Vol. 11(6), pp. 115-122, October-December 2019

DOI: 10.5897/JVMAH2019.0781 Article Number: 69BD79662096

ISSN 2141-2529 Copyright © 2019 Author(s) retain the copyright of this article http://www.academicjournals.org/JVMAH



Journal of Veterinary Medicine and Animal Health

Review

African animal trypanosomosis in cattle in Bénin: A review

Soha Sèsséya Arnaud Sas^{1,4*}, Adjibodé Ayodélé Gabriel⁴, Agbankpé Alidéhou Jerrold¹, Farougou Souaïbou³, Youssao Abdou Karim Issaka², Kpodékon Tchokponhoué Tchégninougbo Marc⁴ and Dougnon Tossou Jacques^{1,4}

Received 5 June, 2019: Accepted 8 August, 2019

African animal trypanosomosis is a disease transmitted by tsetse flies and tabanid to domestic mammals which seriously affects their health status and considerably reduces their productivity. This study proposes to make a synthesis on the bovine trypanosomosis and the various works undertaken in Benin country from 1960 to 2018. After general information on bovine trypanosomosis, the various vectorial agents responsible for the disease are explained. The inventory of fixtures of the pathology since 1960 to 2018 followed by the measures and means of control will be detailed. Then, a point on the state of the trypanosome's chemotherapy -resistance facing circulating drug and of the approaches of solutions and the convincing alternatives will be developed. Finally, a study of the impact of transhumance transmission in the maintenance of bovine trypanosomosis supported by the management of conflicts is exposed. The chemotherapy-resistance of the trypanosomes being a true problem affecting the control of the disease, then it is urgent to find endogenous solutions that are reliable through the valorization of medicinal herbs for an effective control of bovine trypanosomosis in Benin. The present work opens a perspective on the definition of innovative alternatives to fight against the bovine trypanosomosis as well as tackle the chemotherapy-resistance against circulating *Trypanosoma* spp.

Key words: Alternatives, chemotherapy, drug, resistance, *Trypanosoma* spp, Bénin.

INTRODUCTION

Animal trypanosomoses are a major constraint to the development of cattle livestock and other ruminants in Considering itself a true scourge in central and western

Author(s) agree that this article remain permanently open access under the terms of the <u>Creative Commons Attribution</u> <u>License 4.0 International License</u>

¹Research Unit of Applied Microbiology and Pharmacology of Natural substances (URMAPha), University of Abomey Calavi, 01 P. O. Box, 2009 Cotonou, Bénin.

²Laboratory of Animal Biotechnology and Meat Technology (LBATV), University of Abomey Calavi, 01 P. O. Box, 2009 Cotonou, Bénin.

³Research Unit in Communicable Disease (URMAT), University of Abomey Calavi, 01 P. O. Box, 2009 Cotonou, Bénin. ⁴Research Laboratory in Applied Biology (LARBA), University of Abomey Calavi, 01 P. O. Box, 2009 Cotonou, Bénin.

^{*}Corresponding author. E-mail: arnaud.soha@gmail.com; (229) 66 17 16 59.

Africa, African animal trypanosomosis persistence is a major animal health problem with consequences for high morbidity and mortality (Hursey and Slingenbergh, 1995; Lekeux, 2006; Odeniran et al., 2018). It is a cyclic disease transmitted mainly by tsetse flies and other blood-feeding insects of Tabanidae may transmit the trypanosomosis mechnically (Troncy et al., 1981; Itard, 1986; Kumar et al., 2012; Sharma et al., 2012; Singla et al., 2013; Sumbria et al., 2015; Bassi et al., 2018), which seriously affects livestock development. Sometimes called "Nagana", it is a serious parasitosis caused by unicellular parasites: Trypanosoma vivax, T. congolense, Trypanosoma brucei, Trypanosoma Trypanosoma suis (Desquesnes et al., 2009; Ahmed et al., 2016). Disease affects ruminants, pigs, horses and carnivores, but has a serious effect on livestock production and lowers the economic power of cattle producers (Shaw et al., 2014). Trypanosomosis causes a lot of economic losses in livestock production (Hurtado et al., 2016). In 2002, Beninese cattle herd was estimated at 1,300,000 of which 10.50 to 45.00% of each herd may have been infected with trypanosomosis (Codjia, 1989).

Trypanocidal drugs rank first among veterinary pharmaceutical inputs (Vaccines, Anthelmintics, Antibiotics and others) sold by PHARNAVET (Assogba, 2001). Chemotherapy and chemoprevention remain the only means of combating African animal trypanosomosis (Giordani et al., 2016), but several poor or counterfeit products are placed on the market (Sutcliffe et al., 2014; Tchamdja et al., 2016), which promotes the resistance of trypanosome species to circulating trypanocides (Van den Bossche and Delespaux, 2011); which makes it difficult to control the disease.

Beninese government has set up several Livestock Development Programs, the most recent being the Milk and Meat Sector Support Program (PAFILAV). This program aims to control various animal pathologies, including African animal trypanosomosis, which is a real constraint to the development of cattle breeding in Benin. The present work allows us to take stock of all the activities carried out on African animal trypanosomosis in Benin and to propose alternatives in face of the alarming drug resistance that nowadays infringes the cattle development in Benin.

Overview on African animal trypanosomosis

Definition

Animal trypanosomoses is a vector-borne haemoparasitic disease (Juyal et al., 2005; Franco et al., 2017; WHO, 2018). It is the most important and seasonal diseases affecting livestock caused by *Trypanosoma* spp. (Diallo et al., 2018; Jaimes-Dueñez et al., 2018). In Africa, this infection is caused by several species of trypanosomes including *Trypanosoma congolense*, *T. vivax*, *T. simiae*,

T. evansi and T. brucei, which are transmitted cyclically by tsetse flies of the genus Glossina (CFSPH, 2009; Enyaru et al., 2010) and also mechanically by several biting insects (Tabnids, stomoxes, etc.) (Salih et al., 2015). It is a chronic disease of cattle and some mammals that that cause significant anemia as main symptom (Brown, 2008). The disease decreases the haematological indices for infected animals by making low the Packed Cell Volume (PCV) value (Ganyo et al., 2018). Animals infected can produces clinical signs of infection, and occasionally death in half of untreated animals (Osório et al., 2008; Desquesnes et al., 2013). Synonyms

Trypanosomosis or typanosomiasis is the name of several diseases notify among vertebrates caused by haemo-protozoan flagellates (trypanosomes) of the genus Trypanosoma. Referring to animals, it is also call Nagana, Tsetse disease, Tsetse fly disease, trypanosomosis (Spickler, 2018). In humans this includes African trypanosomiasis and Chagas disease. other words can also refer to the disease. Among these words we can include: sleeping sickness; African sleeping sickness; catalepsy; encephalitis; encephalitis lethargica; narcolepsy and sleepy distemper.

Etiology and epidemiology

Etiology

Animal trypanosomosis is an important disease due to trypanosome that causes anemia, loss of condition, and emaciation in livestock (Spickler, 2018; Jaimes-Dueñez et al., 2018). Trypanosomes are protozoan parasites in the family Trypanosomatidae. African animal trypanosomosis is caused by those organisms that are transmitted by Tsetse flies (Spickler, 2018). The three most important species are T. congolense (subgenus Nannomonas), T. vivax (subgenus Duttonella) and T. brucei subsp. brucei (subgenus Trypanozoon). There are three variants of *T. congolense*, called the savannah, forest and kilifi (or Kenya Coast) types. African animal trypanosomiasis can also be caused by *T. (Nannomonas)* simiae, T. (Pycnomonas) suis, T. (Nannomonas) godfreyi and T. (Duttonella) uniforme, and possibly by additional unnamed trypanosomes (Acha and Szyfres, 2003; Leukeux, 2006; Spickler, 2018). All trypanosome parasites are extracellular and flagellated (Leukeux, 2006; Brown, 2008).

Two related parasites, *T. brucei* subsp. *gambiense* and *T. brucei* subsp. *rhodesiense*, are transmitted by the bite of Tsetse flies (genus *Glossina*) cause human African trypanosomiasis, which is also known as sleeping sickness (Camara et al., 2005; Chimelli and Scaravilli, 1997; Spickler, 2018). The primary distinction between this disease and African animal trypanosomosis is that these two organisms can evade the innate resistance

humans possess against other tsetse-transmitted African trypanosomes (Spickler, 2018).Livestock can harbor pathogenic parasites for humans in particular *T. rhodesiense* and *T. gambiense* and act as a reservoir (WHO, 2019).

Epidemiology

Epidemiology of trypanosomosis aims to understand a modal of transmission, the incubation period of the disease, the morbidity and the mortality. The disease is transmitted from animal to animal thought the bite of an infected tabanids and tsetse flies such as *Glossina morsitans*, *G. palpalis* and *G. fusca*. The clinical sign of the disease is quite variable from 4 days up to 5 weeks (Brown, 2008). Morbidity can be expected to be very high when the tsetse challenge is also high. Therefore, sick animals or animals infected with any one of the three species of trypanosome and unless treated will probably die of the disease (Brown, 2008).

Diagnosis

The incubation periods start from 4 days to approximately 6 weeks in ruminants. More virulent isolates seem to have a shorter incubation period, often 2 weeks or less (Spickler, 2018). Diagnosis of animal trypanosomosis is based on two types of procedures. It is essentially about a field diagnosis and laboratory diagnosis (Brown, 2008).

Field diagnosis: based on the observation of pathognomonic signs of bovine trypanosomosis. It characterized by sickness, severe anemia in an endemic area, emaciation, watering. The next is the prompt collection of sample for laboratory diagnosis.

Laboratory diagnosis

This part is realized in two steps, the collection of the samples and the technical procedures:

Samples:

The sample can be the serum, blood, thick and thin blood smears, lymph or buffy coat (OIE, 1992; Brown, 2008).

Laboratory procedures:

Trypanosomosis may be diagnosed by microscopic examination of wet and stained thick or thin blood films, impression smears from lymph node aspirates, Polymerase chain reaction (PCR), Enzyme-linked immuno sorbent assay (ELISA) or indirect fluorescent antibody

(IFA) (OIE, 1992; Brown, 2008).

Multiple vectors of African Animal Trypanosomosis (AAT) in Benin

Seven species of tsetse flies have been recorded in Benin (Table 1). They belong to the subgenus Austenina group), Glossina (*morsitans* group) Nemorhina (palpalis group) (Table 1). Three tsetse species predominate in northeastern Benin; they are Glossina tachinoides, G. palpalis gambiensis and G. morsitans submorsitans (Dehoux, 1993). Tabanus biguttatus is one of the most abundant Tabanid kinds in Benin (Vala and LeClercg, 1993). D'Amico et al., (1996): Desguesnes and Dia (2004) reported in Central Africa and Burkina-Faso that flies, Stomoxys and Tabanids are responsible for the mechanical transmission of T. vivax and T. congolense. Several other hematophagous flies are also involved in the transmission of T. vivax to Nigeria. Several other hematophagous flies species of Tabanidae (T. taeniola, T. biguttatus, T. pluto, T. latipes, T. fasciatus, T. subangustus, T. gratus, T. fuscipes, T. par, T. pertinens, T. secedens, T. albipalpus, T. neocopinus and T. thoracinus) are also involved in the transmission of trypanosomosis to Nigeria (Dipeolu, 1977; Ahmed et al., 2005).

Prevalence of African Animal Trypanosomosis in Bénin from 1960 to 2018

Several studies have been conducted to evaluate the prevalence of African bovine trypanosomosis in Benin using microscopy, serological tests and molecular techniques in tsetse flies and animals. Several reports have been based on the assessment of animal overweight, low hematocrit that may be a consequence of trypanosomosis and microscopy (OIE, 2013). For several studies, the buffy coat technique, smears and thick drops observed using microscopy are used. According to Picozzi et al. (2002) and Uilenberg (2011), 75% of cases of trypanosomosis diagnosis are based on microscopy and serological techniques do not allow diagnose of mixed infections, previous ones and new infections. Epidemiological surveillance techniques by molecular characterization provide more details on the infection status of cattle (Picozzi et al., 2002). A summary of studies examining African animal trypanosomosis in Benin from 1960 to 2017 is presented in Table 2 with a wide range of prevalence (5.00 to 89.80%).

National control campaigns in Bénin

Before independence, zootechnical studies and animal disease control were undertaken on two farms in the State of Benin. The first one is Okpara created in 1952 in the department of Borgou and the second Kpinnou

Table 1. Species of tsetse flies present in Benin.

Group Morsitans (Subgroup Glossina)	Group Palpalis (Subgroup Nemorhina)	Group <i>Fusca</i> (Subgroup <i>Austenina</i>) [Forest]	
[Savannah]	[Riverine]		
Glossina longipalpalis	Glossina palpalis palpalis	Glossina fusca congolensis	
Glossina morsitans submorsitans	Glossina palpalis gambiensis	Glossina medicorum	
-	Glossina tachinoides	-	

Source: Pollock (1982); Itard (1986).

Table 2. Summary of research on trypanosomosis in Benin.

Author and year	Animal species	Tests	Prevalence (%)	Localities
Codjia (1989)_	Cattle	Blood smears	22.5	Borgou, Bénin
Doko et al. (1991)	Cattle	Blood smears	19.3	Abomey-Calavi, Bénin
		IFAT	89.8	
		CATT	50.6	
		Trypanolytic test	3.4	
Lekeux (2006)	Cattle	Blood smears	5 - 63	Bénin, 1991-2004
Farougou et al. (2012)	Cattle	Woo method	6.7	Ouaké, Bénin
	Sheep		3.8	
Hestin (2012)	Cattle	Blood smears	13.14	Departement of Zou, Mono, Borgou, Ouémé

created in 1958 in the department of Mono. This led to the creation of the first National Directorate of Livestock and Animal Industries in 1959 on the eve of independence. Due to political unrest in the country until 1972, no effective restructuring of the livestock service was obtained and this contributed to the persistence of major epizootics including bovine trypanosomosis in livestock in Benin. It took the creation of the Public Society for the Development of Animal Resources (SODERA) in 1972 to put in place effective health control policies against diseases with a strong economic impact, including trypanosomosis. In 1993, the Agricultural Services Restructuring Project (PRSA) was set up, led to the creation of the Ministry of Agriculture, Livestock and Fisheries (MAEP), and contributed to the development of animal production. As a result, two projects to improve cattle breeding and control of bovine trypanosomosis were set up. This is the Livestock Development Project (LDP) and the Milk and Meat Sector Support Project (PAFILAV). The Livestock Development Project (LDP) developed in 1997 was launched in the year 2000 and aimed at strengthening food security, improving the general standard of living of pastoralists and agropastoralists, and reducing poverty through development of sustainable livestock farming based on better exploitation of resources and progressive integration of livestock and agriculture. To achieve its objectives, the project has built on the achievements of the second phase of the Animal Production Development Project (APDP). This Project also had a third phase that

ensured the continuity of the measures that ended on June 30, 2006.

The Milk and Meat Sector Support Project was identified in June 2007 by a mission of the African Bank of Development following the request of the Government of Benin in 2005. The project is part of the OSD 2006 to 2011 and the growth strategy for poverty reduction (SCRP, 2007; 2008; 2009), including the SCRP Priority Actions Program which includes improving agricultural productivity and production. The project will thus contribute to the achievement of the specific objectives assigned to these sectors, included in the Strategic Plan for the Revival of the Agricultural Sector (PSRSA) and aimed at improving the breeding of livestock and control of diseases such as trypanosomosis, other parasitic, viral and bacterial diseases.

The implementation of numerous breeding projects, Pan African Program for the Control of Epizootics (PACE), Livestock Development Project, stage III (LDP III), Livestock Development Project in East Borgou (PDEBE), and Livestock Development Support Project in Borgou (PADEB) over the last ten years has reached cattle and small ruminants. The main activities focused on animal health, zootechnics and research and development (PDE, 2007).

Trypanocidal drug resistance and sub-standard drugs

For several decades, the control of African animal

trypanosomosis has been mainly based on the use of trypanocides for prevention and treatment (Murray et al., 1998; Giordani et al., 2016; Dagnachew et al., 2017). Animal disease control measures through chemotherapy better protect animals than any other methods (Budd, 1999). The increase in trypano-resistance that promotes the resistance of trypanosome species to circulating trypanocidal drugs (Van den Bossche and Delespaux, 2011) is due to the negligence of some unscrupulous trypanocidal drug deliverers, the continued and abusive use of trypanocidal drugs by breeders or unqualified people, under-dosing, finally a limited number of circulating trypanocidal molecules (Assogba, 2001; Dagnachew et al., 2017) as well as the circulation of trypanocidal products of poor quality or counterfeit on the market (Vitouley et al., 2012; Sutcliffe et al., 2014; Tchamdja et al., 2016).

Some studies have also shown the misuse of veterinary drugs as the major cause of drug resistance in Africa (Teko-Agbo, 2008; Gberindyer et al., 2014). These states of affairs make it difficult to control the disease. According to Mainguet (2000), the resistance of a trypanosome species is conserved during the life cycle of the tsetse fly. This period varies from one species to another, 16 months for *T. vivax* and 12 months for *T. congolense*. Trypanocidal drug resistance is a critical issue that needs to be constantly addressed. Resistance to the trypanocidal drugs used is a major problem in livestock farming in Africa (Delespaux et al., 2008) but the breeders do not have a good knowledge on the concept of drug resistance (Sinyangwe et al., 2004).

Cases of resistance of trypanosomes to isometamidiun are reported (Delespaux et al., 2008) throughout Africa and more precisely in Ethiopia and Burkina Faso (Clausen, 2005), Mozambique (Jamal et al., 2005), Nigeria (Ilemobade, 1979) and Bénin (Vitouley, 2014). To reduce the risk of developing drug resistance, the frequency of trypanocidal use must be kept to a minimum (Geerts and Holmes, 1998).

Impact of transhumance in the maintenance of bovine trypanosomosis and management of conflicts in Bénin

In Bénin, Fulani people migrate to southern areas in search of fodder and water due to drought (Agossou et al., 1998). Livestock management is characterized by migratory movements. These displacements sometimes create conflicts between breeders and crop farmers and favor the maintenance of vector-borne diseases such as bovine trypanosomosis (Agossou et al., 1998). Conflicts occur mainly at the beginning of the rainy season when pastoralists prefer new fallow regrowth for the grazing of their animals. This poses a problem of poor occupation of spaces because of the planting of crops (De Haan, 1997). To solve this problem, the Beninese State has made a lot of effort and has taken steps for setting up

transhumance committees at local, national and regional levels. Few improvements have been achieved. With the option of decentralization of the Beninese State and transfer of competence to peasant organizations that emanate from the historic decisions of the National Conference from 19 to 28 February 1990, research and development has been developing since 1996 in the north (Agossou et al., 2003) and Benin Center (Agossou et al., 2003; Maliki et al., 2001), a participatory approach to assist some rural communities in the learning and conflict management experience. Transhumance is regulated by laws, decrees, decisions and regulations made at municipal and national level (Djohy, 2010). The laws that regulate transhumance are: Law No. 87-013 of 21 September 1987 regulating pasturage, keeping of domestic animals and transhumance; Interministerial Order 1992-No. 010/MISAT/MDR/D-CAB of 20 January 1992, establishing, organization, functions and operation of transhumance committees in the Republic of Benin: Law No.93-009 of July 2, 1993 bearing forest regime in the Republic of Benin and Decree No.96-271 of July 2, 1996 bearing the terms of application of law No. 93-009 1993; the 1994 decree July 2, /MISAT/MDR/DGAR concerning the organization of transhumance 1993-1994; Law No. 2002-016 of October 18, 2004, on the fauna regime in the Republic of Benin (Without implementing decree); Interministerial Order 2006-N° 2176/APRM/MSPCL/DCAB/SGM/ DRH/DE/SA of July 7, 2006 mandatory and synchronous rendering the vaccination of the animals against the septicemia, hemorrhagic and the Contagious Peri-pneumonia in the bovine species, throughout the national territory and decree No. 2009-241 of June 9, 2009 establishing the committee responsible for resolving recurrent transhumance problems between Fulani pastoralists and farmers in the national territory.

In all the communes concerned by transhumance, the town halls of the latter take orders to regulate transhumance in their territories. We can quote: decree No. 54/024/MKDI/SG/SGA/ SA of July 27, 2006, on creation, composition and functioning of communal committee of transhumance of the municipality of Kandi; Decree No. 014/M-CKM/SG/BAGD of 1 October 2004 regulating grazing, keeping of domestic animals and transhumance in the commune of Karimama; Decree No. 16/MCKM/SG/BAGD of October 20, 2004, establishing, assigning and functioning of communal, district and village committees, prevention and management of livestock-farmer conflicts in the municipality of Karimama the decree No. 54/024/MOI/SG/SGA/SA September 16, 2009, concerning creation, composition and functioning of communal committee of transhumance of the commune of Ouinhi.

CONCLUSION

Benin is a country with a high agropastoral activity, but

breeding is slow to take off because of sanitary constraints. African animal trypanosomosis is one of the major constraints affecting the economy and productivity of livestock. Despite the control measures implemented for several years through to livestock development projects, bovine trypanosomosis persists and continues to be a major obstacle to improving livestock productivity in Benin. In order to control the disease and ensure good livestock performance, effective control measures against tsetse vectors and extension of control policy to all endemic areas will be required. It is also clear that the government and authorities in charge of livestock development are making better arrangements for the operationalization of the control and eradication of African animal trypanosomosis.

ETHICAL STATEMENT

The present work was conducted under the permission of the University of Abomey-Calavi through the Project FNRSIT-FC 2015.

CONFLICT OF INTERESTS

The authors have not declared any conflict of interests.

REFERENCES

- Acha PN, Szyfres B (2003). Zoonoses and communicable diseases common to man and animals (Vol. 580). Pan American Health Organization, Regional Office of the WHO, Washington, USA 384. https://www.socgenmicrobiol.org.uk/pubs/micro_today/book_reviews/MTMAY02/MTM02_29.cfm
- Agossou V, Baltissen G, Savi A (1998). Prévention de conflits entre agriculteurs et éleveurs: expérience dans quelques villages du Borgou (Nord-Bénin). Bulletin de la Recherche Agronomique Numéro27 mars. pp. 28-42.
- Agossou V, Gnanglè P, Adjé I (2003). Conflits entre agriculteurs et éleveurs: expériences d'apprentissage en prévention de conflits dans quelques villages du centre et Nord Bénin. Protocole de recherche RN 3. Fonds Compétitif/APPRA/PADSE 6 p.
- Ahmed AB, Okiwelu SN, Samdi SM (2005). Species diversity, abundance and seasonal occurrence of some biting flies in Southern Kaduna, Nigeria. African Journal of Biomedical Research 8(2):113-118.
- Ahmed SK, Rahman AH, Hassan MA, Salih EM, Paone M, Cecchi G (2016). An atlas of tsetse and bovine trypanosomosis in Sudan. Parasites and Vectors 9(1):194. https://doi.org/10.1186/s13071-016-1485-6
- Assogba DH (2001). Le marché des médicaments vétérinaires au Bénin. Séminaire sur l'utilisation des trypanocides en Afrique subsaharienne, Dakar, EISMV 6:65-68.
- Bassi PB, de Araújo FF, Garcia GC, Vinícius da Silva M, Oliveira CJF, Bittar ER, Bittar JFF (2018). Parasitological and immunological evaluation of cattle experimentally infected with Trypanosoma vivax. Experimental Parasitology 185:98-106. https://doi.org/10.1016/j.exppara.2018.01.010
- Brown C (2008). Trypanosomiasis (African). Foreign animal diseases. 7th ed. St Joseph, Mo: United States Animal Health Association pp. 405-409.
- Budd L (1999). DFID-funded tsetse and trypanosome research and development since 1980. Economic Analysis 2:123.

- Camara M, Kaba D, Kagbadouno M, Sanon JR, Ouendeno FF, Solano P (2005). La trypanosomose humaine africaine en zone de mangrove en république de Guinée: caractéristiques épidémiologiques et cliniques de deux foyers voisins. Médecine Tropicale 65(2):155.
- Center for Food Security and Public Health (CFSPH) (2009). African animal trypanosomiasis. The Center for Food Security and Public Health. www.cfsph.iastate.edu
- Chimelli L, Scaravilli F (1997). Trypanosomiasis. Brain Pathology 7(1):599-611. https://doi.org/10.1111/j.1750-3639.1997.tb01077.x
- Clausen PH (2005). Studies on the diagnosis, development and distribution of drug- resistant trypanosomes in cattle herds in selected areas of East and West Africa. [Études sur le diagnostic, le développement et la répartition des trypanosomes chimiorésistants dans des troupeaux de bovins dans des zones sélectionnées d'Afrique de l'Est et de l'Ouest.] Thèse, Freie Universitat Berlin 136 p.
- Codjia V (1989). Prospection entomologique et étude de l'incidence de la trypanosomose animale africaine (T.A.A.) et autres hémoparasitoses du bétail dans les districts de Nikki, Kalabe et Segbana. Direction de l'é1evage et des Industries animales. Ministère du développement rural et de faction coopérative, Bénin. Report of Project PNUD/FAO Benin/84/01 I. 68 p.
- Dagnachew S, Biniam T, Addissu A, Meseret T, Hagos A, Tim R, Getachew A, Dave J B, Getachew T, Bruno MG (2017). Prevalence of bovine trypanosomosis and assessment of trypanocidal drug resistance in tsetse infested and non-tsetse infested areas of Northwest Ethiopia. Parasite Epidemiology and Control 2(2):40-49. https://doi.org/10.1016/j.parepi.2017.02.002
- d'Amico F, Gouteux JP, Le Gall F, Cuisance D (1996). Are stable flies (Diptera: Stomoxyinae) vectors of Trypanosoma vivax in the Central African Republic?. Veterinary Research, BioMed Central 27(2):161-170
- De Haan LJ (1997). Agriculteurs et éleveurs au Nord-Bénin: écologie et genres de vie. Karthala Editions P 92.
- Dehoux JP (1993). Lutte contre Glossina tachinoides au Bénin. Utilisation particulière de piège-pneus imprégnés de deltaméthrine. Revue d'élevage et de médecine vétérinaire des pays tropicaux 46(4):581-589.
- Delespaux V, Geysen D, Van den Bossche P, Geerts S (2008). Molecular tools for the rapid detection of drug resistance in animal trypanosomes. Trends in Parasitology 24(5):236-242.
- Desquesnes M, Biteau-Coroller F, Bouyer J, Dia ML, Foil L (2009). Development of a mathematical model for mechanical transmission of trypanosomes and other pathogens of cattle transmitted by tabanids. International Journal for Parasitology 39(3):333-346. https://doi.org/10.1016/j.ijpara.2008.07.004
- Desquesnes M, Dia ML (2004). Mechanical transmission of Trypanosoma vivax in cattle by the African tabanid (Atylotus fuscipes). Veterinary Parasitology 119(1):9-19. https://doi.org/10.1016/j.vetpar.2003.10.015
- Desquesnes M, Holzmuller P, Lai DH, Dargantes A, Lun ZR, Jittaplapong S (2013). Trypanosoma evansi and surra: a review and perspectives on origin, history, distribution, taxonomy, morphology, hosts, and pathogenic effects. BioMed Research International Volume 2013, Article ID 194176, 22 p http://dx.doi.org/10.1155/2013/194176
- Diallo T, Singla LD, Sumbria D, Kaur P and Bal MS (2018). Conventional and molecular diagnosis of haemo-protozoan infections in Cattle and equids from Republic of Guinea and India. Indian Journal of Animal Research 52(8):1206-1211. https://doi.org/10.18805/ijar.v0iOF.6988
- Djohy G (2010). Une analyse des dynamiques sociopolitiques et organisationnelles d'adaptation des éleveurs transhumants dans l'Alibori (Nord Bénin). Thèse d'Ingénieur agronome, FA/UP, Université de Parakou, Bénin P 122.
- Doko A, Guedegbe B, Baelmans R, Demey F, N'diaye A, Pandey VS, Verhulst A (1991). Trypanosomiasis in different breeds of cattle from Benin. Veterinary Parasitology 40(1-2):1-7.
- Enyaru JC, Ouma JO, Malele II, Matovu E, Masiga DK (2010). Landmarks in the evolution of technologies for identifying trypanosomes in tsetse flies. Trends Parasitology 26(8):388-394. https://doi.org/10.1016/j.pt.2010.04.011
- Farougou S, Doko Allou S, Sankamaho I, Codjia V (2012). Prevalence

- of Trypanosome Infections in Cattle and Sheep in the Benin's West Atacora Agro-ecological zone. Tropicultura 30(3):141-146.
- Franco JR, Čecchi G, Priotto G, Paone M, Diarra A, Grout L, Argaw D (2017). Monitoring the elimination of human African trypanosomiasis: Update to 2014. PLoS Neglected Tropical Diseases 11(5):e0005585. https://doi.org/10.1371/journal.pntd.0005585
- Ganyo EY, Boampong JN, Masiga DK, Villinger J, Turkson PK (2018). Haematology of N'Dama and West African Shorthorn cattle herds under natural Trypanosoma vivax challenge in Ghana. F1000 Research 7(314). https://doi.org/10.12688/f1000research.14032.2
- Gberindyer FA, Onyeyili PA, Bosha JA (2014). Quality control properties of some brands of veterinary albendazole boluses common in Nigeria. Journal of Pharmacy and Pharmacology 2:135-139.
- Geerts S, Holmes PH (1998). Drug management and parasite resistance in bovine trypanosomiasis in Africa. Drug management and parasite resistance in bovine trypanosomiasis in Africa. Food and Agriculture Organization (FAO), Record Number: 20000804299, 1, P 31.
- Giordani F, Morrison LJ, Rowan TG, De Koning HP, Barrett MP (2016). The animal trypanosomiases and their chemotherapy: A review. Parasitology 143(14):1862-1889.
 - https://doi.org/10.1017/S0031182016001268
- Hestin T (2012). Les stratégies de développement de l'élevage bovin au Bénin au travers de la mise en place de deux projets consécutifs: le P.D.E puis le P.A.F.I.L.A.V. Thèse n° 083, Université Claude-Bernard Lyon I. 130 p.
- Hursey BS, Slingenbergh J (1995). The tsetse fly and its effects on agriculture in sub-Saharan Africa. World Animal Review pp. 67-73.
- Hurtado OJB, Castro PDJ, Giraldo-Ríos C (2016). Reproductive failures associated with Trypanosoma (Duttonella) vivax. Veterinary Parasitology 229:54-59. https://doi.org/10.1016/j.vetpar.2016.09.017
- Ilemobade AA (1979). Drug sensitivity of mouse –infective Trypanosoma vivax isolates in cattle and sheep. In: International Scientific Council for Trypanosomiasis Research and control (ISCTRC), (16th Meeting, Yaounde, Cameroon) pp. 251-253.
- Itard J (1989). African animal trypanosomiasis. In. Manual of Tropical Veterinary Parasitology. M. Shah-Fischer & R. Ralph Say. (eds) (English translators). CAB International, UK.
- Jaimes-Dueñez J, Triana-Chávez O, Mejía-Jaramillo AM (2018). Spatial-temporal and phylogeographic characterization of Trypanosoma spp. in cattle (Bos taurus) and buffaloes (Bubalus bubalis) reveals transmission dynamics of these parasites in Colombia. Veterinary Parasitology (249):30-42. https://doi.org/10.1016/j.vetpar.2017.11.004
- Jamal S, Sigauque I, Macuamule C, Neves L, Penzhorn BL, Marcotty T, Bossche PVD (2005). The susceptibility of Trypanosoma congolense isolated in Zambezia Province, Mozambique, to isometamidium chloride, diminazene aceturate and homidium chloride. Onderstepoort Journal of Veterinary Research 72(4):333-338. https://hdl.handle.net/10520/EJC86223
- Juyal PD, Singla LD, Kaur P (2005). Management of surra due to Trypanosoma evansi in India: an overview. Infectious diseases of domestic animals and zoonosis in India 75:109-120.
- Kumar H, Gupta MP, Sidhu PK, Mahajan V, Bal MS, Kaur K, Ashuma, Verma S, Singla LD (2012) An outbreak of acute Trypanosoma evansi infection in crossbred cattle in Punjab, India. Journal of Applied Animal Research 40(03):256-259. https://doi.org/10.1080/09712119.2012.667651
- Lekeux M (2006). La trypanosomose bovine africaine: Généralités et situation au Bénin (Doctoral dissertation, Thèse vétérinaire, Université de Lyon).
- Mainguet JM (2000). Apport d'une étude sur le terrain dans le choix d'une stratégie de lutte contre les trypanosomoses bovines en république centrafricaine. Thèse pour obtenir le grade de Docteur vétérinaire soutenue à la faculté de médecine de Créteil P 262.
- Maliki R, Amadji F, Adjé I, Hounou J, Nodichao S (2001). Gestion des espaces agro-pastoraux: conflits entre agriculteurs et éleveurs aucentre Bénin. Acte 2 de l'Atelier Scientifique Sud et Centre du 12 au 13 décembre 2001 à Niaouli pp. 577-589.
- Murray M, D'Ieteren G, Authie E, Wissocq N (1998). Trypanotolerance, an option for sustainable livestock production in areas at risk from trypanosomosis. OIE Revue Scientifique et Technique 17(1):154-175.

- Odeniran PO, Ademola IO, Macleod ET, Welburn SC (2018). Bovine and small ruminant African animal trypanosomiasis in Nigeria–A review. Veterinary Parasitology: Regional Studies and Reports 13:5-13. https://doi.org/10.1016/j.vprsr.2018.03.001
- Office International des Epizooties (OIE) (1992). Manual of Standards for Diagnostic Tests and Vaccines Second Edition 831 p.
- Office International des Epizooties (OIE) (2008). Standardized Techniques for the Diagnosis of Tsetse Transmitted Trypanosomosis. Terrestrial Manual, OIE, Rome, Italy P 49.
- Office International des Epizooties (OIE) (2013). Trypanosomosis (tsetse flies transmitted). In: Terrestrial Manuscript.
- Osório ALAR, Madruga CR, Desquesnes M, Soares CO, Ribeiro LRR, Costa SCGD (2008). Trypanosoma (Duttonella) vivax: its biology, epidemiology, pathogenesis, and introduction in the New World-a review. Memórias do Instituto Oswaldo Cruz 103(1):1-13. http://dx.doi.org/10.1590/S0074-02762008000100001
- PDE (2007). Programme de développement de l'élevage pour les cinq prochaines années (2007-2012). Rapport de Consultation 243 p.
- Picozzi K, Tilley A, Fevre EM, Coleman PG, Magona JW, Odiit M, Welburn SC (2002). The diagnosis of trypanosome infections: applications of novel technology for reducing disease risk. African Journal of Biotechnology 1(2):39-45.
- Pollock JN (1982). Training manual for tsetse control personnel. Vol 1. FAO Publ. No M/P5178/E.
- Salih DA, Hussein AM EI and Singla LD (2015). Diagnostic approaches for tick-borne haemoparasitic diseases in livestock. Journal of Veterinary Medicine and Animal Health 7(2):45-56. http://doi.org/10.5897/JVMAH2014.0345
- Sharma P, Juyal PD, Singla LD, Chachra D and Pawar H (2012). Comparative evaluation of real time PCR assay with conventional parasitological techniques for diagnosis of Trypanosoma evansi in cattle and buffaloes. Veterinary Parasitology 190:375-382. https://doi.org/10.1016/j.vetpar.2012.07.005
- Shaw APM, Cecchi G, Wint GRW, Mattioli RC, Robinson TP (2014).

 Mapping the economic benefits to livestock keepers from intervening against bovine trypanosomosis in Eastern Africa. Preventive Veterinary Medicine 113(2):197-210. https://doi.org/10.1016/j.prevetmed.2013.10.024
- Singla LD, Sharma A, Kaur P, Tuli A, Bhat SA, Bal MS (2013). Bovine trypanosomosis in Punjab: Assessment of seroprevalence by CATT/T. evansi. International Journal of Advanced Research 1(9):364-371.
- Sinyangwe L, Delespaux V, Brandt J, Geerts S, Mubanga J, Machila N, Eisler MC (2004). Trypanocidal drug resistance in eastern province of Zambia. Veterinary Parasitology 119(2-3):125-135. https://doi.org/10.1016/j.vetpar.2003.11.007
- Spickler AR (2018). African Animal Trypanosomiasis. Retrieved from http://www.cfsph.iastate.edu/DiseaseInfo/factsheets.php
- Sumbria D, Singla LD, Sharma A, Bal MS, Kumar S (2015). Multiplex PCR for detection of Trypanosoma evansi and Theileria equi in equids of Punjab, India. Veterinary Parasitology 211(3-4):293-99. https://doi.org/10.1016/j.vetpar.2015.05.018
- Sutcliffe OB, Skellern GG, Araya F, Cannavan A, Sasanya JJ, Dungu B, Mattioli RC (2014). Animal trypanosomosis: making quality control of trypanocidal drugs possible. Revue des Sciences et Techniques, 33(3):813 830.
- Tchamdja E, Kulo A E, Akoda K, Teko-Agbo A, Assoumy AM, Niang EMM, Hoppenheit A (2016). Drug quality analysis through high performance liquid chromatography of isometamidium chloride hydrochloride and diminazene diaceturate purchased from official and unofficial sources in Northern Togo. Preventive Veterinary Medicine 126:151-158. https://doi.org/10.1016/j.prevetmed.2016.02.001
- Teko-Agbo A (2008). Quality of veterinary medicinal products in circulation in Cameroun and Senegal. In: OIE Conference on Veterinary Medicinal Products in Africa, 25–27 March, Paris: OIE. Dakar, Senegal.
- Troncy PM, Itard J, Morel PC (1981). Précis de parasitologie vétérinaire tropicale. Institut d'élevage et de médecine vétérinaire des pays tropicaux.
- Uilenberg G (2011). A Field Guide for the Diagnosis, Treatment and Prevention of African Animal Trypanosomosis. Food and agriculture

- organization of the United Nations, Rome pp. 34-111.
- Vala JC, Leclercq M (1993). On the presence of Tabanus biguttatus Wiedemann (Diptera: Tabanidae) in Benin. In Bulletin & Annales de la Société Royale Belge d'Entomologie 129(7/9):231-236.
- Van den Bossche P, Delespaux V (2011). Options for the control of tsetse-transmitted livestock trypanosomosis. An epidemiological perspective. Veterinary Parasitology 181(1):37-42. https://doi.org/10.1016/j.vetpar.2011.04.021
- Vitouley HS (2014). Trypanocidal drugs and the problem of drug resistance in West Africa, Thesis submitted in fulfillment of the requirements for the degree of Doctor in Veterinary Science (PhD), Faculty of Veterinary Medicine, Ghent University P 197.
- Vitouley HS, Sidibe I, Bengaly Z, Marcotty T, Van Den Abbeele J, Delespaux V (2012). Is trypanocidal drug resistance a threat for livestock health and production in endemic areas? Food for thoughts from Sahelian goats infected by Trypanosoma vivax in Bobo Dioulasso (Burkina Faso). Veterinary Parasitology 190(3-4):349-354. https://doi.org/10.1016/j.vetpar.2012.07.015

- World Health Organization (WHO) (2018). https://www.afro.who.int/health-topics/trypanosomiasis-african.
- World Health Organization (WHO) (2019). https://www.who.int/fr/news-room/fact-sheets/detail/trypanosomiasis-human-african-(sleeping-sickness). Consulted the Mars, 5th 2019.