

*Full Length Research Paper*

# Effects of 1-MCP (1-methylcyclopropene) and STS (silver thiosulphate) on the vase life of cut *Freesia* flowers

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Accepted 16 July, 2010

**This study was to determine the effects of 1-methylcyclopropene (1-MCP) and silver thiosulphate (STS) treatment on the vase life of *Freesia* “Cordula” flowers. 1-MCP and STS extended the vase life of florets and spikes of cut -*Freesia* “Cordula”. The longest spike vase life was also obtained in the 1-MCP (4 ml.l<sup>-1</sup> + 3 hours) treatment as 9.06 days, spike vase life of control group flowers was 6.33 days.**

**Key words:** *Freesia*, 1-methylcyclopropene (1-MCP), silver thiosulphate (STS), vase life.

## INTRODUCTION

*Freesia refracta* (Jacq) is one of the most commonly produced cut – flower species among bulbous, tuberous and rhizomous ornamental plants, especially in European countries (Zencirkiran, 2002). Cut - *Freesia* flowers have a relatively short vase life (c. 5 days). The development of short vase life is thought to be caused by various factors, such as bacteria, respiration and ethylene production (Eason et al., 1997; Ichimura et al., 1999; Zencirkiran, 2002).

To preserve the best quality of cut- flowers after harvest and to make resistant to fluctuations in environmental conditions, treatment with floral preservatives is recommended (Nowak and Rudnicki, 1990). The addition of many substances to vase solutions promotes the development of young flower buds and delays the senescence of mature flowers (Eason and Webster, 1995).

On the other hand, ethylene is one of the factors involved in causing senescence and short vase life of many cut – flowers (Ichimura et al., 2002). The inhibition of ethylene action by pulse pre-treatment with STS has become an important commercial technique for improving the vase life of flowers, especially when they are to be handled in ethylene-contaminated environments such as supermarkets (Serek et al., 1995; Zencirkiran, 2005). However, Silver is a potent environmental pollutant. Therefore, it needs to be replaced by other non – toxic chemicals (Serek et al., 1995; Ichimura et al., 2002).

A volatile compound, 1-methylcyclopropene (1-MCP), is an inhibitor of ethylene action and appears to be

non-toxic chemicals. It has been reported that the vase life of various cut – flowers such as Carnation, *Matthiola*, *Consolida*, *Chrysanthemum*, *Zingiber*, *Anthrimum* and *Delphinium*, can be extended by exposure to 1-MCP (Serek et al., 1995; Sisler et al., 1996 a, b; Ichimura et al., 2002; Hassan and Gerzson, 2002; Almeida et al., 2002).

The objective of the current research was to determine the effects of two chemical treatments on the vase life of *Freesia* “Cordula” flowers.

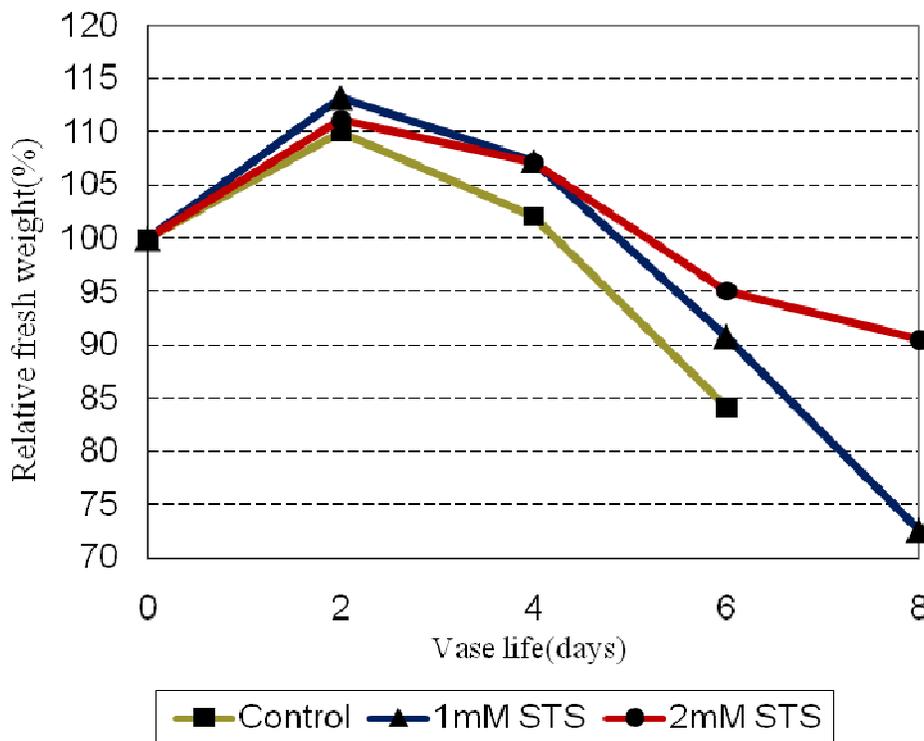
## MATERIALS AND METHODS

### Plant material

Corms of *F. refracta* “Cordula” were used for research material. Corms were kept at 30 ± 1°C in 85 - 90% relative humidity (RH) conditions for 6 weeks to overcome dormancy (Hartsema, 1962). Sprouted corms were planted in plastic house at a distance 10 × 10 cm with shoot tips at the soil surface, on November 2008. During the cultivation period, plant were subjected to natural day length conditions and plastic house temperature was controlled at 18 ± 1°C day, 12 ± 1°C night. Plants were fertilised with 15 N: 6.6 P: 12.5 K/L irrigation water, when 2 - 3 flowers had formed, until bloom as previously described (Wulster and Gianfagna, 1991). The flowers were picked in the morning, when the first coloured buds began to open (Nowak and Rudnicki, 1990), and brought into the laboratory.

### 1-MCP treatment

The stem of each cut flower was re-cut to 28 - 30 cm and placed in a 3 L jar containing water. Flowers were placed with their bases in



**Figure 1.** Effects of STS treatment on relative fresh weight in cut - *Freesia* "Cordula" flowers.

water and then in sealed glass chambers at 20°C, and 1 – MCP (2 and 4nL.L<sup>-1</sup> concentration) was injected into the jar. The flowers were exposed to 1 – MCP for 3 and 6 h (Serek et al., 1995; Sisler et al., 1996a). After the treatments, flowers were transferred to 3 L jar containing 1 L distilled water.

#### STS treatment

Flowers were pulse – treated for 2 h with 1 and 2 mM STS, prepared as described by Reid et al. (1980). After the treatments, flowers were transferred to 3 L jar containing 1 L distilled water. Control flowers were held in jars without chemical treatment.

#### Evaluation of fresh weight loss

Fresh weight loss of flowers was measured every 2 days of the vase life period for the various treatments (Waithaka et al., 2001).

#### Evaluation of vase life

Vase life of cut – flowers was assessed in a controlled environment maintained at 20 ± 1°C, 85 - 90% RH, and a 12 h day length under cool white fluorescent lighting at intensity of 15 mol m<sup>2</sup>s<sup>-1</sup>. The vase life of flowers was terminated when 30% of the florets and whole florets (spike) on a cut stem had senesced (Zencirkiran, 2002; Zencirkiran and Mengüç, 2003).

#### Statistical analysis

In all experiment, flowers were arranged using a completely

randomised design in a controlled environment vase life room. The treatment were replicated three times and each replicate consisting of five flowers. All data were analysed using Minitab version 10.1 for windows. Separation of means was done by Duncan multiple range tests at the 0.05 level.

## RESULTS AND DISCUSSION

When we investigate the range of fresh weight, it can be seen that, different applications had different effects on the values of fresh weight. The increase in fresh weight (%) occurred rapidly during the first two days of vase life in all applications. The maximum increase in the value of the fresh weight (%) was observed in the flowers that are exposed to the application of 2nL.L<sup>-1</sup> + 3 h 1-MCP (Figures 1 and 2).

Typically, cut flowers in the vase initially gain weight and then subsequently lose fresh weight (Joyce et al., 2000). From this point of view, the increase of the fresh weight (%) during the first two days of vase life was consistent with the results of Çelikel and Reid (2002) and Darras et al. (2005). In contrast to the beginning of the vase life, the fresh weight loss was observed after the two days of vase life. We conclude that fresh weight loss can be decreased with applications (STS and 1-MCP) and during the eight days of vase life, the minimum weight loss occurred in the applications of the 2 mM STS (9.46%) and 4nL.L<sup>-1</sup> + 3 h (16%) 1-MCP (Figures 1 and 2).

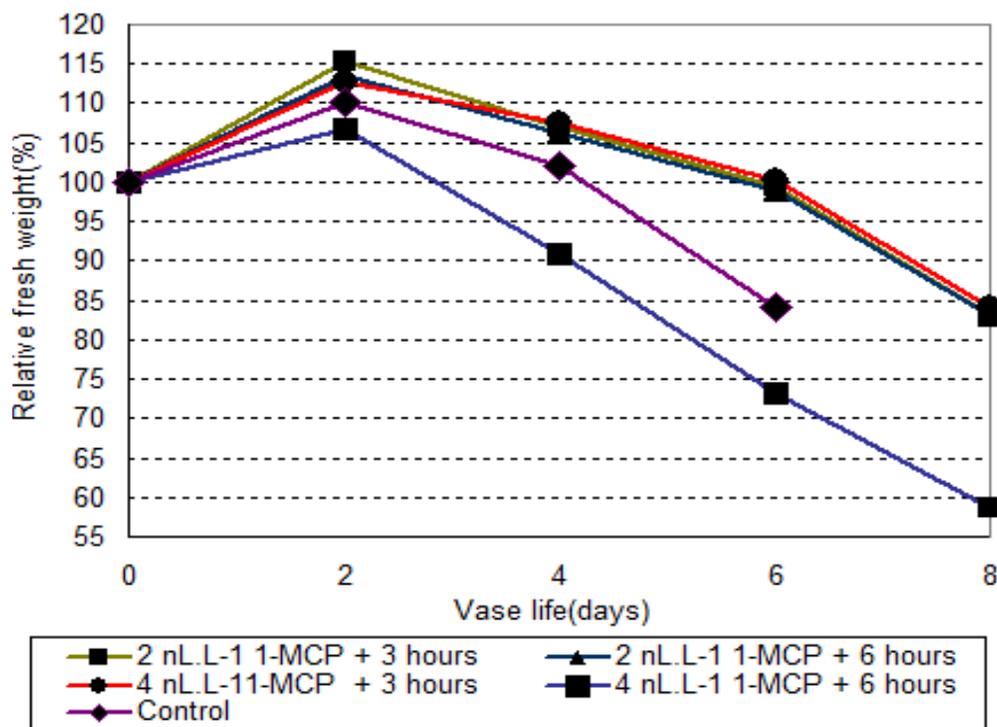


Figure 2. Effects of 1-MCP treatment on relative fresh weight in cut - *Freesia* "Cordula" flowers.

Table 1. Effects of 1-MCP and STS on the vase life of cut *Freesia* "Cordula" flowers.

Treatment	Vase life (days)	
	Floret	Spike
Control	4.66d*	6.33c*
1 mM STS	6.73c	8.11b
2 mM STS	7.06b	8.36b
2 nL.L-1 1-MCP + 3 h	7.06b	8.60b
2 nL.L-1 1-MCP + 6 h	7.40b	8.23b
4 nL.L-11-MCP + 3 h	7.60a	9.06a
4 nL.L-1 1-MCP + 6 h	7.00bc	8.15b

\*Mean separation in columns by Duncan multiple range test, P = 0.05.

As a result of these applications, the decrease in fresh weight (%) leads to increase of the vase life. This result is also in agreement with the results explained by Çelikel and Reid (2002).

Treatments with 1-MCP and STS extended the vase life of florets and spikes of cut -*Freesia* "Cordula" (Table 1). While vase life of control group florets was 4.66 days, it had increased from 2 - 2.94 days in the STS and 1-MCP treatments. The longest spike vase life was also obtained in the 1-MCP (4 nL.L<sup>-1</sup> + 3 h) treatment as 9.06 days, spike vase life of control group flowers was 6.33 days (Table 1). The researches made in the cut - flowers as *Carnation*, *Matthiola*, *Consolida*, *Anthirrinum*,

*Delphinium* by Ichimura et al. (2002), Çelikel and Reid (2002), Serek et al. (1995) and Sisler et al. (1996 a, b) reported that 1-MCP and STS treatments had positively effect on vase life. On the other hand, according to Spikman (1989), STS did not affect the ageing of blooming *Freesia* florets. However, in this research 1-MCP treatments had similar result with STS; even though they had better results (4 nL.L<sup>-1</sup> 1-MCP + 3 h) than STS.

Although STS extends the vase life of cut *Freesia* flowers, it contains silver that is a potent environmental pollutant. There has been some restriction on its commercial use (Serek and Reid, 1993; Ichimura et al., 1998). Therefore, it needs to be replaced by other

non-toxic chemical with the aim of protection of environmental pollution. On the other hand, 1-MCP treatments enhanced satisfactorily, vase life in cut-*Freesia* "Cordula" flowers when compared with control (Table 1). As a result of this research, 1-MCP can be used as alternative treatment in order to extend the vase life of cut - *Freesia* flowers as non-toxic chemical with the aim of protection of environmental pollution.

## REFERENCES

- Almeida AS, Alves RE, Paiva WO, Lima MAC, Almeida JBSA (2002). Quality and Storage of Beehive Ginger (*Zingiber spectabilis*) Following Postharvest Treatment with 1-MCP. *Proc. Am. Soc. Trop. Hort.* 46: 110-111.
- Çelikel FG, Reid MS (2002). Postharvest Handling of Stock (*Matthiola incana*). *HortScience*, 37: 144-147.
- Darras AI, Terry LA, Joyce DC (2005). Methyl jasmonate Vapour Treatment Suppresses Speckling Caused by *Botrytis cinerea* on *Freesia hybrida* L. Flowers. *Postharvest Biol. Technol.*, 38: 175-182.
- Eason JR, Webster D (1995). Development and Senescence of *Sandersonia aurantica* (Hook.) Flowers. *Scientia Hort.* 63: 113-121.
- Eason JR, Vre De LD, Somerfield SD, Heyes JA (1997). Physiological Change Associated with *Sandersonia aurantica* Flower Senescence in Response to Sugar. *Postharvest Biol. Technol.*, 12: 43-50.
- Hartesema OM (1962). Temperature Treatments of *Freesia* Tubers. *Proceeding 16<sup>th</sup> International Congres. Gembloux, Belgium* 5: 298-304.
- Hassan FAS, Gerzson L (2002). Effects of 1-MCP (1-methylcyclopropene) on the Vase Life of Chrysanthemum and Carnation Cut Flowers. *Int. J. Hort. Sci.* 8: 29-32.
- Ichimura K, Shimamura M, Hisamatsu T (1998). Role of Ethylene in Senescence of Cut *Eustoma* Flowers. *Postharvest Biol. Technol.*, 14(2): 193-198.
- Ichimura K, Kojima K, Goto R (1999). Effects of Temperature, 8-Hydroxyquinoline Sulphate and Sucrose on the Vase Life of Cut Rose Flowers. *Postharvest Biol. Technol.*, 15: 33-40.
- Ichimura K, Shimizu H, Hiraya T (2002). Effect of 1-methylcyclopropene (1-MCP) on the Vase Life of Cut Carnation, Delphinium and Sweet Pea Flowers. *Bull. Natl. Inst. Flor. Sci.*, 2: 1- 8.
- Joyce DC, Meara SA, Hatherington ES, Jones P (2000). Effect of Cold Storage on Cut *Grevillea* 'Sylvia' Inflorescences. *Postharvest Biol. Technol.*, 18(1): 49-56.
- Nowak J, Rudnicki RM (1990). *Postharvest Handling and Storage of Cut Flower, Florist Greens and Potted Plants*. Timber Press, Inc., p. 210.
- Reid MS, Paul JL, Farhoomand MB, Kofranek AM, Staby GL (1980). Pulse Treatments with the Silver thiosulphate Complex Extend the Vase Life of Cut Carnations. *J. Am. Soc. Hort. Sci.*, 105: 25-27.
- Serek M, Reid MS (1993). Anti-ethylene Treatments for Potted Flowering Plants-Relative Efficacy of Inhibitors of Ethylene Action and Biosynthesis. *HortScience*. 28: 1180-1181.
- Serek M, Sisler EC, Reid MS (1995). Effects of 1-MCP on the Vase Life and Ethylene Response of Cut Flowers. *Plant Growth Regulation*, 16: 93-97.
- Sisler EC, Dupille E, Serek M (1996a). Effect of 1-methylcyclopropene and methylenecyclopropane on Ethylene Binding and Ethylene Action Cut Carnations. *Plant Growth Regulation*, 18: 79-86.
- Sisler EC, Serek M, Dupille E (1996 b). Comparison of Cyclopropene, 1-methylcyclopropene, and 3, 3- dimethylcyclopropene as Ethylene Antagonists in Plants. *Plant Growth Regulation*, 18: 169-174.
- Spikman G (1989). Development and Ethylene Production of Buds and Florets of Cut *Freesia* Inflorescences as Influenced by Silver thiosulphate, Aminoethoxyvinylglycine and Sucrose. *Scientia Horticulturae*, 39(1): 73-81.
- Waithaka K, Reid MS, Dodge LL (2001). Cold Storage and Flower Keeping Quality of Cut *Tuberosa* (*Polianthes tuberosa* L.). *J. Hort. Sci. Biotech.*, 76(3): 271-275.
- Wulster GJ, Gianfagna TJ (1991). *Freesia hybrida* Respond to Ancyimidol, Cold Storage of Corms and Greenhouse Temperatures. *Hort Sci.* 26(10): 1276-1278.
- Zencirkiran M (2002). Cold Storage of *Freesia refracta* "Cordula". *New Zealand J. Crop Hort. Sci.*, 30: 171-174.
- Zencirkiran M, Mengüç A (2003). Cold Storage of *Alstroemeria pelegrina* "Ostara". *New Zealand J. Crop Hort. Sci.*, 31: 255-259.
- Zencirkiran M (2005). Effects of Sucrose and Silver thiosulphate Pulsing on Stem – base Cracking and Vase Life in *Leucojum aestivum* Flowers. *J. Hort. Sci. Biotech.*, 80(3): 332-334.