Short Communication

Evaluation of the bacteriological characteristics of poultry litter as feedstuff for cattle

E. M. Ngodigha^{1*} and O.J. Owen²

¹Department of Fisheries/Livestock Production Technology, Niger Delta University, Wilberforce Island, P.M.B. 071, Yenagoa, Bayelsa State, Nigeria.

Accepted 17 February, 2009

The bacteriological characteristics of poultry litter as feedstuff for cattle was evaluated by subjecting it to heat treatment by deep stacking. Litter samples were assayed for pathogens before heat treatment commenced. The litter was bagged, stacked and covered with thick, black plastic cellophane in a roofed building for 21 days. The temperature of the stack was monitored with a thermometer and the readings recorded. At the end of the heat treatment, particles were removed from the litter with a mechanical sieve. Litter samples were subjected to proximate analysis, mineral composition profile, cultured in a McConkey medium and then incubated for 24 – 48 h at 37 – 42°C for various pathogens. Results showed that litter dry matter (DM) and crude protein (CP) contents were 87 and 20% respectively. Mineral composition varied from 0.10% for sodium to 4.50%, for phosphorus. The isolation temperature of pathogens in the untreated poultry litter ranged between 37°C for Salmonella sp. and Mycobacterium, 41°C for Clostridium and Escherichia coli to 42°C for Staphylococcus. No pathogen was isolated after heat treatment (40.1 – 55°C) for 21 days. Poultry litter can be used as feedstuff for cattle if processed properly to eliminate pathogens and the nutrient levels equalized.

Key words: Bacteriological characteristics, cattle, deep stacking, pathogen, poultry litter.

INTRODUCTION

The rising cost of animal feeds has continued to be a major problem in developing countries as feed cost is about 70 – 75% of the total cost of production, compared to about 50 – 60% in developed countries (Nwogu et al., 2003). There is therefore the need to source for alternative feed ingredients that can lead to a reduction in the cost of feed and hence the total cost of production. Poultry litter has such potential as it has been found to contain protein, which can replace soya bean meal in rabbit diets (Onimisi and Omage, 2006).

Processing of poultry litter is necessary for destruction of potential pathogens, improvement of handling and storage characteristics, and maintenance or enhancement of palatability (Fontenot, 2000). Pathogenic microbial organisms gain access to the animal body through conta-

minated feed and water (Youdeowei et al., 1999). The presence of these pathogenic microorganisms impact negatively on feed utilization and physiological functions within the animal system.

According to Simonsen et al. (1987), poultry accounted for 50.1% of all non-human *Salmonella* isolates. Clegg et al. (1995) and Hogg et al. (1990) also confirmed the isolation of many microorganisms from the faeces of hens.

E. coli infection, which is responsible for major losses in the poultry industry, is commonly found in poultry litter and faecal material (Fontenot, 2000). Bio-deterioration takes place in poultry manure as part of the metabolic activities of the bacteria present in it (Onion et al., 1981). Some of these microorganisms are pathogenic. The degree of bio-deterioration depends on the moisture content, relative humidity, temperature and pH of the bacterial medium. This study was therefore designed to evaluate the bacterial profile in poultry litter so as to authenticate its safety as cattle feed ingredient.

²Department of Animal Science, Rivers State University of Science and Technology, P.M.B 5080, Port Harcourt, Rivers State, Nigeria.

^{*}Corresponding author. E-mail: m_ngodighaa@yahoo.co.uk. Tel: +234 8033413211.

Table 1. Chemical composition of deep stacked layers' poultry litter (dry matter basis).

Components	Composition
Dry matter (% DM)	87.00
Crude protein (CP) %	20.00
Energy (KJ/kg)	2601.72
Crude fibre %	10.4
Ether extract (EE) %	2.20
Ash %	18.50

Table 2. Mineral composition of deep stacked layers' poultry litter.

Components	Composition (%)	
Phosphorus	4.50	
Calcium	2.00	
Sodium	0.10	
Potassium	2.05	
Magnesium	0.48	

MATERIALS AND METHODS

Shovels were used to collect poultry litter from the layers' section of the Teaching and Research Farm of the Rivers State University of Science and Technology, Port Harcourt (4° 52'N; 6° 57' E). The litter was placed in empty feed bags. Before subjecting the litter to heat treatment by deep stacking, samples were collected to assay for bacterial profile at the Microbiology Laboratory of the Institution.

The litter was subjected to a heat cycle treatment created by deep stacking. This was achieved by stacking the bagged poultry litter, which was then covered with thick, black plastic cellophane in a roofed building for 21 days. The temperature of the pile was constantly monitored with a thermometer and the readings recorded. At the end of the heat treatment, particles were removed from the litter with a mechanical sieve. Litter samples were subjected to proximate analysis and mineral composition profile (Tables 1 and 2), procedures of AOAC (1990). Samples were also cultured in a McConkey medium and then incubated for 24 – 48 h at $37-42^{\circ}\text{C}$ for various pathogens (Table 3).

RESULTS AND DISCUSSION

The chemical and mineral compositions of the deep stacked layers' litter are shown in Tables 1 and 2. The dry matter (DM) and crude protein (CP) contents were 87 and 20%, respectively. The values obtained for energy, crude fibre, ether extract and ash were 2601.72 KJ/kg, 10.4, 2.20 and 18.50%, respectively. The mineral composition of the recycled poultry litter in Table 2 shows that Phosphorus (P), Calcium (Ca), Sodium (Na), Potassium (K) and Magnesium (Mg) have values of 4.5, 2.00, 0.10, 2.05 and 0.48%, respectively. Phosphorus (P) had the highest value when compared with the other minerals.

The bacterial organisms isolated in the layers' poultry litter before and after heat treatment are shown in Table 3. In the present study, the isolation temperature of pa-

Table 3. Pathogenic organisms isolated in layers' poultry litter before and after heat treatment.

Pathogens	Before heat treatment	After heat treatment	Isolation temperature (°C)
Escherichia coli	+	NPI	41
Salmonella sp.	+	NPI	37
Mycobacterium	+	NPI	37
Shigella	NPI	NPI	-
Listeria	NPI	NPI	-
Mould	NPI	NPI	-
Yeast	NPI	NPI	-
Clostridium	+	NPI	41
Proteus	NPI	NPI	-
Staphylococcus	+	NPI	42

^{+,} Scanty; NPI, No pathogen isolated.

thogens in untreated poultry litter ranged between 37°C for *Salmonella* and *Mycobacterium*, 41°C for *Clostridium* and *E. coli* to 42°C for *Staphylococcus*.

The differing bacterial presence in the untreated poultrylitter confirmed the findings of Olutiola et al. (1991). The high core temperatures, $(41 - 42^{\circ}C)$ and (37)- 39.5°C) reported by Charles (1975) and Fielding (1991) in birds and rabbits respectively, confers a naturalinfection-control device against some bacteria. Most of the bacteria isolated in this study were those that are commonly experienced by man and animals in their dayto-day exposure and to which their bodies have developed some degree of relative resistance (Onimisi and Omage, 2006). These findings are in agreement with earlier results obtained by Fuller (1973); Thornton and Gracev (1976); and Awonivi et al. (2004), who used maggot meal based diets (another unconventional feed ingredient) on broiler chickens and isolated bacterial organisms in their visceral organs.

With regard to the safety aspects of using poultry litter in cattle rations, the results obtained in this study showed that, pathogens were not isolated after heat treatment (40.1 - 55.0°C) for 21 days. This was due to the action of heat to which the pathogens were subjected (Davis, 1999). It is worthy of note that the survival of microorganisms in dietary ingredients varies widely with its moisture content (Onion et al., 1981). The destruction of bacterial organisms observed after the heat treatment may also be aided by a reduction in the moisture content of the poultry litter (Fontenot, 2000). In this study, a moisture content of 13% was observed. The maximum temperature obtained during the heat treatment was 55°C. This result is in agreement with the findings of Chaudhry (1990), who obtained a temperature of 54.6°C at 15% moisture content. Fontenot (2000) reported that increaseing the moisture content above 35% resulted in lower maximum temperatures. The maximum temperature therefore appears to be related to the moisture level in the litter.

The observed gradual increase in temperature to the maximum level attained (55°C) during the heat treatment, was due to the biological activity of the microorganisms which might have contributed to the killing of pathogenic organisms.

Conclusion

The nutritional value of poultry litter as feedstuff for cattle has been well established by many authors. Poultry litter can be used as feedstuff for cattle if processed properly to eliminate pathogens and the nutrient levels equalized. This study, which evaluated the bacterial profile in poultry litter, is vital and complementary to its suitability as feedstuff in cattle rations.

REFERENCES

- AOAC (1990). Association of Analytical Chemists. Official methods of analysis. 15th edition, Arlington Virginia, U.S.A.
- Awoniyi TAM, Adebayo A, Aletor VA (2004). A study of some erythrocyte indices and bacteriological analysis of broiler chickens raised on maggot-meal based diet. Int. J. Poult. Sci. 3 (6): 386-390.
- Charles OW (1975). The economic feasibility of processed animal waste. World Poult. Sci. J. 31: 271-275.
- Chaudhry SM (1990). Processing and nutritive value of poultry litter and slaughtering house by-products. Ph.D thesis, Virginia Polytechnic and State University, Blacksburg. pp. 68-79
- Clegg FG, Jones T, Smart JL, McMaurty MJ (1995). Bovine botulism associated with broiler litter waste. Vet. Record. 117: 22-30.
- Davis GV (1999). Feeding broiler litter to beef cattle. CES Publication, University of Arkansas, U.S.A.
- Fielding D (1991). Rabbits. Tropical Agriculture Series. CTA/Macmillan Education Limited, London. pp. 21- 25.
- Fontenot JP (2000). Utilization of poultry litter as feed for beef cattle. Anim. Residuals Managt. 19: 234-252.
- Fuller R (1973). Ecological studies on the *Lactobacillus* flora associated with crop epithelium of fowl. J. Appl. Bacteriol.. 36: 331-337.

- Nwogu FC, Egbunike GN, Ononogbu CE, Fapohunda JB, Ogbonna JU (2003). Biologic and economic effects of including different agroindustrial by-products in weaner rabbit production. Proceedings of 8th
- Hogg RA, White VJ, Smith GR (1990). Suspected botulism in cattle associated with poultry litter. Vet. Record. 126: 476- 488.
- Annual Conference. Animal Science Association of Nigeria (ASAN). Minna. pp. 36-38.
- Olutiola PO, Famurewa O and Sontag HG (1991). An introduction to General Microbiology. pp. 21-27.
- Onimisi PA, Omage JJ (2006). An evaluation of poultry litter as feedstuff for growing rabbits. Livestock Res. Rur. Dev. 18 (11): 5-10.
- Onion AHS, Allsop DO, Eggins HOW (1981). Bio-deterioration and microbial activities. In: Smith's Introduction to Industrial Mycology. 7th edn. (London) Ltd.
- Simonsen B, Bryan FL, Christian JHB (1987). Prevention and control of food-borne *salmonellosis* through application of Hazard Analysis Critical Control Point (HACCP). Int. J. Food Micr. 4: 227-247.
- Thornton H, Gracey JF (1976). Bacteria and viral diseases. In: Textbook of Meat Hygiene. 6th Ed. E.L.B and Balliere Tindall.
- Youdeowei A, Ezedinma FOI, Onsi OC (1999). Introduction to Tropical Agriculture. Longman Ltd., Lagos. pp. 20-130.