = standard error, CV(%) = coefficient of variation, PLH = plant height, AWL = awn length, PDER=peduncle extrusion, PDL = peduncle length, SPL = spike length, TSW = thousand seed weight, NSPS = number of seed per spike, NFTPP = number of fertile tiller per plant, DFL = days to 50% flowering, DMA = days to maturity.

and altitude groups. The same results were reported on morphological diversity of Ethiopia barley landraces by different authors (Tanto et al., 2009; Abay et al., 2009; Eticha et al., 2010; Dejene et al., 2010; Tanto et al., 2010; Muhe and Alemayehu, 2011; Jalata et al., 2011).

Morphological variation within regions

Estimate of genotypic variance for regions of origin among accessions showed highly significant difference ($p < 0.01$) for plant height, peduncle extrusion, spike length, thousand seed weight, number of seeds per spike, days to 50% flowering and days to maturity in all regions. Similarly, awn length from Arsi, Sidamo, and Wellega; peduncle length from Bale, Shewa, Sidamo, Wellega and Wello; number of fertile tillering per plant from Arsi and Tigray significantly varied (Table 6). Analyses of diversity pattern, among accessions from different regions for

quantitative characters revealed existence of morphological diversity within regions indicating differences in agro-ecological conditions across regions contributing for the observed morphological diversity. Similar results were also reported in several studies (Negassa, 1985; Demissie and Bjornstad, 1996; Dejene et al., 2010).

Morphological variation within altitudinal gradients

Most of the morphological characters showed significant variation among altitude groups except peduncle length in altitude group I (1500-2000) and IV (3001-3500), spike length in altitude group I and number of fertile tiller per plant in altitude group II (2001-2500) and IV (Table 6). The altitude group III (2501-3000) showed significant genotypic variation for all characters measured. In general, high genotypic variation was observed in an

Table 6. Estimate of genotypic variances for nine regions of origin and four altitude groups based on ten quantitative characters

Region	PLH	AWL	PDER	PDL	SPL	TSW	NSPS	NFTPP	DFL	DMA
Arsi	19.06**	0.09*	0.87**	4.58 ^{ns}	0.27*	6.77**	54.59**	0.13**	19.18**	33.52**
Bale	24.51**	0.02 ^{ns}	1.22**	8.33*	0.26*	6.66**	58.59**	0.01 ^{ns}	32.20**	43.46**
Gojam	35.38**	0.02 ^{ns}	1.31**	5.51 ^{ns}	0.24*	6.36**	63.94**	0.03 ^{ns}	34.63**	28.93**
Gonder	40.41**	0.02 ^{ns}	0.88**	1.74 ^{ns}	0.34*	9.64**	45.06**	0.01 ^{ns}	11.23**	25.56**
Shewa	33.41**	0.07 ^{ns}	1.34**	9.96*	0.47**	8.11**	63.52**	0.03 ^{ns}	15.39**	36.98**
Sidamo	25.89**	0.31**	1.30**	8.79*	0.26*	11.46**	58.05**	0.00	30.05**	54.18**
Tigray	53.59**	0.04 ^{ns}	0.81*	5.84 ^{ns}	0.25*	7.66**	91.18**	0.12**	35.47**	44.66**
Wellega	29.10**	0.54**	4.02**	9.32*	0.46**	9.29**	75.45**	0.06 ^{ns}	28.03**	36.29**
Wello	29.10**	0.05 ^{ns}	1.24**	10.44*	0.55**	7.77**	50.95**	0.00	18.85**	39.11**
Altitude group (m.a.s.l)										
Group I	31.18**	0.30**	1.50**	4.10 ^{ns}	0.13 ^{ns}	5.37**	54.89**	0.07*	33.32**	46.53**
Group II	19.63**	0.08*	1.56**	6.85*	0.27*	8.83**	59.28**	0.05 ^{ns}	31.08**	40.49**
Group III	26.52**	0.08*	1.17**	7.48**	0.35*	5.96**	48.93**	0.09*	15.69**	35.39**
Group IV	31.67**	0.10*	0.78**	5.73 ^{ns}	0.28*	11.31**	37.72**	0.03 ^{ns}	10.88**	22.58**

*, **, and ns indicates significance at $P = 0.05$, $P = 0.01$ and non-significant, respectively. PLH = plant height, AWL = awn length, PDER = peduncle extrusion, PDL = peduncle length, SPL = spike length, TSW = thousand seed weight, NSPS = number of seed per spike, NFTPP = number of fertile tiller per plant, DFL = days to 50% flowering, DMA = days to maturity; M.a.s.l. = meter above sea level; Group I (1500-2000), Group II (2001-2500), Group III (2501-3000) and Group IV (3001-3500).

altitude groups II and III, which comprised the major barley growing areas in the country. Similar result was reported by Demissie and Bjornstad (1996) and Dejene et al. (2010) where they found high variation concentration in areas between 2000-3000 and 2400-3000 m.a.s.l. respectively. This high variation attributed to mixed farming system, which is typically found in areas of higher elevation usually above 2000 m.a.s.l. Tanto et al. (2009) also reported the reduction of area of cultivation for barley as altitude decreased which indicated that barley is cool climate crop.

Cluster analysis

Cluster analysis for accessions

Cluster analysis grouped the 102 accessions in to five distinct groups (Table 7). Numbers of accessions per cluster varied from 44 accessions in cluster I to 4 accessions in cluster V. Cluster means and percent of populations (accessions) in each cluster are presented in Tables 7 and 8. Forty four accessions were found in cluster I, which was 43.1% of the total experimental materials. This cluster has been characterized by intermediate plant height, relatively the heaviest thousand seed weight, relatively higher number of fertile tillers per plant, early flowering and early maturity. Accessions grouped under cluster I were scattered along all regions and more at altitude group I (1500-2000) and II(2501-3000). Cluster II accounts 22.6% of the population and included 23 accessions and had shorter peduncle extrusion, longer days to 50% flowering and longer days

to maturity. Majority of these accessions were collected at altitude group III (2501-3000) from all regions except Shewa and Tigray. Relatively accessions with shorter plant height, earlier days to 50% flowering, earlier maturity, and smaller thousand seed weight were grouped under cluster III which contribute 17.7% to the population (18 accessions).

Cluster IV consisted of thirteen accessions, 12.8% of the population, characterized by high number of seeds per spike and moderate in days to 50% flowering and days to maturity; which includes more accessions collected from Shewa and from all altitude groups. This cluster, cluster IV, contains accessions which have high number of seeds per spike and early mature, especially accession number 4879, 243571, 235068 and 242093. Cluster V included four accessions (3.9% of the population) and characterized by taller plant height, longer awn length, peduncle extrusion, peduncle length, spike length, and heavier thousand seed weight, fewer number of seeds per spike, lower number of fertile tillers per plant, relatively late days to 50% flowering and days to maturity, in which accessions were collected from Arsi, Bale and Wellega from altitude groups II (2001-2500), III (2501-3000) and IV (3001-3500).

Although the cluster analysis grouped barley accessions with greater morphological similarity, the cluster did not necessarily included all accessions from the same or adjacent sites. This result is in agreement with the work of Dejene et al. (2010) who reported that, clustering of accessions based on the agronomic characters revealed no distinct regional grouping patterns in which accessions from same or adjacent regions appeared in different clusters.

Table 7. Distribution of 102 barley accessions over five clusters by nine regions of origin and four altitude groups based on 10 quantitative characters

Regions	Clusters					No. of accessions
	I	II	III	IV	V	
Arsi	4	4	3	-	1	12
Bale	5	3	2	-	1	11
Gojam	5	6	-	-	-	11
Gonder	2	7	2	-	-	11
Shewa	2	-	4	6	-	12
Sidamo	6	1	3	1	-	11
Tigray	9	-	-	3	-	12
Wellega	7	1	1	-	2	11
Wello	4	1	3	3	-	11
Total	44	23	18	13	4	102
Altitude groups						
Group I	20	4	-	1	-	25
Group II	13	1	8	4	1	27
Group III	9	11	7	5	1	33
Group IV	2	7	3	3	2	17
Total	44	23	18	13	4	102

Table 8. The summary of cluster mean of 102 barley accessions for 10 quantitative characters

Characters	Cluster means				
	I	II	III	IV	V
Plant height	99.8	108.4	100.8	113.1	115.5
Awn length	11.5	11.0	11.8	11.8	11.7
Peduncle extrusion	14.9	12.5	13.1	14.4	17.1
Peduncle length	40.0	32.7	37.9	37.6	46.7
Spike length	8.2	8.3	7.6	7.5	8.6
Thousand seed weight	47.4	45.6	39.1	42.2	50.3
Number of seed per spike	25.1	40.7	44.6	58.5	25.5
Number of fertile tiller per plant	4.1	3.7	3.4	3.4	3.3
Days to 50% flowering	70.3	88.9	75.9	78.5	83.0
Days to maturity	116.0	138.2	117.6	125.1	135.5
Number of accessions	44	23	18	13	4

Clustering indicated that environment had an impact on the performance of barley and specifically altitude had great contribution for the variability of the characters.

Cluster analysis for regions

Regional cluster analysis grouped the nine regions of barley accessions in to four groups based on 10 quantitative characters (Table 9). Arsi, Bale, Gojam and Wellega grouped in to cluster I characterized with the longest spike length and earlier flowering. Cluster II was characterized with the longest plant height, awn length,

peduncle extrusion, peduncle length, and number of seeds per spike, in which Shewa, Wello and Sidamo were grouped in this cluster. The shortest plant height, awn length, peduncle length, spike length, the heaviest thousand seed weight, the lowest number of seeds per spike and the highest Number of fertile tillers per plant were clustered under cluster III, in which Tigray is the source of collection for this cluster. Cluster IV comprised one region (Gonder) which was characterized with shorter peduncle extrusion; longer spike length, smaller number of fertile tiller per plant, delayed flowering and maturity. The same results were reported by Dejene et al. (2010) and Demissie and Bjornstad (1996).

Table 9. The summary of cluster means of nine regions of barley accessions for their 10 quantitative characters.

Characters	Cluster means			
	I	II	III	IV
Plant height	105.11	106.53	96.00	102.06
Awn length	11.35	11.91	11.08	11.25
Peduncle extrusion	14.77	14.52	12.36	11.74
Peduncle length	39.01	39.26	34.15	34.28
Spike length	8.24	7.97	7.77	7.82
Thousand seed weight	46.62	43.06	46.96	42.30
Number of seed per spike	31.84	42.52	31.37	40.16
Number of fertile tiller per plant	3.81	3.55	4.09	3.41
Day to 50% flowering	59.45	76.14	72.90	84.12
Day to maturity	124.83	119.97	119.76	130.34
Regions	Arsi, Bale, Gojam Wellega	Shewa, Wello Sidamo	Tigray	Gonder
Number of regions	4	3	1	1

Principal component analysis

Principal component analysis for accessions

The principal component analysis exhibited variances of 30, 17, 15 and 10%, were extracted for the first four principal components and accounts about 72% of total variation (Table 10). Days to 50% flowering, days to maturity, number of seeds per spike and peduncle extrusion showed greater loading for the variation in the first principal components. Similarly, thousand seed weight, days to maturity, spike length and number of seeds per spike contributed major variation in the second principal component. The variation in the third principal component were mainly due to number of fertile tiller per plant, peduncle extrusion, plant height, awn length and peduncle length, while the fourth principal component showed 10% of total variation with greater loading from awn length, plant height and spike length. In line with the present finding, Demissie and Bjornstad (1996) employed principal component analysis for detecting variation in 49 barley populations in which the first four PCs contributed 63% of total variation. Generally days to 50% flowering, days to maturity, and number of seeds per spike were the most loading characters for the variation among accessions.

Principal component analysis for regions

Principal component analysis showed that 83% of total variation among regions was extracted for the first three principal components having eigenvalue greater than one (Table 10). Peduncle extrusion, peduncle length, days to flowering, days to maturity and plant height gave the most

loading contribution for the variation in first principal component which contributed 34% of the variation. The second principal component contributed 31% of the variation in which thousand seed weight, number of seed per spike, awn length and number of fertile tillers per plant contributed greater variation. Similarly, days to maturity, days to flowering, spike length and plant height were the most loading contributors for the third principal component.

Principal component analysis for altitude groups

The first two principal components extracted 93% of total variation among altitude groups having eigenvalue greater than one (Table 10). Number of seed per spike, peduncle extrusion, number of fertile tiller per plant and days to 50% flowering were the most loading contributors in the first principal component. Similarly, spike length, awn length and peduncle length were showed greater loading in the second principal component.

Diversity index

Estimates of Shannon Weaver diversity index over regions of origin and altitude groups showed high diversity index for the six qualitative characters studied. Phenotypic diversity was very high for kernel row number ($H'=0.99$), grain color ($H'=0.90$) and spike attitude ($H'=0.85$) and low for lemma color ($H'=0.48$) (Table 11). This is due to high ecological heterogeneity of the country, which is favorable condition for barley landrace cultivation. Except lemma color, all characters were high in phenotypic diversity over all regions of origin and

Table 10. Eigenvectors, total variance, cumulative variance, and eigenvalues for ten quantitative characters of 102 barley landrace in Ethiopia for first four, three and two principal components for accessions, regions and altitude groups respectively

Characters	Eigen vectors for accessions				Eigen vectors for regions			Eigen vectors for altitude groups	
	PC1	PC2	PC3	PC4	PC1	PC2	PC3	PC1	PC2
PLH	-0.18	0.16	0.39	-0.51	0.30	0.25	0.32	0.32	0.23
AWL	0.14	-0.23	0.38	0.62	0.29	0.43	-0.01	0.24	-0.54
PDER	0.36	0.01	0.42	-0.11	0.49	0.05	0.10	-0.34	-0.24
PDL	0.29	-0.24	0.35	-0.28	0.42	0.09	0.21	-0.32	-0.27
SPL	0.21	0.35	0.31	0.42	0.27	-0.23	0.45	-0.22	0.56
TSW	0.24	0.57	0.12	-0.08	0.22	-0.51	0.01	-0.31	0.26
NSPS	-0.40	-0.34	0.19	0.16	-0.20	0.47	-0.02	0.35	-0.16
NFTPP	0.25	0.25	-0.45	0.17	0.08	-0.38	-0.23	-0.33	-0.03
DFL	-0.47	0.27	0.14	0.18	-0.35	0.06	0.52	0.33	0.24
DMA	-0.41	0.37	0.18	0.04	-0.31	-0.19	0.54	0.32	0.18
Eigen value	2.96	1.70	1.52	1.02	3.4	3.08	1.8	7.47	1.86
% of total variance	30	17	15	10	34.0	31.0	18.0	75.0	19.0
% cumulative variance	30	47	62	72	34.0	65.0	83.0	75.0	93.0

PLH = plant height, AWL = awn length, PDER = peduncle extrusion, PDL = peduncle length, SPL = spike length, TSW = thousand seed weight, NSPS = number of seed per spike, NFTPP = number of fertile tiller per plant, DFL50% = days to 50% flowering, DMA = days to maturity, PC = principal component.

altitude groups for this study. The same results were reported by different authors (Demissie and Bjornstad, 1996; Abdi, 2011; Lakew and Alemayehu, 2011).

Regional diversity index

Estimate of diversity index (H') pooled over regions of origin showed high phenotypic diversity among six qualitative characters (Table 11). The mean H' varied from 0.66 for Tigray to 0.83 for Arsi. Arsi, Wellega and Wello showed greater diversity index followed by Bale, Gojam, Gonder and Sidamo. Tigray and Shewa showed lower genetic diversity index. Among all characters, kernel row number from Gonder, grain color from Gojam, Shewa, and Wellega, spike density from Arsi and Tigray showed high genetic diversity index. Lemma color and awn color showed lower genotypic diversity index in most regions.

Altitudinal diversity index

Altitude groups showed high phenotypic diversity among six qualitative characters. The mean H' pooled over characters for four altitude groups varied from 0.65 for altitude group I (1500-2000 m.a.s.l) to 0.84 for altitudes group III (2501-3000 m.a.s.l) with mean value of 0.77 ± 0.07 (Table 11). Kernel row number from altitude group II (2001-2500), and III, grain color from altitude group I and III and spike density from altitude group III and IV (3001-3500) showed the highest diversity index. Altitude group II indicated lower genetic diversity index for lemma color.

Difference in altitude gradient and agro-ecological setting gave high diversity variation in cereal crops especially barley landraces. The result indicated high H' in Ethiopia barley landrace in altitude group III (2501-3000 m.a.s.l). Diversity index decreased at an altitude above 3000 m.a.s.l. This result is in agreement with the work of Demissie and Bjornstad (1996) and Abdi (2011). Similarly, mean diversity index for characters increases with altitude reaching a maximum between 2400 and 2800 m.a.s.l and decreasing beyond that altitude (Engels, 1991). This indicates high genotypic diversity in barley is related to high rainfall and lower temperature at high altitudes, which shows barley that is a cool season crop.

Conclusions

One hundred and two barley accessions were evaluated for ten quantitative and six qualitative characters to assess morphological diversity and association of traits in Ethiopian food barley (*H. vulgare* L.) landraces in relation to regions of origin and altitudes. Analysis of variance and genetic diversity index indicated the existence of morphological diversity and association of traits in Ethiopian food barley (*H. vulgare* L.) landraces in relation to regions of origin and altitudes. Cluster analysis grouped one hundred two accessions in to five distinct groups. Number of accessions per cluster varied from 44 accessions in cluster I to 4 accessions in cluster V. Shannon-Weaver diversity index showed high and comparable levels of phenotypic diversity among the accessions. Phenotypic diversity was very high for kernel row number ($H'=0.99$), grain color ($H'=0.90$) and spike

Table 11. Estimate of Shannon-Weaver diversity index (H') of 102 Ethiopia barley landraces for nine region of origins and four altitude groups by six qualitative characters.

Region	Qualitative characters						Mean $H' \pm SE$
	SPA	KRN	ANC	LMC	GRC	SPD	
Arsi	0.84	0.82	0.95	0.57	0.85	0.98	0.83 \pm 0.05
Bale	0.93	0.85	0.47	0.42	0.74	0.85	0.71 \pm 0.08
Gojam	0.90	0.90	0.62	0.46	0.96	0.82	0.77 \pm 0.07
Gonder	0.71	0.99	0.57	0.39	0.86	0.81	0.72 \pm 0.08
Shewa	0.58	0.92	0.44	0.42	0.96	0.87	0.69 \pm 0.10
Sidamo	0.87	0.91	0.52	0.38	0.90	0.86	0.74 \pm 0.09
Tigray	0.69	0.76	0.43	0.34	0.75	0.99	0.66 \pm 0.09
Wellega	0.90	0.69	0.91	0.61	0.96	0.89	0.82 \pm 0.05
Wello	0.73	0.94	0.93	0.58	0.87	0.92	0.82 \pm 0.05
Altitude group (m.a.s.l)							
Group I	0.57	0.69	0.52	0.53	0.97	0.66	0.65 \pm 0.06
Group II	0.90	0.95	0.61	0.47	0.93	0.82	0.78 \pm 0.08
Group III	0.70	0.98	0.84	0.59	0.97	0.96	0.84 \pm 0.06
Group IV	0.93	0.89	0.49	0.28	0.49	0.93	0.66 \pm 0.11
Total Mean	0.85	0.99	0.70	0.48	0.90	0.72	0.77 \pm 0.07

SPA = Spike attitude; KRN = Kernel row number; ANC = Awn color; LMC = Lemma color; GRC = Grain color; SPD = Spike density, m.a.s.l = meter above sea level; Group I (1500-2000), Group II (2001-2500), Group III (2501-3000) and Group IV (3001-3500).

attitude ($H'=0.85$) and low for lemma color ($H'=0.48$). The mean H' pooled over characters for four altitude groups, were varied from $H'=0.65$ for altitude group I (1500-2000) to $H'=0.84$ for altitude group III (2501-3000). Greater genotypic diversity index was observed in Arsi, Wellega and Wello and also high genotypic diversity was observed in altitude groups II (2001-2500), and III (2501-3000), which comprised the major barley growing areas in the country. Days to flowering, days to maturity and numbers of seeds per spike, from quantitative characters and kernel row number, grain color and spike attitude from qualitative characters contributed much of the variances among accessions. Based on the observed variation both for quantitative and qualitative characters, it could be concluded that studying the phenotypic diversity among barley accessions is important to identify the genetic potential of parental lines and increase the efficiency of the barley breeding programmers.

Conflict of Interest

The authors have not declared any conflict of interest.

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REFERENCES

- Abay F, Bjornstad A, Melinda S (2009). Measuring on farm diversity and determinants of barley diversity in Tigray, Northern Ethiopia. *Momona Ethiop. J. Sci.* 1(2):44-66.
- Abdi A (2011). Barley genetic resources collection and conservation in Ethiopia. Mulatu B. and Grando S. (Eds.), *Barley research and development in Ethiopia. Proceedings of the 2nd National Barley Research and Development Review Workshop.* November 28-30, 2006, HARC, Holetta, Ethiopia. pp 19-30.
- Alemayehu F, Parlevliet JE (1997). Variation between and within Ethiopian barley landraces. *Euphytica* 94:183-189.
- Amanda B (2008). *The biology of Hordeum vulgare L. (barley).* 2nd Edition, The University of Adelaide. Australia, 404 pp.
- Azhaguvel P, Komatsuda T (2007). A phylogenetic analysis based on nucleotide sequence of a marker linked to the brittle rachis locus indicates a diphyletic origin of barley. *Ann. Bot.* 100:1009-1015.
- Badr A, Muller K, Schafer-Pregl R, El Rabey H, Effgen S, Ibrahim HH, Pozzi C, Rohde W, Salamini F (2000). On the origin and domestication history of barley. *Mol. Biol. Evol.* 17:499-510.
- Blattner FR, Badani MAG (2001). RAPD data do not support a second center of barley domestication in Morocco. *Genet. Resour. Crop Evol.* 48: 13-19.
- CSA (Central Statistical Agency) (2012). *Crop production forecast sample survey results: Area and crop production forecast for major crops for 2011/12 (Private Peasant Holding, Meher Season).* Central Statistical Agency, Addis Ababa, Ethiopia.
- Dai F, Eviatar N, Dezhi W, Jordi C, Meixue Z, Long Q, Zhonghua C, Avigdor B, Guoxiong C, Guoping Z (2012). Tibet is one of the centers of domestication of cultivated barley. *PNAS* 10: 1-5.
- Dejene T, Andrea MB, Jens L (2010). Morphological diversity of Ethiopian barleys in relation to geographic regions and altitudes. *Hereditas* 147: 154-164.
- Demissie A, Bjornstad A (1996). Phenotypic diversity of Ethiopian

- barley in relation to geographical regions, altitude range and agro-ecological zones: as aid to germplasm collection and conservation strategy. *Hereditas* 124: 17-29.
- Engels JMM (1991). Genetic diversity in Ethiopia barley in relation to altitude. *Genet. Resour. Crop Evol.* 41:67-73.
- Eticha F, Sinebo W, Grausgruber H (2010). On-farm diversity and characterization of barley landraces in the highlands of west Shewa, Ethiopia. *Ethnobot. Res. Appl.* 8:025-034.
- Federer WT, Raghavarao D (1975). On augmented designs. *Biometrics* 31:39-35.
- Grando S, Helena GM (2005). Food Barley: Importance, Uses and Local Knowledge. Proceedings on International Workshop on Food Barley Improvement. Hammamet, Tunisia. ICARDA, Aleppo, Syria. pp. 14-17.
- Harlan JR (1976). Barley. In: Evolution of crop plants. NW. Simmonds (Ed). Longman Press. UK. pp. 93-98.
- IPGRI (1994). Descriptors for barley. International Plant Genetic Resources Institute, Rome, Italy, 52 pp.
- Jalata Z, Amsalu A, Habtamu Z (2011). Variability, heritability and genetic advance for some yield and yield related traits in Ethiopia barley (*Hordeum vulgare L.*) landraces and crosses. *Int. J. Plant Breed. Genet.* 5: 44-52.
- Lakew B, Alemayehu A (2011). Advances and experiences in barley landrace improvement in Ethiopia. In: Mulatu, B. and Grando, S. (Eds.). Barley Research and Development in Ethiopia. Proceedings of the 2nd National Barley Research and Development Review Workshop. November 28-30, 2006. HARC, Holetta, Ethiopia. pp. 31-46.
- Leino MW, Jenny H (2010). Nineteenth century seeds reveal the population genetics of landrace barley (*Hordeum vulgare L.*). *Mol. Biol. Evol.* 27: 964-973.
- Muhe K, Alemayehu A (2011). Diversity and agronomic potential of barley landraces in variable production system in Ethiopia. *World J. Agric. Sci.* 7: 599-603.
- Negassa M (1985). Patterns of phenotypic diversity in an Ethiopian barley collection, and the Arusi- Bale highland as a center of origin of barley. *Hereditas* 102: 139-150.
- SAS Institute Inc. (2004). SAS/STAT, Statistical Software. Version 9.1.3, SAS Institute Inc., Cary, North Carolina, U.S.A.
- Tanto T, Domenico R, Elena B, Roberto P (2009). Genetic diversity of barley landraces from the central highlands of Ethiopia: Comparison between the Belg and Meher growing seasons using morphological traits. *Genet. Resour. Crop Evol.* 56: 1131-1148.
- Tanto T, Domenico R, Elena B, Roberto P (2010). Adaptation and diversity along an altitudinal gradient in Ethiopian barley (*Hordeum vulgare L.*) landraces revealed by molecular analysis. *BMC Plant Biol.* 10: 121.